

W.B.

STRESS AND RECOVERY OF BENTHIC
INFAUNA FROM OCEAN DISCHARGE
OFF ORANGE COUNTY, CALIFORNIA

SOUTHERN CALIFORNIA COASTAL WATER
RESEARCH PROJECT

November 1974

Effects of Diversion to Deepwater off Orange County, California

On 31 March 1971, a discharge of 133 MGD of primary treated wastewater was diverted from a one-mile outfall to a five-mile long outfall off the Orange County Coast by the County Sanitation Districts of Orange County. Biological data taken by Marine Biological Consultants and by Gary Smith (S.I.O.) and sediment metals data taken by D. Young and J. Galloway suggested that after 17 years of continuous discharge (from 27 to 133 MGD) the inshore benthic environment at the nearshore outfall recovered within a year. In addition, offshore data suggested that no major effects in the benthic environment were found during the first year of discharge out the long outfall.

We propose to re-examine these data, complete analyses of remaining un-analyzed samples (especially from Smith) and initiate sampling at sites not now being surveyed. Such a program should provide urgently needed data on both the reversibility of effects from nearshore discharge of a large volume of primary effluent and provide insight into the kinds of environmental responses (or lack of them) engendered by a modern, well-designed sub-thermocline diffuser system.

I. Work Plan

A. Previous data (cited above) will be collated and reviewed for its adequacy to describe the significance of environmental changes which may have occurred at the old outfall site as a result of effluent diversion. This will include pre- and post-

discharge switch quantitative information on fish abundance, diversity and disease incidences, benthic infaunal species abundances and diversities and on sediment metals (copper and lead) and chlorinated hydrocarbons.

B. Completion of Benthic Infaunal Samples

Between July 1970 and September 1972, Dr. Gary Smith, Scripps Institution of Oceanography, collected a total of 342 benthic samples at six stations near the outfalls and at two control stations. The outfall stations involved one transect at each outfall with stations at 0.25, 0.5 and 1.0 mile downcoast of the outfalls. Sampling was approximately bimonthly with six samplings before and six after the discharge switch. Two replicates from each station were analyzed for total sulphides ($\text{mgS}^{\equiv}/\text{g}$ sediment dry weight) in the upper 1 cm layer and for total organic carbon (% organic carbon, dry weight basis). However, only the 0.25 mile stations were completely analyzed for benthic infaunal species. The remaining samples (288), now at the Allan Hancock Foundation, are available for further analysis to confirm or supplement Dr. Smith's conclusions. We propose to initially analyze two samples from each of the 0.5 and 1.0 mile stations (4 stations) for six of the 12 sampling periods. This amounts to 48 samples at an approximate cost of \$60/sample (total \$3,000) by a qualified outside laboratory (U.S.C. or M.B.C.).

These data will be examined for changes in diversity and abundance and their relation to the available physical data

and to the already analyzed 0.25 mile infaunal data.

C. New SCCWRP and ORCOSAN Sampling and Analysis

Until September 1974, the County Sanitation Districts had no benthic sampling program other than special sampling for metals and pesticides. Thus, a nearly two-year gap exists in our knowledge between Smith's last sampling and the new program instituted at the new outfall site by ORCOSAN.

ORCOSAN sampling will continue quarterly as of September 1974 and will provide information on benthic infaunal and chemistry changes from that time on at the new outfall site. However, the sampling will not provide additional data on recovery at the one-mile outfall nor an assessment of the benthic fauna and chemistry over a larger region surrounding both outfalls.

Such information will be necessary to adequately assess past and future effects. Therefore, we propose to conduct one major, but complementary, benthic sampling program at sites about ORCOSAN's present grid of stations and to include three of Dr. Smith's previous sites at the one-mile outfall. This program is described below.

In addition, we will continue to participate in the continuing quarterly otter trawl program (established in 1969).

STATIONS

Sampling will initially be conducted at 24 stations offshore of Huntington Beach, Newport Beach and Corona del Mar. The stations selected surround the ORCOSAN stations and extend their benthic sampling pattern inshore to 10 fm (including Gary

Smith's three inshore stations labeled A, B, and C), upcoast to a distance of 5 miles from the outfalls, offshore to depths of 100 fm and downcoast to Corona del Mar. These stations are labeled in the accompanying tables and figures as S-1 through S-21 (SCCWRP stations) and B-1 through B-17 including Smith's offshore stations labeled D, E, and F (Orange Co. San Stations).

SAMPLING

Benthic sampling will be done using the Orange Co. San. Petersen grab aboard Mr. Munson's vessel or one with similar capabilities and navigational equipment (fathometer accurate to a depth of 100 fm, Loran, and radar).

Three replicate samples will be taken at each station. Each sample will be examined for sediment color and consistency, then placed in a bucket to measure wet volume. Each sample will then be washed separately through 0.5 mm mesh screen. Retained organisms will be preserved in buffered formalin and later transferred to isopropyl alcohol for sorting, identification, weighing and counting.

One or two additional grab samples will be taken at each station to obtain sediment samples for:

- a. particle size distribution - 100 gm
- b. total carbon determination - 100 gm
- c. C.O.D. determination - 100 gm
- d. metals determination - 100 gm (chromium, copper, zinc, selenium and antimony)
- e. CHC determination - 100 gm (DDT, DDE, Dieldrin and PCBs)

At each station, the sequence of taking biological and physical samples will be randomized; ie., the physical samples will be taken first at some stations, second at others, third at others, etc. This should reduce bias introduced by ship drift in the comparison between biological and physical data.

ANALYSIS OF SAMPLES

Biological Samples. The biological samples from the initial survey will be analyzed by a qualified laboratory under SCCWRP direction, and a reference collection of specimens will be maintained for future reference. A duplicate collection will be given to the Orange County Sanitation District.

Species abundance lists from each replicate and station will be coded, keypunched and stored for calculation of diversity indices (Gleason, Simpson, Shannon-Weaver, Brillouin, standard deviation and scaled versions of these indices) and similarity indices (Bray-Curtis). The latter will be used to conduct a site-group analysis which will also include data taken by ORCOSAN.

These procedures will (a) produce a series of charts showing site-groupings and their distribution and geographical diversity gradients and (b) data for regression against physical-chemical parameters (below).

Sediment Particle Size Analysis will be done from one sample at each station by the Geology Dept. laboratory at USC (Dr. D. Gorsline). Data on median and mean distributions (mm), sorting coefficients and percent gravel, sand, silt and clay will be obtained.

Total Carbon will be done from one sample from each station by the Geology Department laboratory at USC or a similar suitable laboratory.

To obtain compatible information at existing Orange County stations, we will request samples from ORCOSAN and submit them with those from our survey.

C.O.D. will be measured by submitting samples to the ORCOSAN laboratory so results will be comparable.

Metals (chromium, copper, zinc, selenium and antimony) will be analyzed by absorption spectroscopy at SCCWRP.

Chlorinated Hydrocarbons will be analyzed by gas liquid chromatography at SCCWRP.

These physical and chemical data (including sediment color and consistency) will be used to prepare geographical charts showing gradients in these parameters. These data will then be examined for possible gradients associated with the discharge sites and with other important factors such as depth. Analyses will then be conducted to determine if correlations exist between these physical parameters and the biological data such as diversity, species richness, site-groupings, abundance and dominant species.

Following this initial survey, we will examine the data to determine sites and sampling frequencies for future surveys which will adequately assess chronic effects of the discharges and separate such effects from seasonal fluctuation in species and physical parameters.

SCCWRP PROPOSED STATIONS

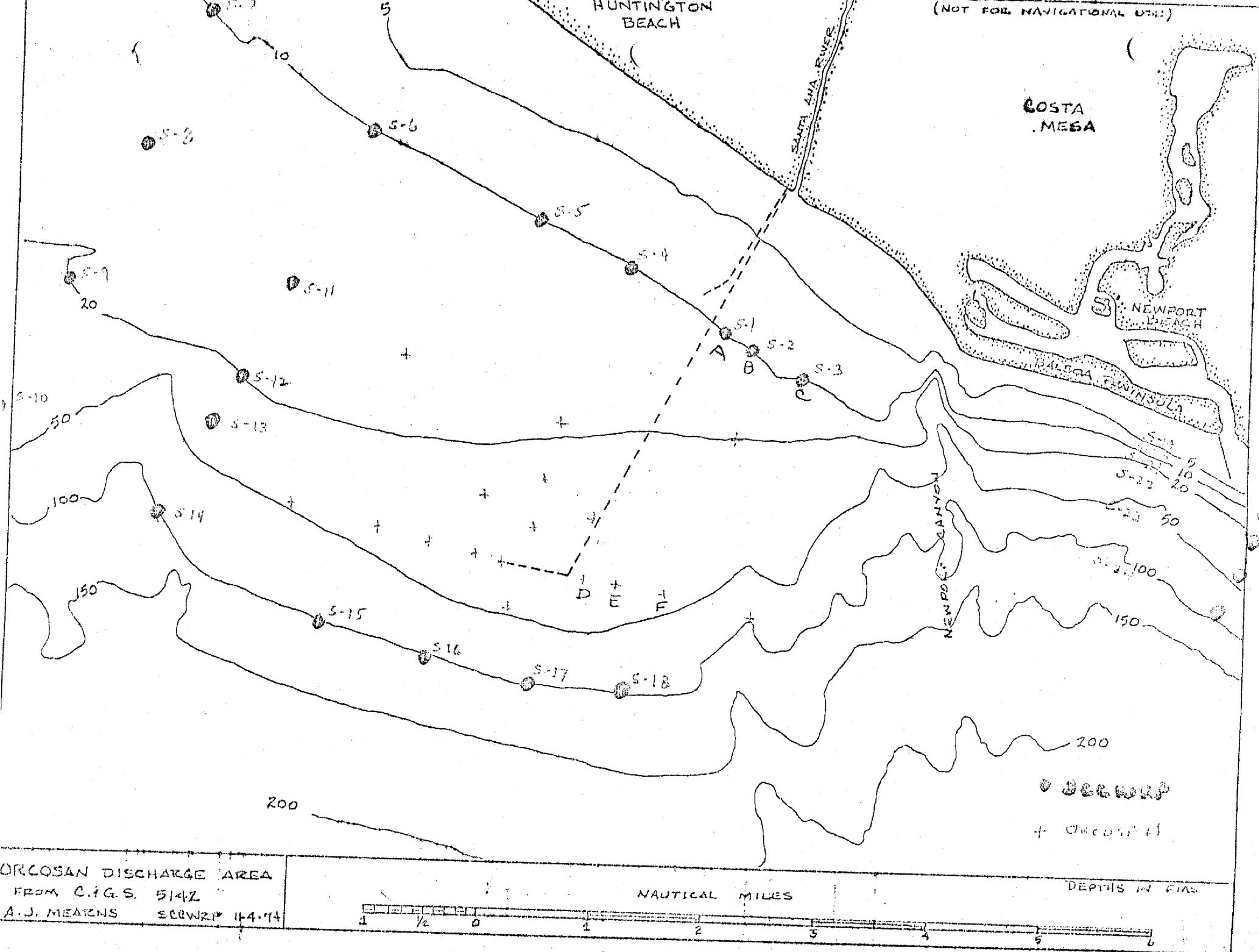
San Pedro Grid

<u>Station</u>	<u>Depth (fm)</u>	<u>Location (approximate only)</u>				
S-1	10	1/4 mi SE	of 1-mi outfall	"	"	"
S-2	10	1/2 mi SE	"	"	"	"
S-3	10	1.0 mi SE	"	"	"	"
S-4	10	0.85 mi NW	"	"	"	"
S-5	10	1.75 mi NW	"	"	"	"
S-6	10	3.2 mi NW	"	"	"	"
S-7	10	5.1 mi NW	"	"	"	"
S-8	15	4.9 mi NW	"	5-mi diffuser end	"	"
S-9	20	4.7 mi WNW	"	"	"	"
S-10	31	4.9 mi W	"	"	"	"
S-11	15	3.1 mi NW	"	"	"	"
S-12	20	3.0 mi WNW	"	"	"	"
S-13	31	3.0 mi SWN	"	"	"	"
S-14	100	3.2 mi W	"	"	"	"
S-15	100	1.8 mi WSW	"	"	"	"
S-16	100	1.2 mi SSW	"	"	"	"
S-17	100	1.0 mi S	"	"	"	"
S-18	100	1.5 mi SSE	"	"	"	"

Corona del Mar Control Transect

S-19	8	0.4 mi SE	of Newport Jetty	"	"	"
S-20	15	0.5 mi SSE	"	"	"	"
S-21	20	0.6 mi SSE	"	"	"	"
S-22	31	0.8 mi S	"	"	"	"
S-23	50	1.1 mi S	"	"	"	"
S-24	100	1.4 mi S	"	"	"	"

(NOT FOR NAVIGATIONAL USE)

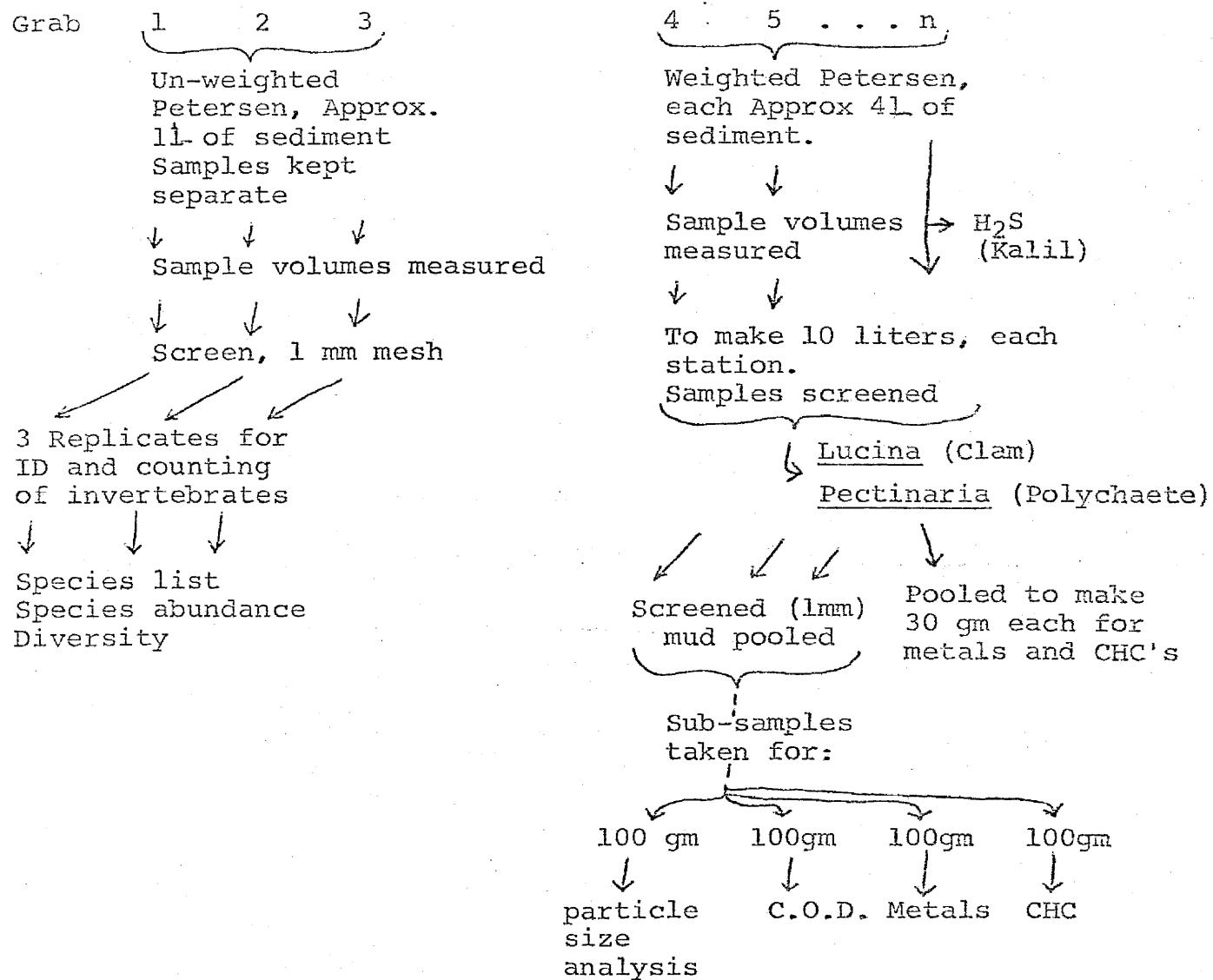


APPENDIX II

INFORMATION ON PRESENT BENTHIC
MONITORING PROGRAMS IN THE
ORANGE COUNTY OUTFALL AREA

FLOW DIAGRAM FOR BOTTOM SAMPLING
AT ORANGE COUNTY SANITATION
SHIPBOARD FATE OF SAMPLE

Five or More Samples are Taken at Each Station



MEMORANDUM

October 30, 1974

To: Willard Bascom, Charles Greene, Jack Word, Dave Young

From: Alan Mearns

Re: Present Sampling Program (Benthic) at Orange County

The County Sanitation Districts of Orange County initiated their new quarterly and weekly ocean sampling programs in September and October 1974, as specified by the Santa Ana Regional Water Quality Control Board (Order No. 74-11, pp. 22-33).

I talked with the laboratory staff at Orange County Sanitation to obtain specific information on progress and procedures used in their sediment sampling program.

Seventeen bottom stations are taken each quarter of the year (August complete; November and January planned) using a Petersen grab sampler with 0.1m² maximum area. Approximately 4 stations are completed in one ship day requiring four or more ship days to complete a survey. These are done on Tuesdays and Thursdays, in the morning hours to avoid effects of afternoon wind and chop on vessel and gear performance.

The vessel used is Fred Munson's out of Newport.

At each station, at least five grab samples are taken as described below and as shown in the accompanying flow diagram:

1. Three separate replicate samples are taken using the Petersen without addition of lead weights (obtain small samples). Each sample volume is measured in a calibrated bucket, then the sample is washed thru 1 mm screen. Retained organisms are preserved in labeled jars for sorting, identification and counting.
2. Two or more samples are taken using the Petersen fitted with lead weights (to obtain large sediment volumes). Each sample volume is measured and sampling is continued until 10 liters of sediment are obtained. Hydrogen sulfide is measured by Kalil. After each volume measurement, the samples are washed thru 1 mm screen and both the sediment and screened fractions are retained. The screened fractions are sorted to obtain 30 gm of the clam Lucina and 30 gm of the polychaete Pectinaria for analysis of metals and chlorinated hydrocarbons.

Memorandum
October 30, 1974
To: WB, CG, JW, DY
From: AM
Re: Present Sampling Program at OC

The sediments from the washings are subsampled as follows:

100 gm for particle size analysis.
100 gm for C.O.D. (which replaces total carbon as specified by SARWQCB).
100 gm for metals.
100 gm for chlorinated hydrocarbons.

This completes the work at one station.

All analyses are performed at Orange County Sanitation. The physical/chemical determinations are made using methods cited in the following publications:

Particle Size Analysis -

Water Quality Control Plan, Ocean Waters of California SWRCB 1972 and Methods for Chemical Analysis of Waters and Wastes, EPA 1971

Metals -

Great Lakes Region: Chemical Laboratory Manual for Bottom Sediments. Compiled by the Great Lakes Regional Committee on Analytical Methods. EPA. December 1969. (Including H₂O₂, Nitric acid, HCl and Aqua Regia methods).

Pesticides -

Analysis of Pesticide Residues in Human and Environmental Samples. EPA. Revised November 1972.

C.O.D. (Chemical Oxygen Demand) -

Standard Methods for Wastewater Analysis.

Benthic Infauna -

References and methods recommended by SCCWRP (J. Word)
Biologists training for ID work include:

Ida Duesberg
Ann Martin
Fred O'Brien
Mike Hines

BOTTOM SAMPLING STATIONS
ORANGE COUNTY SANITATION DISTRICTS
ADOPTED JUNE 7, 1974

<u>Station Sediments</u>	<u>Depth fm</u>	<u>Approximate Distance and Bearing from Diffuser end</u>
B-0	32	End point of Diffuser
B-1	30	0.31 mi @ 265°
B-2	28	0.31 mi @ 015°
B-3	45	0.31 mi @ 195°
B-4	31	0.62 mi @ 105°
B-5	31	0.93 mi @ 265°
B-6	25	0.93 mi @ 310°
B-7	23	0.93 mi @ 015°
B-8	28	0.93 mi @ 060°
B-9	31	0.93 mi @ 105°
B-10	30	1.55 mi @ 265°
B-11	18	1.55 mi @ 015°
B-12	32	1.55 mi @ 105°
B-13	45	2.48 mi @ 265°
B-14	15	2.48 mi @ 310°
B-15	20	2.48 mi @ 060°
B-16	80	2.48 mi @ 105°

Bottom Sampling Stations (cont'd)

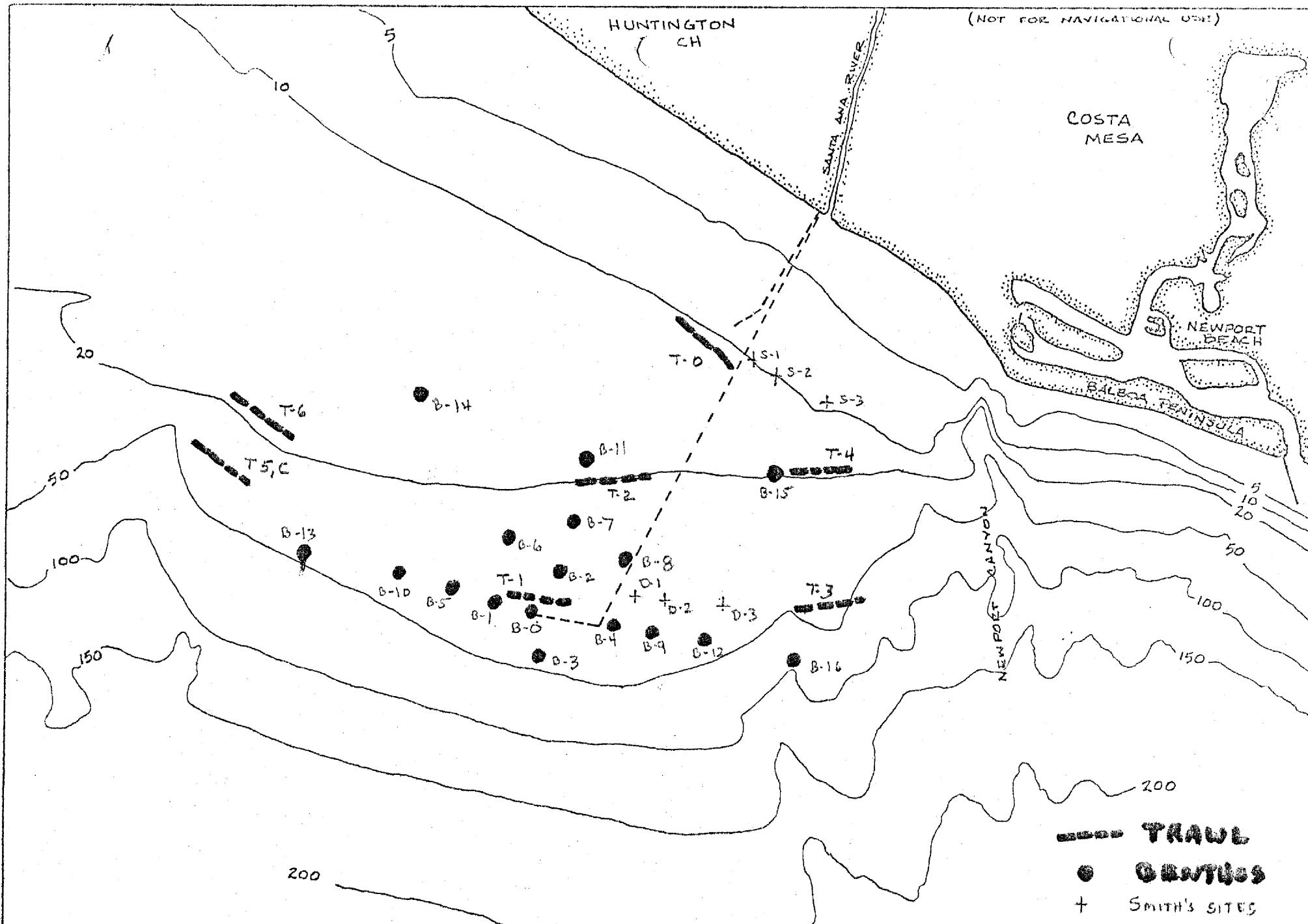
<u>Station Trawls</u>	<u>Depth fm</u>	<u>Approximate Distance and Bearing from Diffuser end</u>
T-0	10	3.1 mi @ 035° (old Diffuser end)
T-1	31	New Diffuser end (0.12 mi @ 020°)
T-2	20	1.48 mi @ 030°
T-3	35	2.79 mi @ 090°
T-4	20	3.1 mi @ 060°
T-5	23	3.41 mi @ 295°
T-6	20	3.41 mi @ 305°

BENTHIC STATIONS SAMPLED

BY GARY SMITH (S.I.O.)

SEPTEMBER 1970 to SEPTEMBER 1972

<u>Smith's Stations</u>	<u>Depth fm</u>	<u>Approximate Location</u>
A	10	1/4 mi from outfall
B	10	1/2 mi from outfall
C	10	1 mi from outfall
D	31	1/4 mi from outfall
E	31	1/2 mi from outfall
F	31	1 mi from outfall



ORCOSAN DISCHARGE AREA
FROM C.I.G.S. 5142
A. J. MEARNS SCCWRP 11-4-74

NAUTICAL MILES

0 1 1½ 2 3 4 5 6

SCCWRP
CHLORINATED HYDROCARBON ANALYSIS

SAMPLE AND LOCATION Orange Co. Sediments

DESCRIPTION

DATE 11/16/75 UNITS REPORTED ppb dry weight

SAMPLE NUMBER	SE30	NC30	NJ30	SC30	SB30
SAMPLE DESCRIPTION					
CONSTITUENT					
op'-DDE					
pp'-DDE	10.2	16.7	18.9	8.0	24.9
op'-DDD					1.7
pp'-DDD	1.5	2.7			8.5
op'-DDT			0.3		
pp'-DDT	0.6		1.5		
Heptachlor Epoxide					
Aldrin					
Dieldrin					
TOTAL DDT					
TOTAL PESTICIDE (DDT)	12.3	19.4	20.7	8.6	35.1
Arochlor 1242 *	100.8	52.6	166.8	68.6	121.4
Arochlor 1254 *	27.2	26.2	68.5	22.8	61.9
Arochlor 1260					
TOTAL PCB	128.0	78.8	235.3	91.4	183.3
TOTAL CHLORINATED HYDROCARBONS	140.3	98.2	256.0	99.4	218.4

SCCWRP 4-2

* PCB's based on "saponified" sample

UNIVERSITY OF SOUTHERN CALIFORNIA
UNIVERSITY PARK
LOS ANGELES, CALIFORNIA 90007

DEPARTMENT OF GEOLOGICAL SCIENCES
TELEPHONE: (213) 746-2717

December 18, 1975

BRIEF ANALYSIS
SCCWRP SAMPLE SET

Samples are all silty sands with major modes in fine sand and coarse silt. Clay fractions are negligible in all samples and in all instances less than 5% of total sediment weight. Since the amount of clay was very small (clay in the particle size sense) the samples were analysed in terms of regular Wentworth size grades down to the 4 micron diameter (boundary between silt and clay) and then the clay-sized particles were lumped. This will influence the tail of the size distribution as plotted on log probability paper and give the appearance of a third component distribution below the silt range. In actual fact the silt and clay is all one normal distribution and is a matrix for a single or double mode sand size distribution. Most samples show a poorly sorted, normal medium sand with fine sand-silt matrix. A few samples, as for example in SB 30, a coarse mode also appears. This is probably either an artifact of compositing several discrete sedimentation units in a grab sample or may represent shell or gravel clasts in a sand mass.

Soluble organic fraction is everywhere less than 6% and has a mode at about 1.5% by weight. This is probably also a separate addition to the basic sediment. All samples were treated with a 50:50 acetone/distilled water washing solution and decanted and washed with distilled water. Dry residue are after acetone leaching therefore.

General size parameters show the influence of the dominant fine sand-silt component and all have mean diameters in the range from 30-100 microns with changes due to the amount of the coarse mode contributed. Only one sample (7C) falls below the range considering normal error of analysis. In this sample the median is in the fine sand and silt range and the weighted mean is thus perhaps not really that far off the group.

Mean diameter, standard deviation and skewness are relatively insensitive to the small tail contribution and thus lumping the clay grade has little or no effect on those for comparison purposes. Kurtosis is markedly changed by the degree of subdivision in the sizing process and should be used with caution in inter-group comparisons.

SCCWRP Sediment samples: Moment measures. (Mean diameter ϕ , mean diameter mm, sorting coefficient, skewness, kurtosis)

NC-50 NB-50

SC-50 SE-50

MEASURE	NJ50	NB50	SB50	NE30	SB30	SC30	SC8	SE50	SEE50
M.D. ϕ	4.32	3.95	3.93	3.89	3.43	3.28	2.72	3.44	2.96
M.D. mm	0.0499	0.0644	0.0652	0.0671	0.0922	0.1024	0.1508	0.0917	0.1283
So.	1.32	1.10	1.20	1.11	1.26	1.75	0.77	1.09	1.21
Sk.	2.55	3.51	3.22	3.49	2.08	1.14	2.45	2.88	1.37
K.	6.93	13.39	11.24	13.74	8.14	2.23	15.68	13.03	3.69
									SB-50
	NC 25	SF 30	NC 30	NB 20	NN 25	NE 20	NJ 25	SB 25	SC 50
M.D. ϕ	4.22	3.17	3.84	4.92	3.85	4.08	3.99	4.34	3.83
M.D. mm	0.0534	0.1107	0.0694	0.0328	0.0690	0.0587	0.0627	0.0492	0.0700
So.	1.29	1.48	0.95	1.77	0.88	1.31	1.27	1.40	1.32
Sk.	2.43	2.01	3.22	1.38	2.22	2.41	2.72	2.37	2.43
K.	7.31	6.79	11.79	1.28	5.04	7.28	8.76	5.51	7.36
									NE 50
	SE 30	NJ 30	NB 25	NE 8	SC 20	SE 25	SC 25	NE-50	SE 8
M.D. ϕ	3.57	4.03	4.04	3.84	3.84	4.34	4.51	3.98	2.84
M.D. mm	0.0841	0.0610	0.0606	0.0695	0.0697	0.0491	0.0436	0.0632	0.1390
So.	1.30	1.16	1.26	0.91	1.24	1.40	1.55	1.18	0.71
Sk.	2.34	3.18	2.61	2.69	3.26	2.33	1.79	3.41	3.95
K.	8.16	11.28	8.39	15.19	10.82	5.45	3.61	12.26	28.45
							100		
	NB 30	SC 100 #1	NB 100	NN 30	NB 8	NF 180	SB 8	NE 25	1255
M.D. ϕ	3.86	4.75	4.70	4.00	2.50	4.56	3.75	4.02	1.57
M.D. mm	0.0684	0.0370	0.0384	0.0623	0.1762	0.0423	0.0738	0.0613	0.3360
So.	1.15	1.53	1.57	1.10	0.50	1.59	1.06	1.32	0.80
Sk.	3.34	1.90	1.89	3.13	-0.69	1.62	1.51	2.47	2.74
K.	12.45	3.20	3.15	10.80	3.66	3.18	6.32	8.20	20.58
	10C	7C	4D	5D	5C				
M.D. ϕ	3.88	5.69	4.35	1.95	5.25				
M.D. mm	0.0678	0.0193	0.0489	0.2582	0.0261				
So.	1.44	1.89	1.75	1.75	2.24				
Sk.	2.27	0.94	1.52	2.05	0.22				
K.	7.04	-0.32	2.77	4.75	0.04				

Size Grade	NJ 50 DB 50	NC 50 DB 50	NB 50 DB 50	NE 30	SB 30	SC 30	SC 8	SC 50 SE 50	SE 50 SEE 50
-1 - 0	0	0	0	0	0.23	0.57	0	0	0
0 - 1	0	0	0	0	0.58	5.22	0	0.24	2.94
1 - 2	0	0	0	0.13	4.67	13.64	9.59	0.95	10.28
2 - 3	0.74	0	3.28	2.28	26.02	29.03	63.73	28.12	47.03
3 - 4	49.60	74.94	73.08	74.73	53.08	28.74	23.50	57.90	27.92
4 - 5	37.86	17.00	15.54	16.45	9.27	6.83	2.02	8.36	6.89
5 - 6	3.57	2.07	2.42	1.42	1.86	9.94	0.49	1.22	1.28
6 - 7	2.77	1.76	1.66	1.70	1.55	2.23	0.25	1.11	1.30
7 - 8	1.53	1.17	1.20	0.96	1.13	1.42	0.21	0.69	2.31
8 - 10	3.93	2.26	2.83	2.33	1.61	2.30	0.20	1.41	0
									SB 50
	NC 25	SF 30	NC 30	NB 20	NN 25	NR 20	NE 25	SB 25	SE 50
-1 - 0	0	0.44	0	0	0	0	0	0	0
0 - 1	0.24	1.03	0	0	0	0	0.19	0	0
1 - 2	0.36	13.24	0	0.80	0	1.60	0.77	0	0.26
2 - 3	1.78	35.31	1.81	1.60	3.24	2.80	2.51	0	17.85
3 - 4	51.50	33.84	78.02	29.67	73.58	59.10	69.84	55.61	54.96
4 - 5	35.57	10.32	13.46	40.23	14.93	26.13	15.94	29.38	17.59
5 - 6	3.07	1.84	1.63	7.20	2.05	3.03	4.51	4.95	2.97
6 - 7	2.66	1.10	1.73	7.42	4.94	2.84	1.93	3.78	2.37
7 - 8	1.49	0.29	2.71	4.69	1.26	1.39	1.31	2.04	1.31
8 - 10	3.34	2.60	0.64	8.38	0	3.11	2.99	4.25	2.69
								NE 50	
	SE 30	NJ 30	NB 25	NE 8	SC 20	SE 25	SC 25	NE 50	SE 8
-1 - 0	0	0	0	0	0	0	0.15	0	0
0 - 1	0.48	0	0.23	0.22	0	0	0.23	0	0
1 - 2	1.91	0	0.91	1.12	0.48	0	0.38	0	1.12
2 - 3	27.07	1.28	1.70	3.13	2.39	0.83	1.55	0.39	69.40
3 - 4	50.40	66.65	65.21	65.33	86.86	53.32	41.99	74.83	26.30
4 - 5	13.23	24.27	22.27	25.29	0	30.84	37.40	18.06	1.91
5 - 6	2.05	2.38	2.90	2.85	3.26	5.07	5.53	2.32	0.66
6 - 7	1.68	1.54	2.40	0.73	2.77	3.74	4.81	1.48	0.12
7 - 8	1.00	1.17	1.52	0.28	1.50	1.94	2.56	0.93	0.13
8 - 10	2.18	2.71	2.87	1.06	2.78	4.26	5.40	2.96	0.36

Size Grade	NB 30	SC 100 ^{#1}	NB 100	NN 30	NB 8	NF 100 ¹⁰⁰	SB 8	NE 25	I255
-1 - 0	0	0	0	0	0	0.21	0	0.07	1.11
0 - 1	0	0	0	0	1.55	0.21	0	0.59	12.73
1 - 2	0	0	0	0	8.06	0.83	0.87	0.66	70.30
2 - 3	4.67	0	0.79	0.13	70.76	2.48	22.15	3.28	12.18
3 - 4	74.79	32.35	35.61	71.20	11.63	37.75	38.42	62.69	2.44
4 - 5	13.37	45.49	42.05	20.44	0	37.85	32.51	23.82	0.73
5 - 6	1.80	8.12	8.24	2.35	0	7.92	3.85	2.40	0.12
6 - 7	1.77	4.59	3.87	1.75	0	4.47	0.90	1.99	0.15
7 - 8	1.15	3.18	2.95	2.40	0	2.57	0.46	1.17	0.10
8 - 10	2.45	6.18	6.49	1.73	0	5.72	0.85	3.34	0.15

10 C 7C 4D 5D 5C

-1 - 0	0.44	0	0.77	0.21	0
0 - 1	0.44	0	0.31	28.13	4.66
1 - 2	1.75	0	0.77	45.14	4.35
2 - 3	6.57	0	4.13	8.19	0.62
3 - 4	69.20	12.73	53.75	5.46	9.33
- 5	12.29	39.97	21.43	6.34	38.00
5 - 6	1.87	14.05	5.09	2.34	13.31
6 - 7	1.89	10.67	4.27	1.63	9.41
7 - 8	1.86	8.24	3.23	1.08	7.76
8 - 10	3.69	14.34	6.27	1.47	12.55

SCCWRP Sediment Samples: Sample weights before treatment and weight % soluble organics.

SAMPLE #	DRY WT. UNTREAT.	% SOL. ORG.						
NJ 50	21.0954	1.40						
NCW 50	18.7836	1.57						
NB 50	31.1816	1.21						
NE 30	25.7923	1.34						
SB 30	21.4620	1.42						
SC 30	35.8990	1.24						
SC 8	21.7449	0.70						
SCSE 50	21.4989	1.20						
SE SEC 50	33.3549	0.89						
NC 25	20.7838	1.67						
SF 30	34.2971	1.06						
NC 30	25.8372	1.20						
NB 20	18.2012	2.14						
NN 25	26.5510	1.05						
NE 20	25.5939	1.40						
NE 25	34.8760	1.34						
SB 25	24.3737	1.37						
SB SC 50	24.2930	1.24						
E 30	28.7722	1.06						
NJ 30	30.9657	1.17						
NB 25	24.2817	1.63						
NE 8	30.0621	3.40						
SC 20	24.8766	1.13						
SE 25	29.0587	1.56						
SC 25	33.5974	1.12						
NE NC 50	26.8258	1.12						
SE 8	25.4179	1.47						
NB 30	30.1006	1.29						
SC 100 #1	30.1297	1.31						
NB 100	32.4943	1.27						
NN 30	33.6843	1.13						
NB 8	28.9600	1.17						
NF 188 ¹⁰⁰	24.9864	1.35						
SB 8	30.5285	0.97						
NE 25	36.8965	1.33						
1255	38.6288	0.75						
10 C	24.1707	1.54						
7C	7.3530	6.33	T. H.					
4 D	24.7120	1.40						
5 D	41.0553	1.01						
5 C	7.8038	4.77						

CNIDARIA
HYDROZOA

Hydriida, unid. 91101

Monobrachium parasitum Mereschkovsky 91113

ANTHOZOA

Anthozoa, unid. 91665

Ceriantharia, unid. 92130

Edwardsiella californica McMurrich 92075

Filigella mitsukurii Kinoshita 91875

?Renilla kollikeri Pfeffer 91950

Pennatulacea, unid. 92017

Virgulariidae, unid. 91986

PLATYHELMINTHES

Platyhelminthes, unid. 92500

NEMERTEA

Nemertea, unid. 93000

NEMATODA

Nematoda, unid. 94500

ENTOPROCTA

Barentsia sp. 93985

ANNELIDA

POLYCHAETA

ERRANTIA

Anaitides sp. 21210

Chloeia pinnata Moore 21012

Diopatra ornata Moore 23011

Drilonereis falcata Moore 23331

D. mexicana Fauchald 23335

Drilonereis sp. 23330

Eteone californica Hartman 21243

E. dilatae Hartman 21244

Eulalia quadrioculata Moore 21254

Eulalia sp. 21250

Eunice americana Hartman 23111

Eusigillion spinosum Hartman 23111

Exogone gemmifera Pagenstecher 22181

E. lourei Berkeley & Berkeley 22182

E. uniformis Hartman 22183

Glycera americana Leidy 22811

G. branchiopoda Moore 22812

G. capitata Oersted 22813

G. convoluta Keferstein 22814

G. oxycephala Ehlers 22817

G. robusta Ehlers 22818

Glycera sp. 22810

Glyceridae, unid. 22800

Glycinde polygnatha Hartman 22912

Glycinde sp. 22910

Goniada brunnea Treadwell 22923

G. littorea Hartman 22924

Goniada sp. 22920

Goniadidae, unid. 22900

Gyptis brevipalpa Hartman-Schroeder 21923

Hyalinocia juvenalis Moore 23021

Langerhansia heterochaeta (Moore) 22221

Lumbrineris californiensis Hartman 23215

L. cruzensis Hartman 23216

L. lagunae Fauchald 23214

L. latreilli Audouin & Milne Edwards 23222

L. tetaura (Schmarda) 23229

Lumbrineris sp. 23210

Marpysa bellii oculata Treadwell 23131

Nephtys caecoides Hartman 22623

N. californiensis Hartman 22624

N. cornuta franciscana Clark & Jones 22625

N. ferruginea Hartman 22626

Nephtys sp. 22620

Nereidae, unid. 22400

Nereis sp. 22470

Ninoe gemmea Moore 23242

Nothria iridescentis (Johnson) 23035

Nothria sp. 23030

Onuphidae, unid. 23000

Onuphis eremita Audouin & Milne Edwards 23051

O. nebulosa Moore 23054

Onuphis sp. 23050

Ophiodromus pugettensis (Johnson) 21941

- Paleanotus septis (Johnson) 20921
 Paranaitis polynoides (Moore) 21321
 Peisidice era Johnson 20521
 Pholoe glabra Hartman 20621
 Phyllodocidae, unid. 21300
 Platynereis bicanaliculata (Baird) 22521
 Polyodontes panamensis (Chamberlin) 20531
 Protodorvillea gracilis (Hartman) 23531
 scaleworm, unid. 20200
 Sigambra tentaculata (Treadwell) 22042
 Sphaerodordium biseriale (Berkeley & Berkeley) 22711
 Sphaerodordium sp. 22910
 Steggoa californiensis Hartman 21351
 Sthenelais tertiaglabra Moore 20642
 Sthenelais verruculosa Johnson 20643
 Sthenelanella uniformis Moore 20651
 Streptosyllis sp. 22310
 Syllidae, unid. 22200
 Typosyllis sp. 22280
- EDENTARIA**
- Aedicira pacifica (Hartman) 25212
 A. ramosa (Annenkova) 25213
 Amage anops (Johnson) 28011
 Ammotrypane aulogaster Rathke 27011
 Ampharete arctica Malmgren 28032
 A. labrops Hartman 28034
 Ampharete sp. 28030
 Ampharetidae, unid. 28000
 Amphictesis scaphobranchiata Moore 28043
- Amphisamytha bioculata (Moore) 28051
 Apopronospio pygmaeus (Hartman) 25615
 Aricidea neosuecica Hartman 25223
 A. suecica Eliason 25224
 A. wassi Pettibone 25225
 Aricidea sp. 25220
 Armandia bioculata Hartman 27021
 Axiothella rubrocincta (Johnson) 27522
 Boccardia basilaria Hartman 25511
 Capitella capitata (Fabricius) 27231
 Capitellidae, unid. 27200
 Chaetozone corona Berkeley & Berkeley 26332
 C. setosa Malmgren 26335
 Chaetozone sp. 26330
- Chone ecaudata (Moore) 28621
 C. veleronis Banse 28626
 Chone sp. 28620
 Cirriformia sp. 26350
 Clymenura gracilis Hartman 27551
 Cossura candida Hartman 26511
 Decamastus gracilis Hartman 27261
 Euchone incolor Hartman 28652
 Flabelligeridae, unid. 26700
 Haploscoloplos elongatus (Johnson) 25121
 Harmothoe crassicirrata Johnson 20271
 H. lunulata (delle Chiaje) 20276
 H. priops Hartman 20277
 Harmothoe sp. 20270
 Hesionidae, unid. 21942
 Hesperone complanata (Johnson) 20292
 Heteroclymene glabra Moore 27571
 Laonice cirrata (Sars) 25541
 Loimia medusa (Savigny) 18291
 Magelona pitelkai Hartman 25813
 M. sacculata Hartman 25814
 Magelona sp. 25810
 Maldane cristata Treadwell 27611
 M. sarsi Malmgren 27613
 Maldanidae, unid. 27600
 Megalomma splendida (Moore) 28724
 Melinna heterodonta Moore 28111
 M. oculata Hartman 2812
 Myriochele gracilis Hartman 27711
 Nerinides acuta (Treadwell) 25571
 N. maculata Hartman 25572
 N. pigmentata (Reish) 25573
 Notomastus sp. 27330
 Paraonidae, unid. 25200
 Paraonis gracilis oculata Hartman 25251
 Parapronospio pinnata (Ehlers) 25614
 Pectinaria californiensis Hartman 27921
 Pherusa sp. 26760
 Pista disjuncta Moore 28344
 P. fasciata (Grube) 28346
 P. moorei Berkeley & Berkeley 28348
 Pista sp. 28340
 Poecilochaetus johsoni Hartman 26011
 Polycirrus perplexus Moore 28362

SEDENTERIA (Cont)

- Polydora citrona Hartman 25583
— Polydora sp. 25580
— Potamilla neglecta (Sars) 28761
— Praxillella affinis pacifica Berkeley 27661
— P. gracilis (Sars) 27662
✓ Prionospio cirrifera Wiren 25611
✓ P. malmsgreni Claparede 25613
— Sabellaria cementarium Moore 27832
Sabellidae, unid. 27800
Scalibregma inflatum Rathke 26931
— Scoloplos armiger (Mueller) 25163
— Spio sp. 25650
✓ Spiochaetopterus costarum (Claparede) 26241
Spionidae, unid. 25600
✓ Spiophanes bombyx (Claparede) 25662
✓ S. cirrata Berkeley & Berkeley 25666
✓ S. missionensis Hartman 25664
✓ Spiophanes sp. 25660
✓ Sternaspis fossor Stimpson 27111
✓ Streblosoma crassibranchia Treadwell 28421
Terebellidae, unid. 28200
✓ Terebellides stroemi Sars 28512
✓ Tharyx sp. 26380
— Travisia brevis Moore 27061
— T. foetida Hartman 27062

SIPUNCULOIDEA

- Sipunculoidea, unid. 94300

ECHIUROIDEA

- Listriolobus pelodes Fisher 94411

ARTHROPODA

CRUSTACEA

OSTRACODA

Cylindroberididae, unid. 52043

Cyprinidae, unid. 52250

Cythereidae, unid. 52324

Euphilomedes carcharodonta (Smith) 52029

E. longiseta Juday 52031

E. oblonga Juday 52033

E. producta Poulsen 52032

Euphilomedes sp. 52028

Myodocopidae, unid. 52001

~~RUTIDERMA~~ Philemedes lomae (Juday) 52086

Rutiderma rostrata Juday 52088

Rutiderma sp. 52083

Scleroconcha sp. 52037

CIRRIPEDIA

Scalpellum californicum Pilsbry 54311

Scalpellum sp. 54310

MALACOSTRACA

NEBALIACEA

Nebaliacea, unid. 55000

mysidacea

Mysidacea, unid. 58000

Acanthomysis macropsis Tattersall 58733

Acanthomysis sp. 58730

Archaeomysis grebnitzkii Czerniavsky 58512

Archaeomysis sp. 58510

mysidacea (cont)

Neomysis kadiakensis Ortmann 58723

Neomysis sp. 58720

CUMACEA

- Anchicolurus occidentalis Calman 59111
Campylas is canaliculata Zimmer 59148
C. hartae Lie 59149
C. rubromaculata Lie 59151
Campylaspis sp. C 59143
Cumella sp. A 59131
Cumella sp. B 59132
Cumella sp. C 59133
Cyclaspis sp. A 59162
Cyclaspis sp. B 59163
Cyclaspis sp. 59160
Diastylidae, unid. 59169
Diastylis californica Zimmer 59171
Diastylis sp. A 59172
Diastylis sp. 59170
Diastylopsis tenuis Zimmer 59271
Eudorella pacifica Hart 59191
Hemilamprops californica Zimmer 59211
Lampropidae, unid. 59219
Lamprops carinata Hart 59221
L. quadriplicata Smith 59222
Lamprops sp. 59220
Leptostylis sp. A 59361
Leucon subnasica Given 59241
Leucon sp. 59240
Mesolamprops bispinosa Given 59321
Oxyurostyliis pacifica Zimmer 59251

TANAIDACEA

- Leptochelia sp. 60410
Leptognathia sp. 60420

ISOPODA

VALVIFERA 61100

- Edotea sublittoralis Menzies & Barnard 61381
Neastacilla californica (Boone) 61221
Syndotea magnifica Menzies & Barnard 61343

ANTHURIDEA 61600

- Hallophasma geminata Menzies & Barnard 61751

GNATHIIDEA 63400

- Gnathia crenulatifrons Monod 63514

- Gnathia sp. 63510

- ASELLOTA 60700 63700
Austrosignum tillerae Menzies & Barnard 64631
Munna sp. 64640
Pleurogonium sp. 64620

AMPHIPODA

GAMMARIDEA 70000

- Gammaridea, unid. 70000
Ampelisca brevisimulata Barnard 70211
(A. compressa Holmes) 70215 = A. acassiei Judd
A. cristata Holmes 70212
A. cucullata Barnard 70229
A. hancocki Barnard 70219
A. indentata Barnard 70234
"A. macrocephala Liljeborg 70222
A. milleri Barnard 70226
Ampelisca pacifica Holmes 70214
A. pugetica Stimpson 70216
A. cf. shoemakeri Barnard 70239
Ampelisca sp. 70210
Amphideutopus oculatus Barnard 70671
Aoridae, unid. 70600
Byblis veleronis Barnard 70251
Byblis sp. 70250
Eohaustorius washingtonianus (Thorsteinson) 72421
Gammaropsis sp. 72890
Gitana calitemplado Barnard 70331
Heterophoxus oculatus (Holmes) 74671
Isaeidae, unid. 72800
Isaeidae sp. A 72801
Lembos audbetti Barnard 70621
Listriella diffusa Barnard 73515
L. goleta Barnard 73511
L. melanica Barnard 73513
Listriella sp. 73510
Lysianassidae, unid. 73600
Mandibulophoxus uncirostratus (Giles) 74661
Megaluropus longimerus Schellenberg 72231
? Melphisana bola Barnard 74012
Metaphoxus frequens Barnard 74641
Metopa cf. dawsoni Barnard 75325
Microjassa litotes Barnard 72931
Monoculodes sp. 74130
Oedicerotidae, unid. 74100

GAMMARIDEA (Cont)

- Paraphoxus bicuspidatus Barnard 74611
P. heterospidatus Barnard 74623
P. obtusidens (Alderman) 74615
Paraphoxus sp. 74610
Parapleustes sp. 74810
Photis bifurcata Barnard 72811
P. brevipes Shoemaker 72812
P. californica Stout 72813
P. macrotica Barnard 72817
Photis sp. 72810
Pleustidae, unid. 74900
Rudilemboides stenopropodus Barnard 70651
Stenothoidae, unid. 75300
Synchelidium sp. 74110
Tiron biocellata Barnard 75521
Westwoodilla caeca (Bate) 74121

CAPRELLIDEA 79000

- Caprella californica Stimpson 79012
C. equilibra Say 79011
C. mendax Mayer 79022
Caprella sp. 79010
Mayerella banksia Laubitz 79071
Tritella pilimana Mayer 79052
Tritella sp. 79050

EUPHAUSIACEA

- Euphausiacea, unid. 66000

DECAPODA

- Decapoda, unid. 89900

CARIDEA

- Crangon communis Rathbun 82928

ANOMURA

- Anomura, unid. 86000
Callianassa sp. 86210
Callianassidae, unid. 86200
Lepidopa myops, zoea Stimpson 86922
Porcellanidae, unid. 87200

BRACHYURA

- Brachyura, unid. 88000
Cancer sp. 88910
?Parapinnixa sp. 89430
Pinnixa franciscana Rathbun 89447
P. occidentalis Rathbun 89446
Pinnixa sp. 89440
Pinnotheridae, unid. 89400
Randallia ornata (Randall) 88411

INSECTA

- Insecta, unid.

MOLLUSCA

GASTROPODA 30000

MESOGASTROPODA

- Balcis micans (Carpenter) 31722
B. oldroydi (Bartsch) 31725
B. rutila (Carpenter) 31723
Bittium quadrifilatum Carpenter 33327
Caecum crebricinctum Carpenter 33141
Epitonium cf. acrostephanus (Dall) 31935
Eulima californica (Bartsch) 31711
Fartulum occidentale Bartsch 33131
Neverita recluziana (Deshayes) 34261

NEOGASTROPODA

- Amphissa sp. 35920
Kurtzia arteaga (Dall & Bartsch) 37131
Kurtziella beta (Dall) 37122
K. plumbea (Hinds) 37124
Mitrella sp. 35930
Nassarius fossatus (Gould) 36022
Olivella baetica Carpenter 36512

OPISTHOBRANCHIA

- Acteocina intermedia Willet 38271
Acteocina sp. 38270
Aglaja ocelligera (Bergh) 38245
Aglaja sp. 38240
Cephalaspidea sp. A 3801
Cyllichna diegensis (Dall) 38282
Gastropteron pacificum Bergh 38261
Odostomia sp. C 38020
Rictaxis punctocaelatus (Carpenter) 38131

Turbonilla sp. D 38044
Turbonilla sp. E 38045
Turbonilla sp. G 38046
Turbonilla sp. J 38047
Volvulella californica Dall 38193
V. cylindrica (Carpenter) 38191
V. panamica Dall 38192
Volvulella sp. 38190

LECYPODA 40000
Pelecypoda, unid. 40000

NUCULOIDA
Acila castrensis (Hinds) 40211
Nucula sp. 40220
Nuculana sp. 40320
Yoldia scissurata (Dall) 40343

MYTILOIDA

Amygdalum pallidulum (Dall) 41121
Crenella decussata (Montagu) 41131
C. divaricata (Orbigny) 41132
Crenella sp. 41130
Megacrenella columbiana (Dall) 41231
Modiolus neglectus Soot-Ryen 41182
Mytilidae, unid. 41100

VENEROIDA

Adontorhina cyclica Berry 43211
Axinopsida serricata (Carpenter) 43221
Compsomyax subdiaphana (Carpenter) 43581
Cooperella subdiaphana (Carpenter) 43911
Cyclocardia ventricosa (Gould) 42212
Cyclocardia sp. 42210
Gari californica (Conrad) 44411
Lepton merceum Carpenter OROBITELLA CHACCI 42931
Lucina nuttallii (Conrad) 43151
Lucinoma annulata (Reeve) 43142
Macoma acolasta Dall 44226
M. carlottensis Whiteaves 44224

VENEROIDA (Cont)
✓ Macoma sp. 44220
Mactra californica Conrad 44012
Mysella pedroana (Dall) 42923
Mysella sp. C 42927
✓ Mysella sp. 42920
✓ Nemocardium centifilosum (Carpenter) 43461
Parvilucina sp. 43180
Siliqua lucida (Conrad) 44731
Solen sp. 44710
Tellina bodegensis (Hinds) 44211
T. modesta Carpenter 44216
Tellina sp. 44210
Thyasira flexuosa (Montagu) 43233

MYOIDA

Corbula luteola Carpenter 44911
Corbula sp. 44910
Panopea generosa (Gould) 45041

PHOLADOMYOIDA

Cardiomya californica (Dall) 46012
C. cf. pseustes Dall 46016
Cuspidaria parapodema Barnard 46022
Lyonsia californica Conrad 45621
Periploma discus Stearns 45711
Thracia curta Conrad 45831
T. diegensis Dall 45933

SCAPHOPODA

Scaphopoda, unid. 48600
Cadulus fusiformis Pilsbry & Sharp 48761
Dentalium rectius Carpenter 48692

APLACOPHORA

Chaetoderma cf. nitidulum (Loven) 48127
Chaetoderma sp. 48110

PHORONIDA 44050

Phoronida, unid. 44050
Phoronis sp. 44060

ECTOPROCTA 93300

GYMNOLAEMASTA 93301

CTENOSTOMATA 93866

Alcyonium mammillatum Alder 93872

BRACHIOPODA 94001

Glottidia albida (Hinds) 94005

ECHINODERMATA

ASTEROIDEA 96200

Asteroidea, unid. 96200

✓ Astropecten verrilli De Loriol 96222

OPIHUROIDEA 97000

✓ Ophiuroidea, unid. 97000

ECHINOIDEA 96600

✓ Echinoidea, unid. 96600

Dendraster excentricus (Eschscholtz) 96641

HOLOTHUROIDEA 97600

Holothuroidea, unid. 97600

Caudina sp. 97690

Leptosynapta sp. B 97673

HEMICHORDATA 98000

Enteropneusta, unid. 98000

CHORDATA

ASCIDIACEA 98100

Ascidiae, unid. 98100

CEPHALOCORDATA 98600

✓ Branchiostoma californiense Andrews 98611

VERTEBRATA

PISCES

Pisces, unid.

Icelinus quadriseriatus (Lockington) 16487

Syphurus atricauda (Jordan & Gilbert) 139000

Amphiodia sp. 97140

Amphiodia occidentalis 97161

Amphiodia (Amphipriva) 97170

A. (A) urtica 97171

A. (A). digitata 97172

Amphiuridae 97120

Amphichondrus granulosus 97131

Amphipholis squamata 97191

Amphiura sp. 97200

A. arcystata 97233

Ophura lutkeni 97352

Astropectinidae 96220

Astropecten sp. 96220

A. verrilli 96222

Brissopsis sp. 96680

OK, Good job.
- Gary Williams, supervisor, proposed
- Get Mearns help in getting
- back to the stations,
apm

MEMORANDUM

To: Alan Mearns

From: Charles Greene

Re: Benthic Grab Survey of Orange County Outfalls
(29-31 July 1975)

Date: July 16, 1975

In compliance with the revised EPA Research Plan (Biology), a detailed sampling program for the Orange County outfall area has been designed. This survey will provide follow-up data on Gary Smith's study and a broad detailed data base for comparison with future studies. The plan is based on a review of the results obtained from the preliminary survey of this area conducted by SCCWRP in February and March, 1975.

The survey is tentatively scheduled for 29-31 July 1975, with the Marine Surveyor.

It is recommended that the 52 stations shown on Figure 1 be occupied and samples taken with the weighted (G. Smith's) Van Veen Grab (0.1 m^2) according to the following schedule:

A. Biological Samples (C. Greene)

1. Three replicate samples will be taken at 26 stations and a single sample at each of the remaining 26 stations (Figure 1 and Table 1).
2. Sample volume, color, texture, and odor, if any, should be recorded along with station number and depth for each sample.
3. Samples collected at the 6 Smith stations (Figure 1 and Table 1) are to be washed over 1.0 and 0.5 mm screens. All remaining samples are to be washed over a 1.0 mm screen.
4. Screened samples shall be preserved in 10% buffered formalin and submitted to an outside agency for sorting.

B. Physical/Chemical Samples (B. Mitchell & D. Young)

1. These samples will be sub-samples from the biological samples and consist of the following:
 - a. One $2\frac{1}{2}$ " diameter core/sample (98 cores) - to be capped and frozen up-right.
 - b. One 1" x 2" core/station at the 23 station indicated on Table 1 to be preserved for acid volatile sulfide analysis.
 - c. One 50-100 gms of sediment (4-5 cm deep) from each station for grain-size analysis.
 - d. One scoop (1-2 cm from surface) in small glass jar and frozen for DDT, PCB, and ACB's.
2. Physical/chemical samples will be returned to SCCWRP at the end of each day for storage (freezing) or immediate analysis (BOD). The sub-sample for BOD analysis is to be taken from the sediment grain-size samples.

A detailed schedule of times, places, and personnel will be circulated next week.

John [unclear]

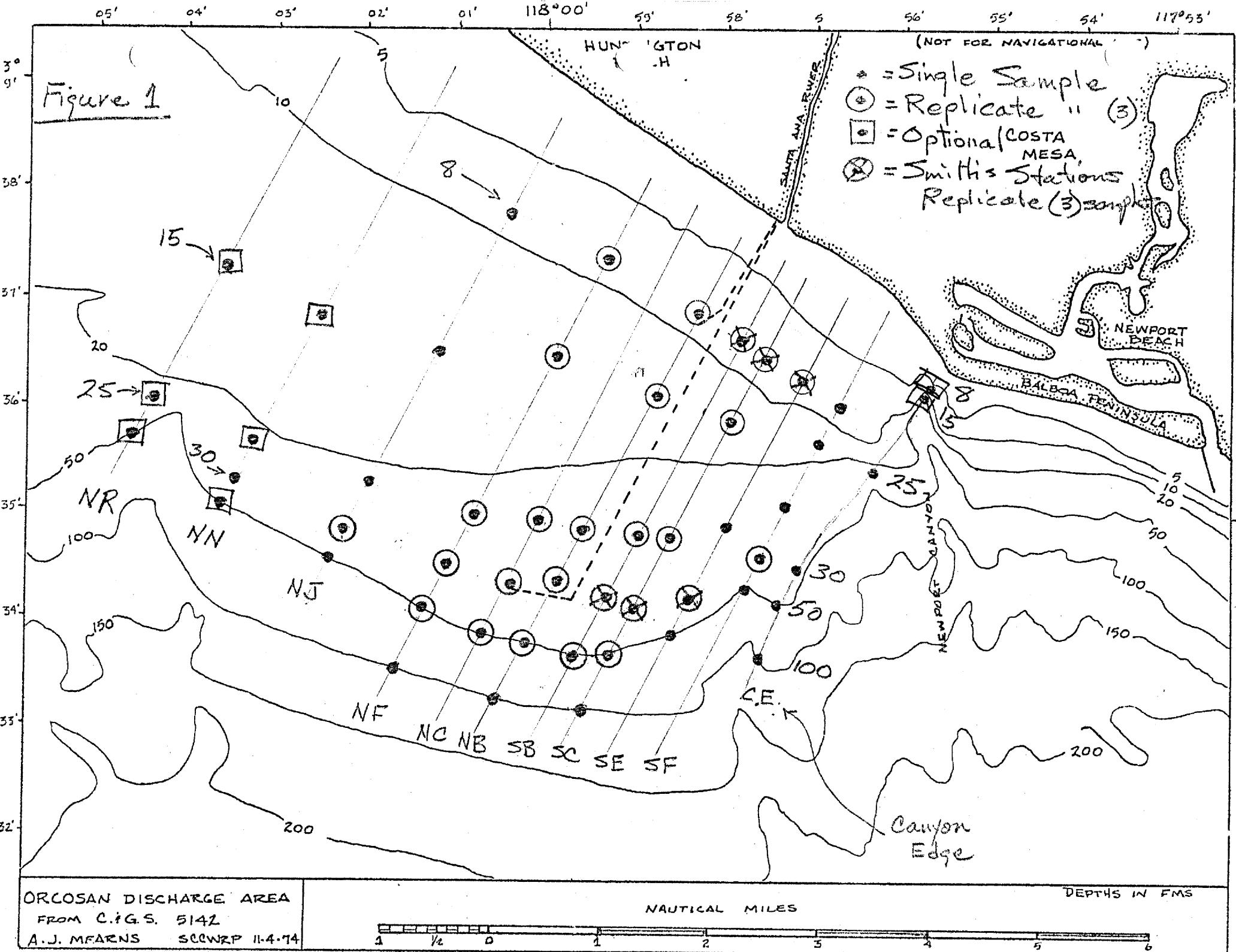


Table 1. Summary of grab samples and physical/chemical sub-samples to be collected off Orange Co. - 29-31 July 1975.

	DEPTH					
	8 (FM)	15 (FM)	25 (FM)	30 (FM)	50 (FM)	100 (FM)
C E	1 Grab	1 Grab	1 Grab	1 Grab	1 Grab	1 Grab
S F	1 Grab	1 Grab	1 Grab	3 Grab	1 Grab	
S E	3 Grabs *		1 Grab	3 Grabs *	1 Grab	
S C	3 Grabs *	3 Grabs *	3 Grabs *	3 Grabs *	3 Grabs *	1 Grab
T R	T S B	3 Grab *		3 Grabs *	3 Grabs *	3 Grabs *
A N S	N B	3 Grabs	3 Grabs *	3 Grabs *	3 Grabs *	1 Grab *
E C T	N C		3 Grabs *	3 Grabs *	3 Grabs *	
N F	3 Grabs	3 Grabs	3 Grabs *	3 Grabs *	3 Grabs *	1 Grab
N J	1 Grab *	1 Grab	1 Grab	3 Grabs *	1 Grab	
N N		1 Grab Opt.	1 Grab Opt.	1 Grab *	1 Grab Opt.	
N R		1 Grab Opt.	1 Grab Opt.		1 Grab Opt.	

- 1) 2 1/2" core to depth of sample - 1/sample = 104 cores
- 2) 1-2 cm deep scoop of surface for DDT - 1/station = 52
- 3) 50-100 gm surface sediment (4-5cm) - 1/station = 52
- 4) * 1" x 2" core placed in preservative - see above

MEMORANDUM

To: Willard Bascom
From: Mike Moore
Re: Orange County Benthic Grab Survey

February 18, 1975

Of the 47 stations to be sampled on the 15th of February, only 15 were completed due to high seas and high winds. We found it impossible to stay on station and to get Dr. Gary Smiths modified van veen grab to penetrate and obtain a sample.

The stations that were completed are now being analyzed for biological community structure and for volatile solids as stated in the project outline.

A brief outline of area sampled: (see chart) Depths: 8, 15, 25, 30, 50, 100 fathoms. Bottom type: Varied from solid sand in the shallow stations to a silty-clay in deeper stations. Shell fragments were found in large quantities in the 8-50 fathom stations. Field Biology Observation: In the canyon stations we found what appeared to be a polychaete community (Terebelids, malacophids) whereas in the flatland stations we found what appeared to be a brittle star, sea urchin community (Ophiofragmus, Lytichinus). This was only from a quick inspection of the screened sediments and is not intended to be an accurate statement of the community structure of the area.

Orange County - Two surveys in 1975

Purpose

1. To provide background data for a detailed comparison of areas surrounding the sewage diffuser pipe in Huntington Beach. The detailed comparison would be to determine effects (if any) of a modern sewage diffuser on the benthic marine environment.
2. To compare stress and recovery at both the one mile and the five mile outfall.

Surveys

- I. A rough descriptive analysis of an area approximately four miles by five miles surrounding both discharge sites consisting of forty stations. One bottom grab sample for each

station will be taken and analyzed for:

1. Dominant species
2. Volatile solids

One day in the field will be needed to provide these data to justify the limiting of the size of the sample area for the detailed comparison survey and to provide the background data needed for that survey.

- II. A detailed chemical and biological analysis of the area of expected effect. One field day will be needed to obtain samples.

A. Parameters

1. Water column

- a. temperature
- b. D.O. on boat
- c. Salinity C. Smith
- d. pH

2. Bottom types

- a. sand-silt Observation on boat (Word and Moore)
- b. silt-clay
- c. sediment grain size analysis (Greene, Gorsline)

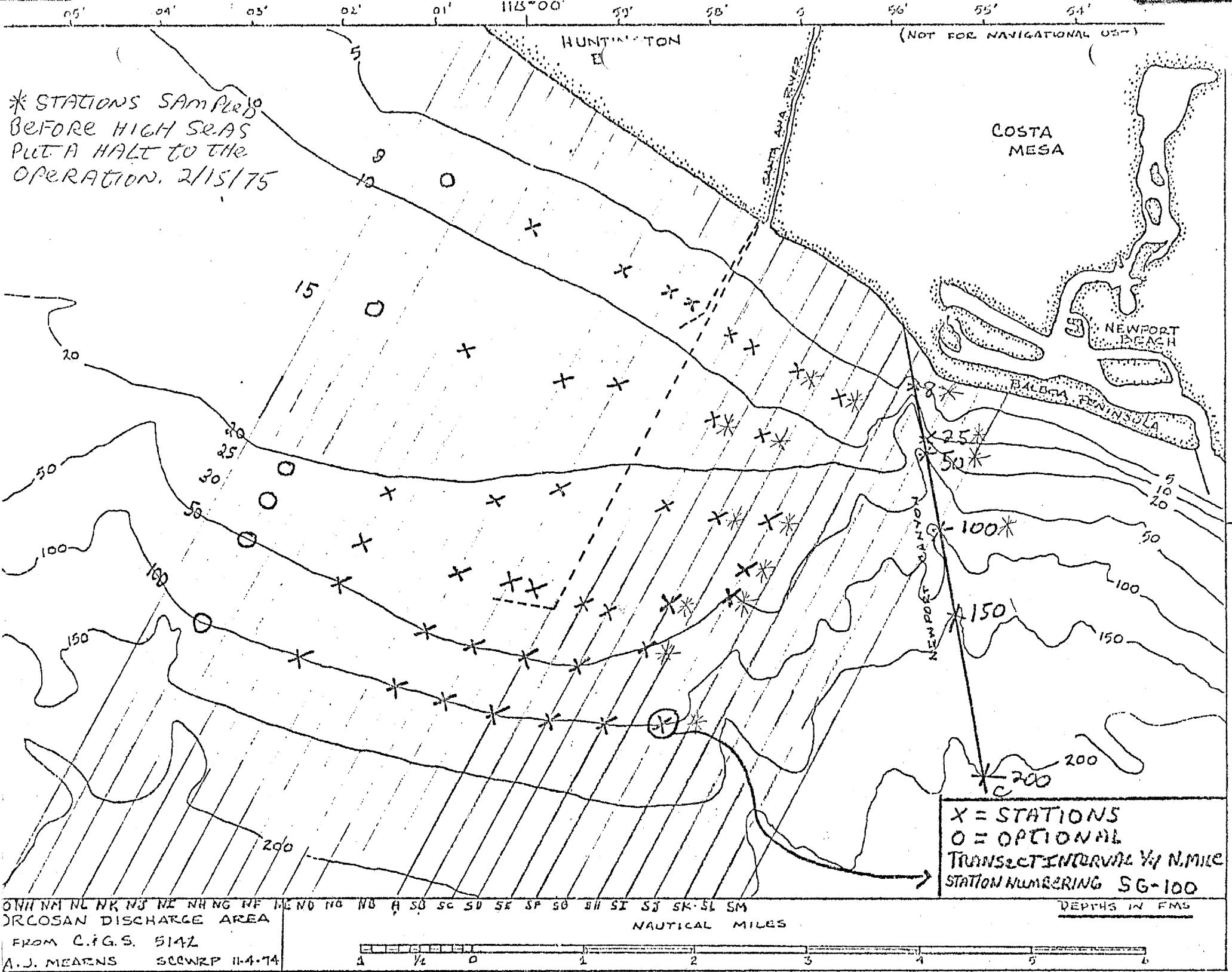
3. Outfall effects physical measurements

- a. COD
- b. BOD Most samples
 1. BOD/COD
- c. Volatile solids (all samples)
- d. organic carbon (2 or 3 for each range of COD levels) and organic nitrogen (M.B.C.)
- e. metals
 1. Cr (total), Cu, Cd, -(Ni, Zn), Se, At
- f. chlorinated hydrocarbons (DDT, DDE, ACBs)
- g. Hexane extractibles (oil and greese)

4. Biological Analysis

- a. Two grabs will be screened and preserved on board boat to be later analyzed by outside laboratory under supervision of SCCWRP M.B.C. \$60/sample.

A total of three benthic grabs per station will be used for the biological analysis. One sub core from each of these grabs will be used for the physical parameters.



MEMORANDUM

To: Jack Word

April 17, 1975

From: Brad Myers

Re: Orange County Sanitation District Outfall:
Preliminary Study

I completed the initial investigation of benthic organisms from ORCOSAN with an average effort of about 3 hours per sample. Several species that I feel occur uncommonly were observed, as well as a depth extension for Cephalaspidean sp A.

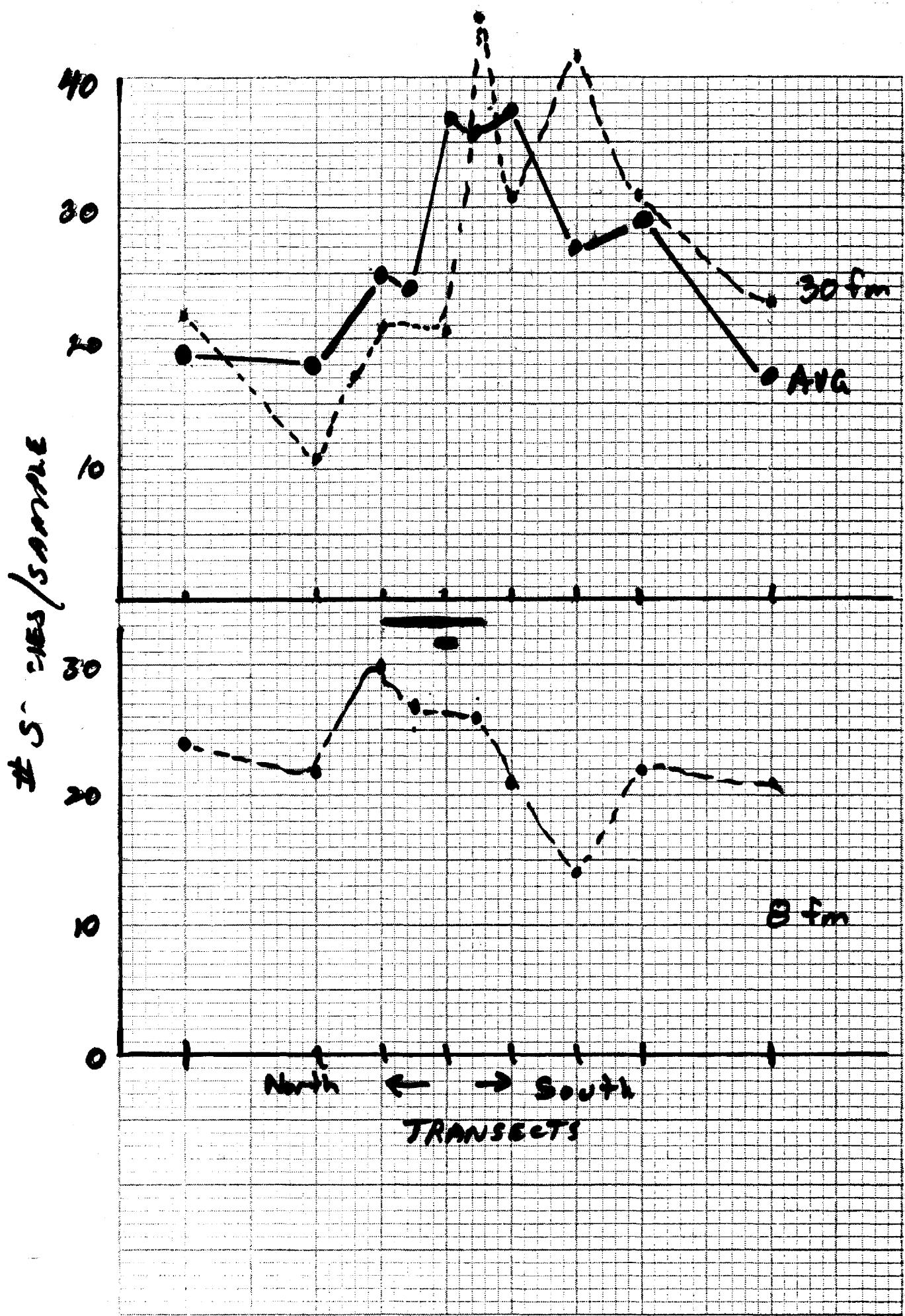
I attempted to observe any trends of populations in the area of the outfall by plotting occurrence and relative abundance for 10 species (or assemblages) on station charts. The observed trends were most evident in Euphilomedes carcharodonta and E. producta both detritus feeders, whose highest abundances occurred within the area of the sludge field (see overlay for sludge field). Prionospio occurred in high abundance near the outfall at 3 of the 4 stations where it was abundant. All stations where Lucinidae (Lucinoma, parvilucina, etc.) occurred in high abundance corresponded with the sludge field around the outfall. None of the other species plotted showed any trends relating to the sludge field.

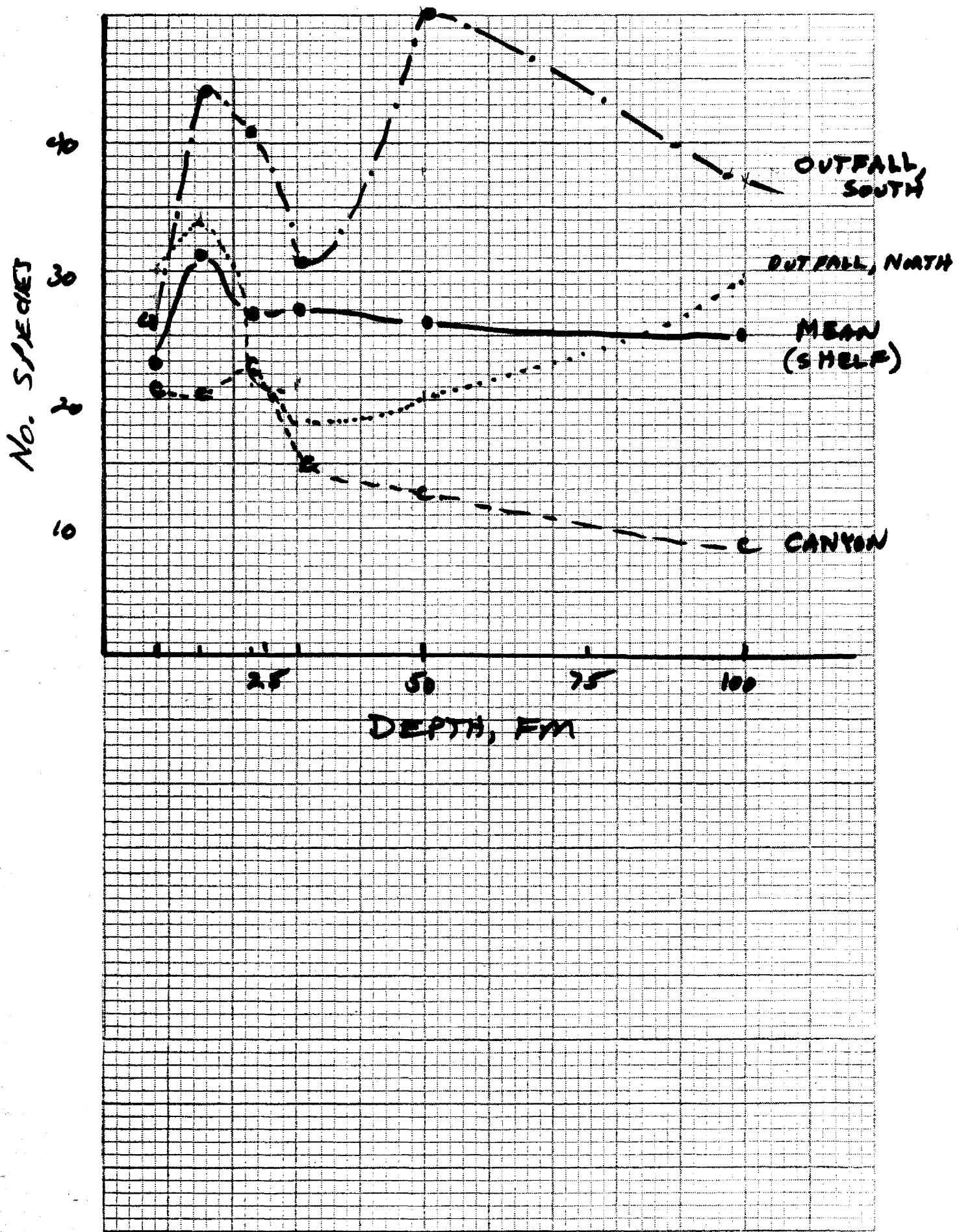
I also applied an overlay of grainsize data to the plotted species, however no correlation could be made from the information. An interesting point however was that Pectenaria did not occur in abundant numbers in any sandy areas, nor was it collected at any stations less than 25 fm (See plotted chart).

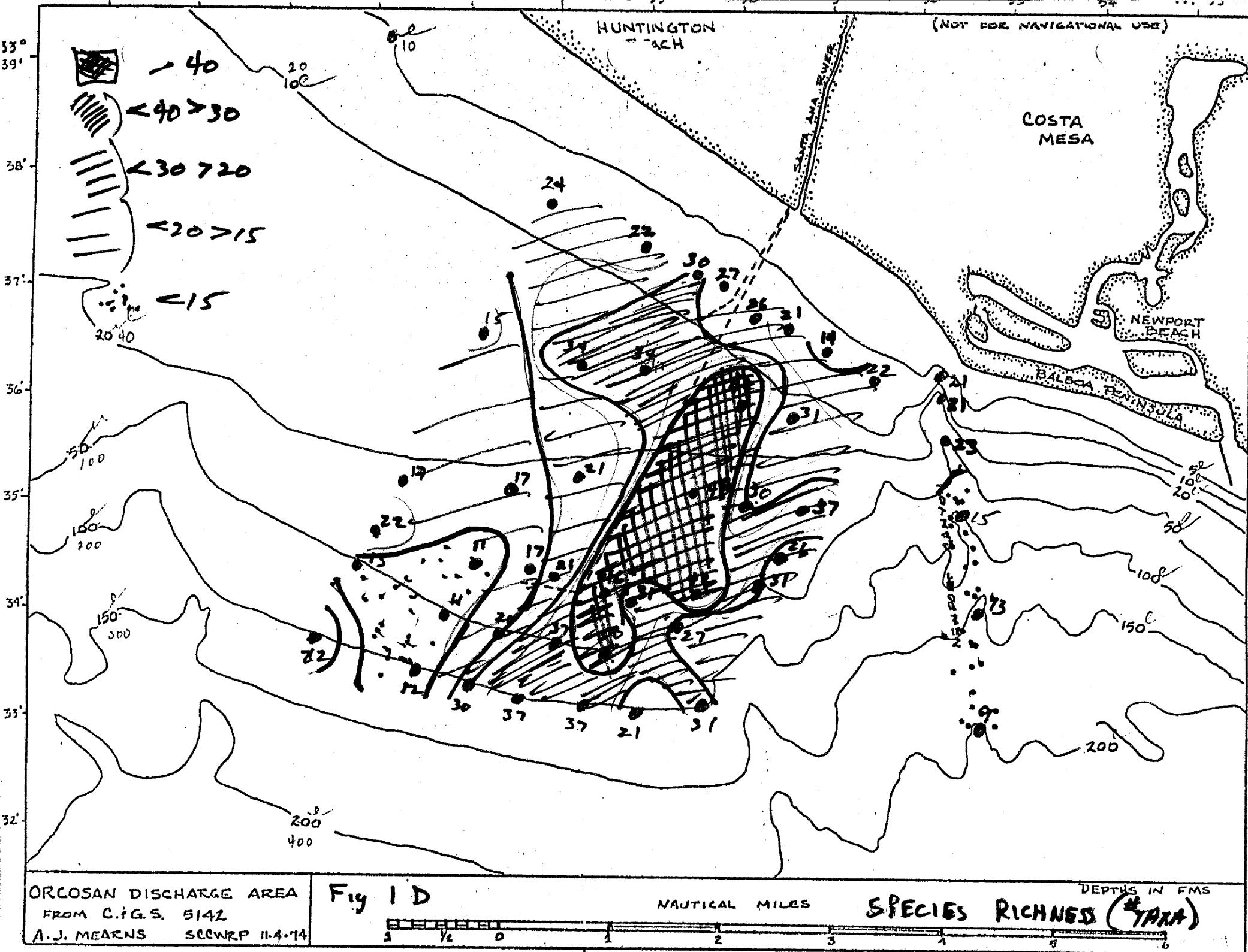
Numbers of species observed were also plotted by stations, as well as mean numbers of species by transect and strata, this showed a general increase in number of species near the outfall, as well as a generally higher number of species south of the pipe (between the pipe and the canyon). The canyon transect showed very low species diversity and numbers of animals, generally decreasing with depth.

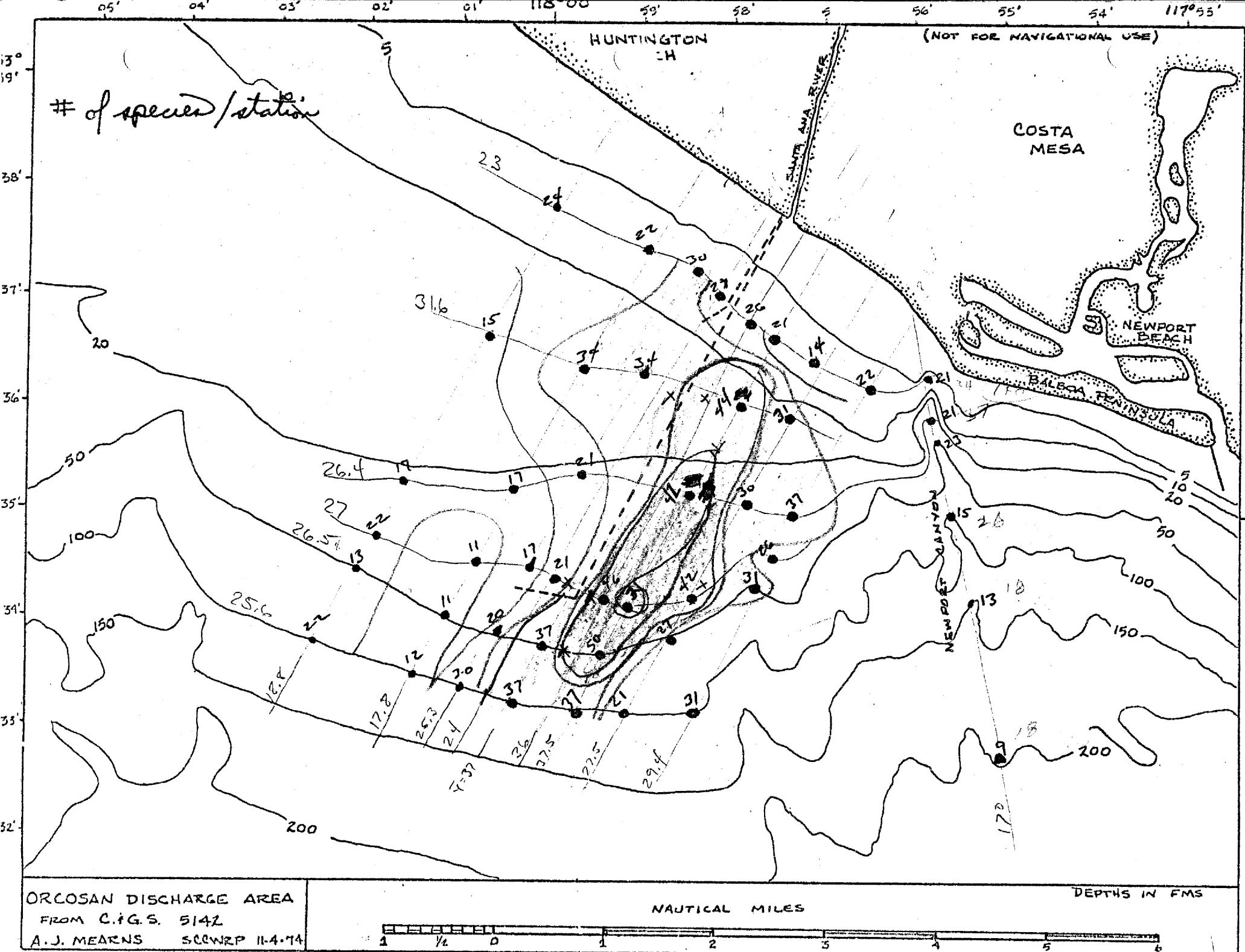
Please look at information and see if you have any other observations you can add.

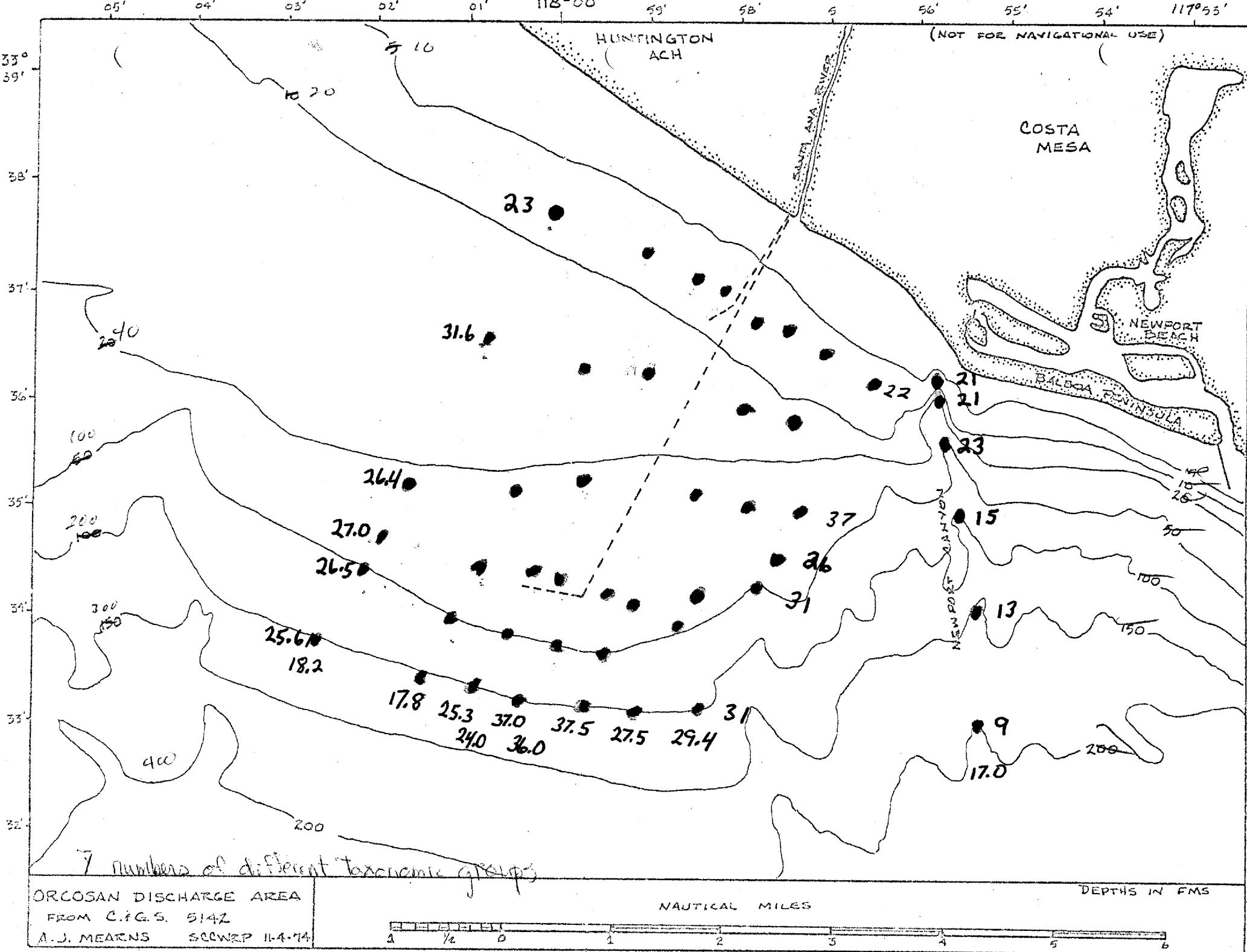


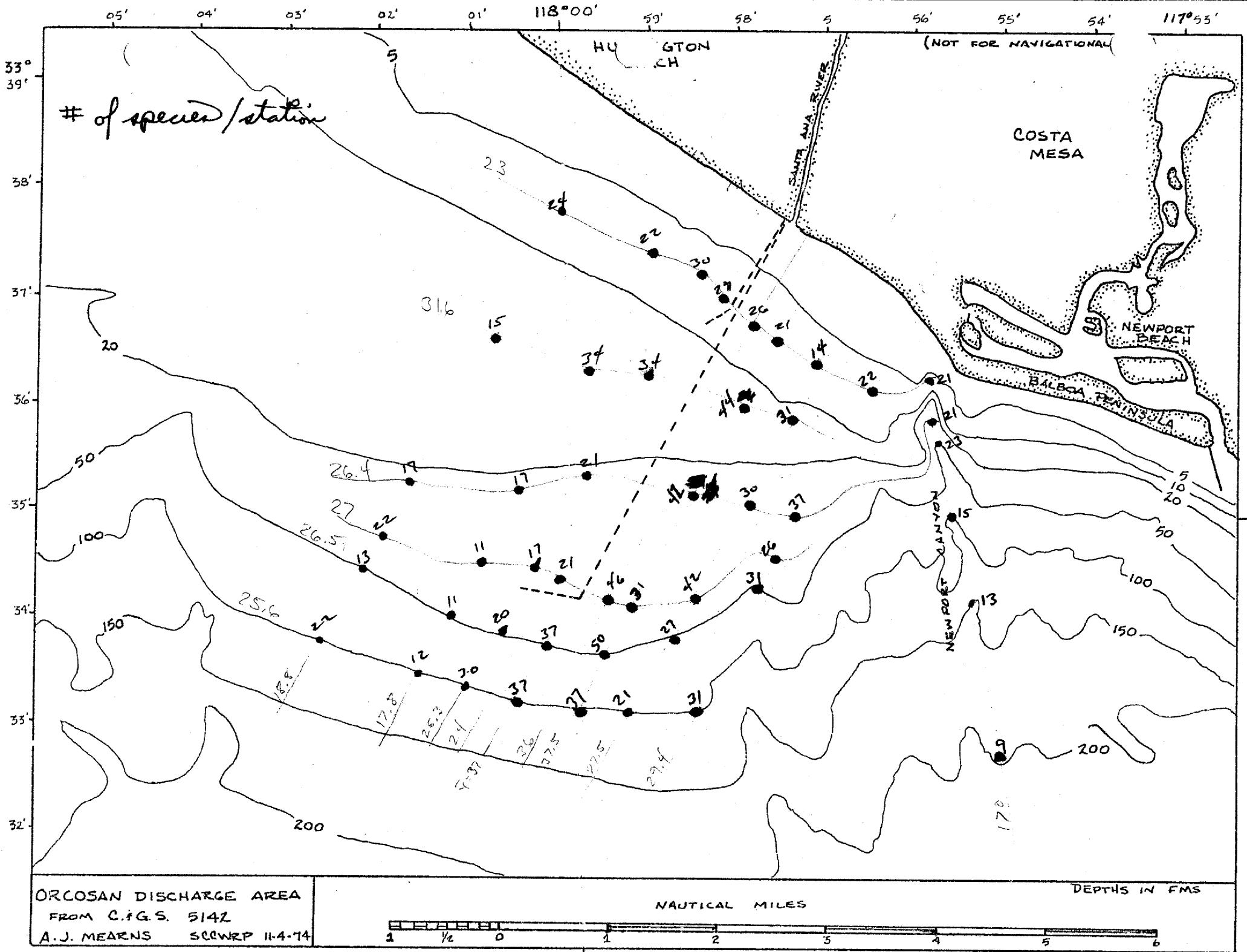












05' 04' 03' 02' 01' 118-00 59' 58' 5 56' 55' 54' 117°53'

(NOT FOR NAVIGATION)

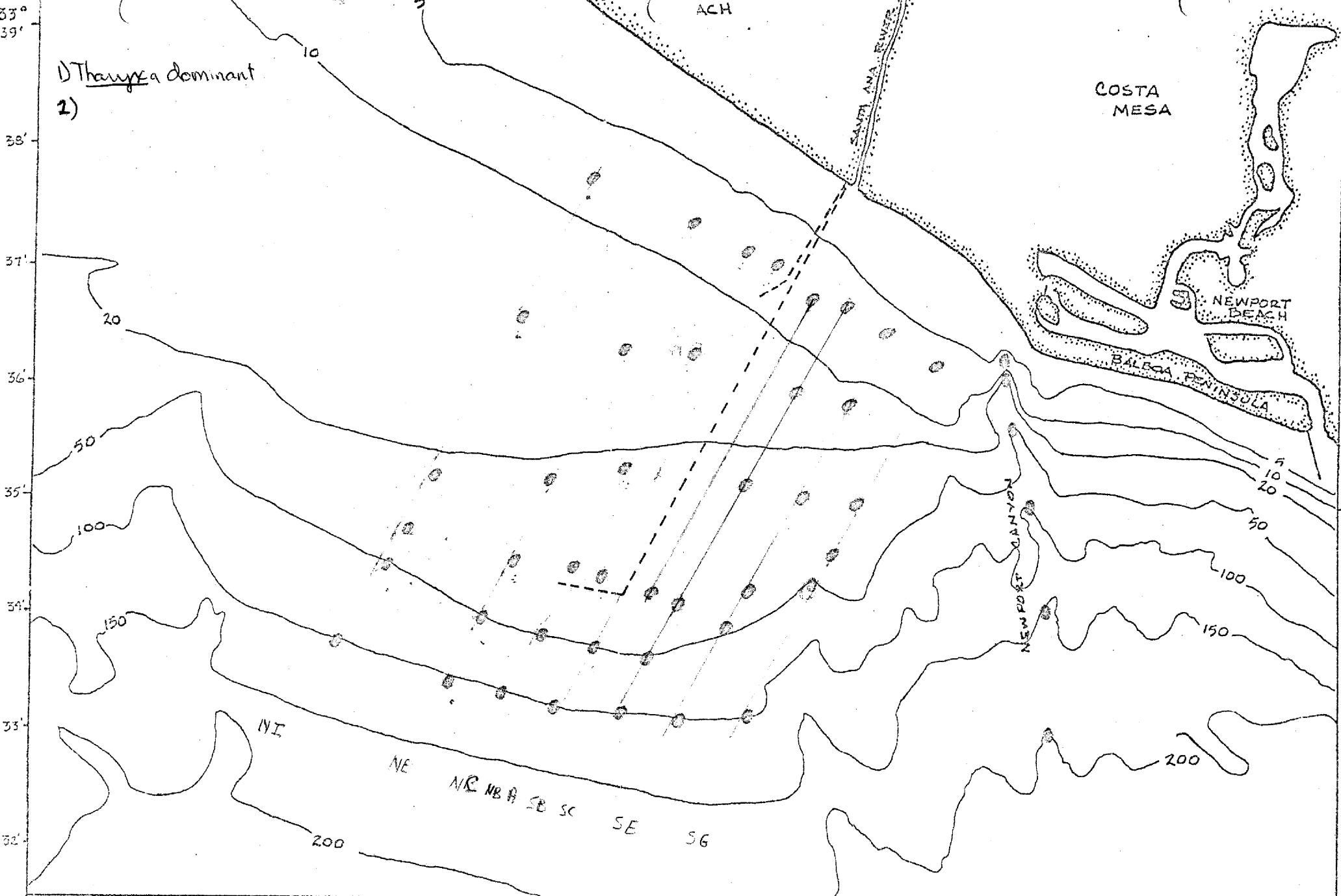
HUNTINGTON
BEACH

- 1) Thalassia dominant
2)

COSTA
MESA

NEWPORT
BEACH

BALBOA
PENINSULA

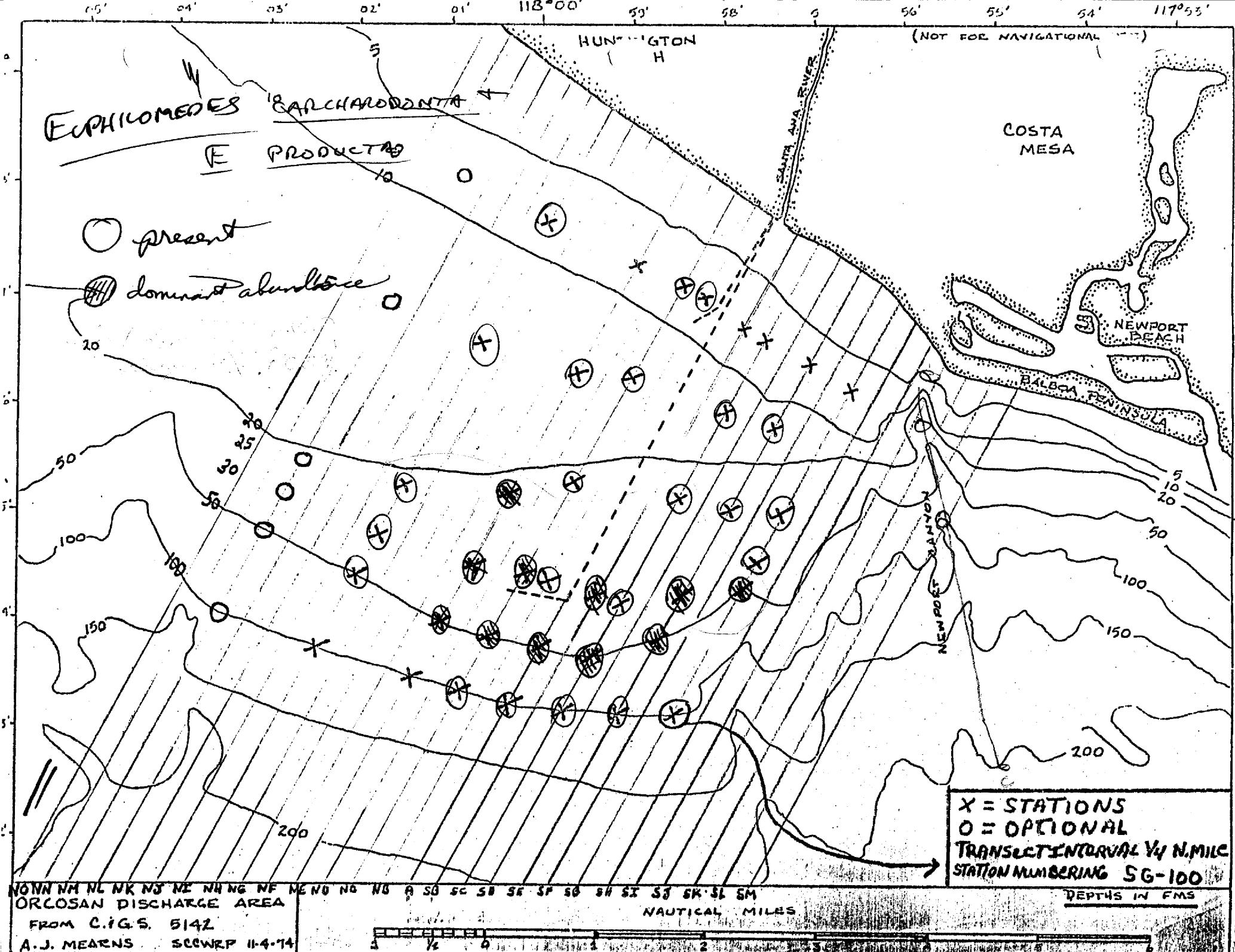


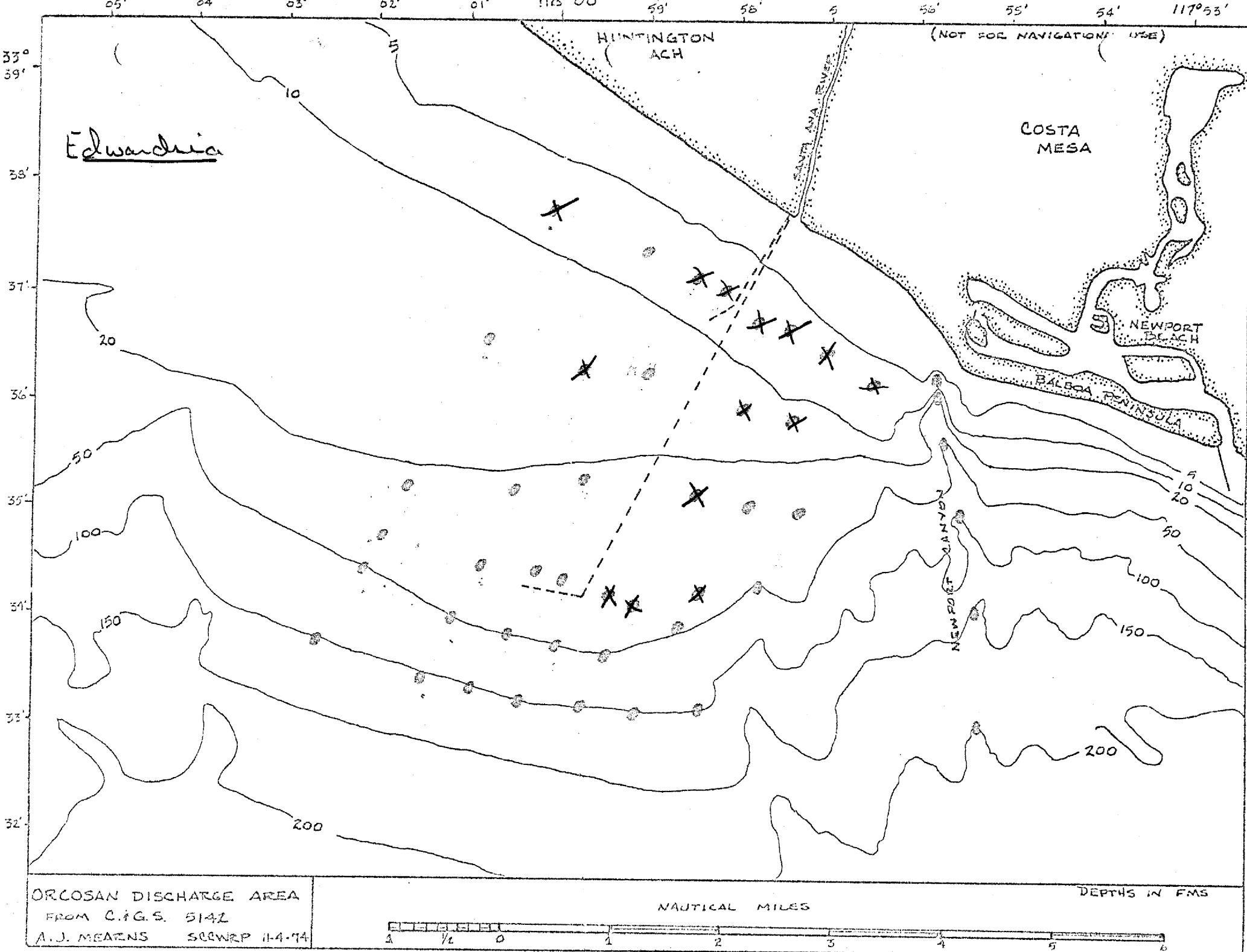
ORCOSAN DISCHARGE AREA
FROM C.G.S. 5142
AUGUST 1974
SCSWRP 11-4-74

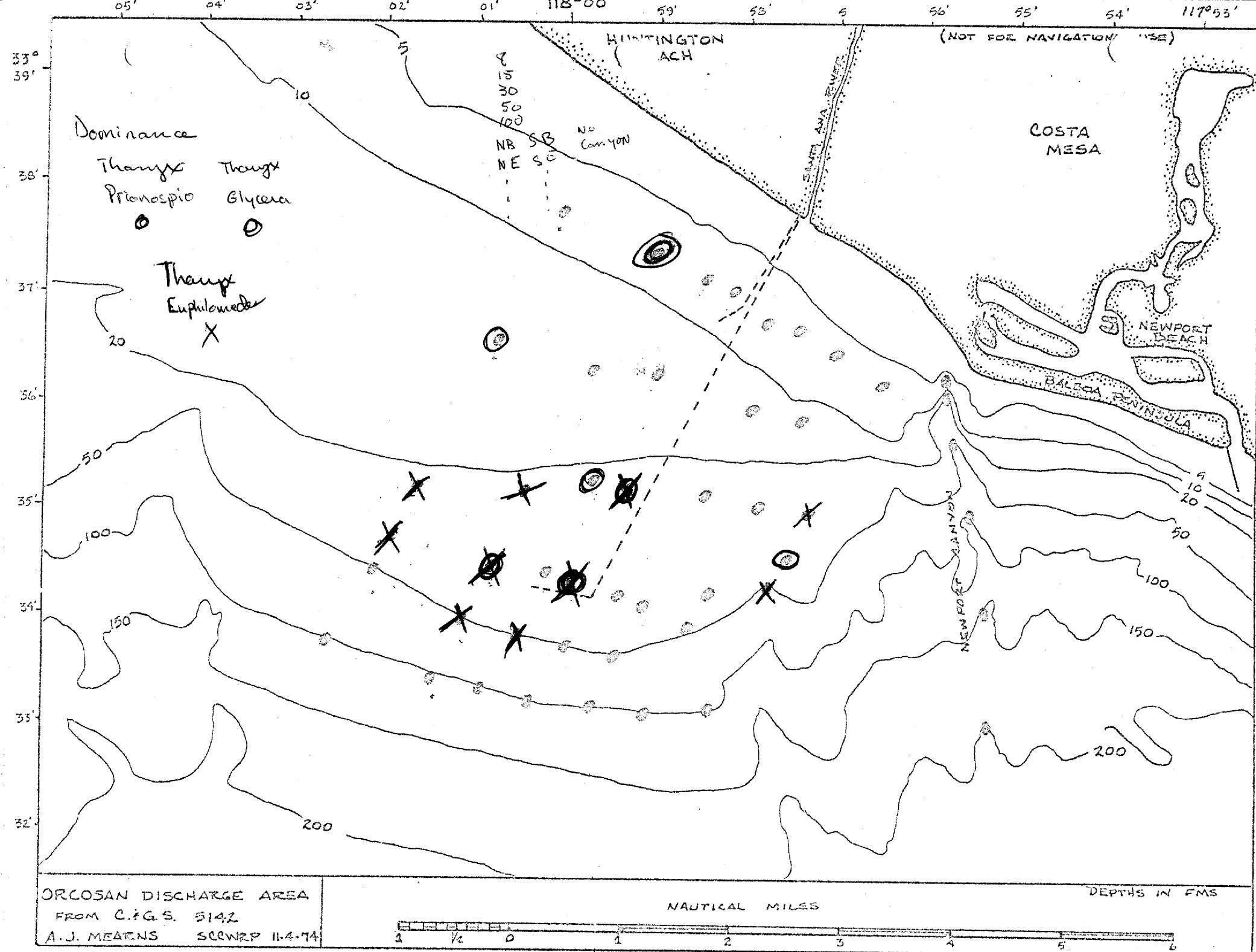
NAUTICAL MILES

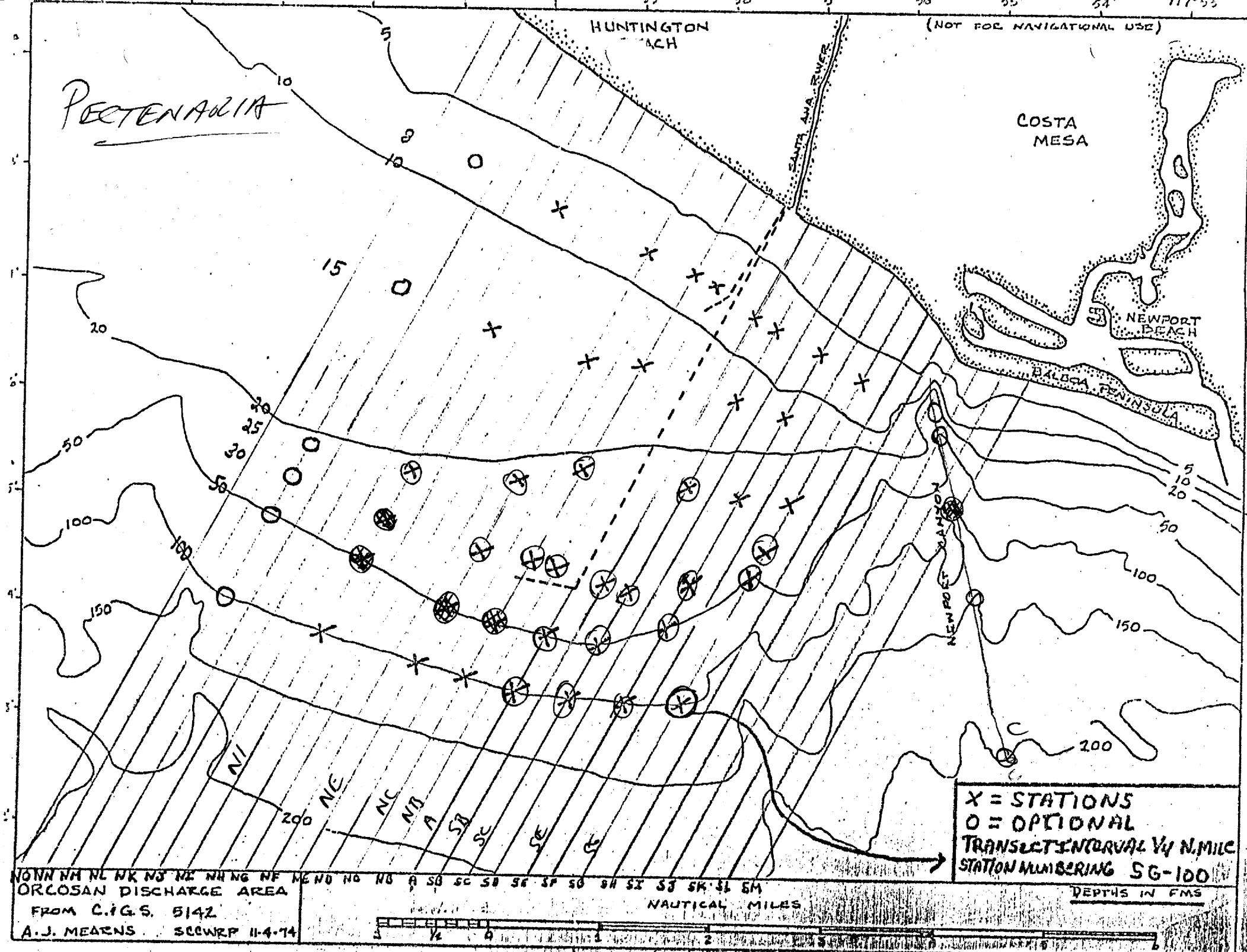
DEPTHS IN FMS

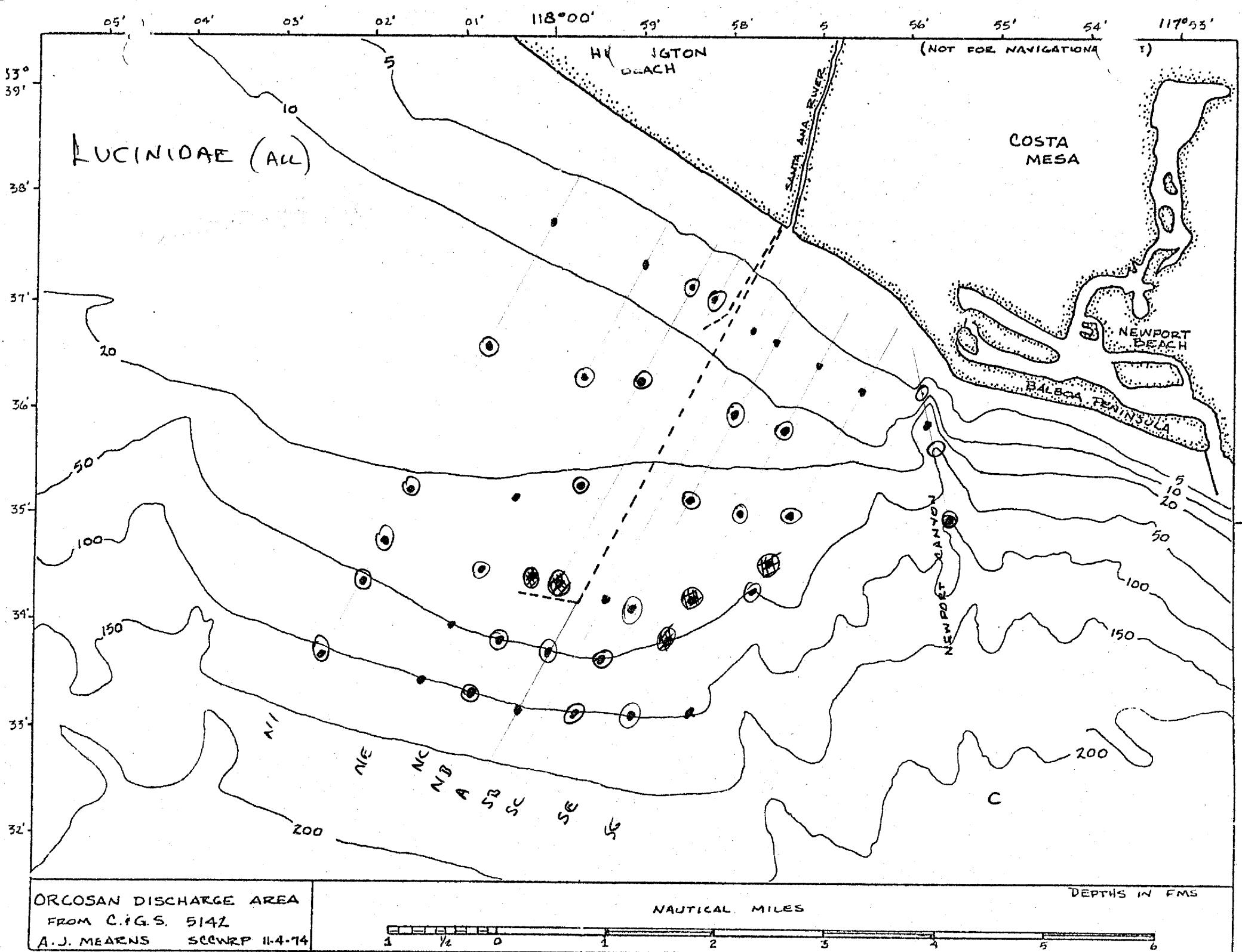






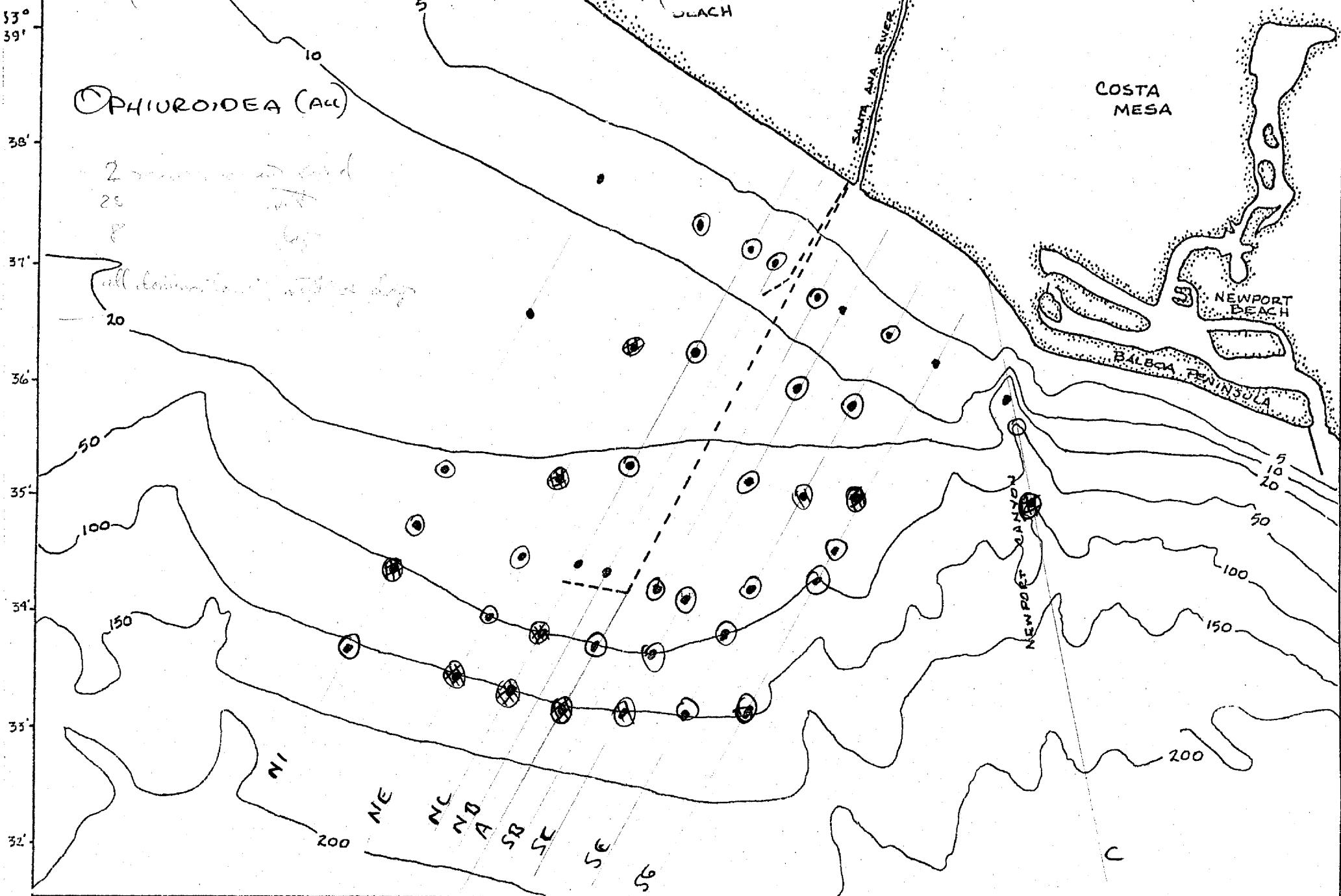






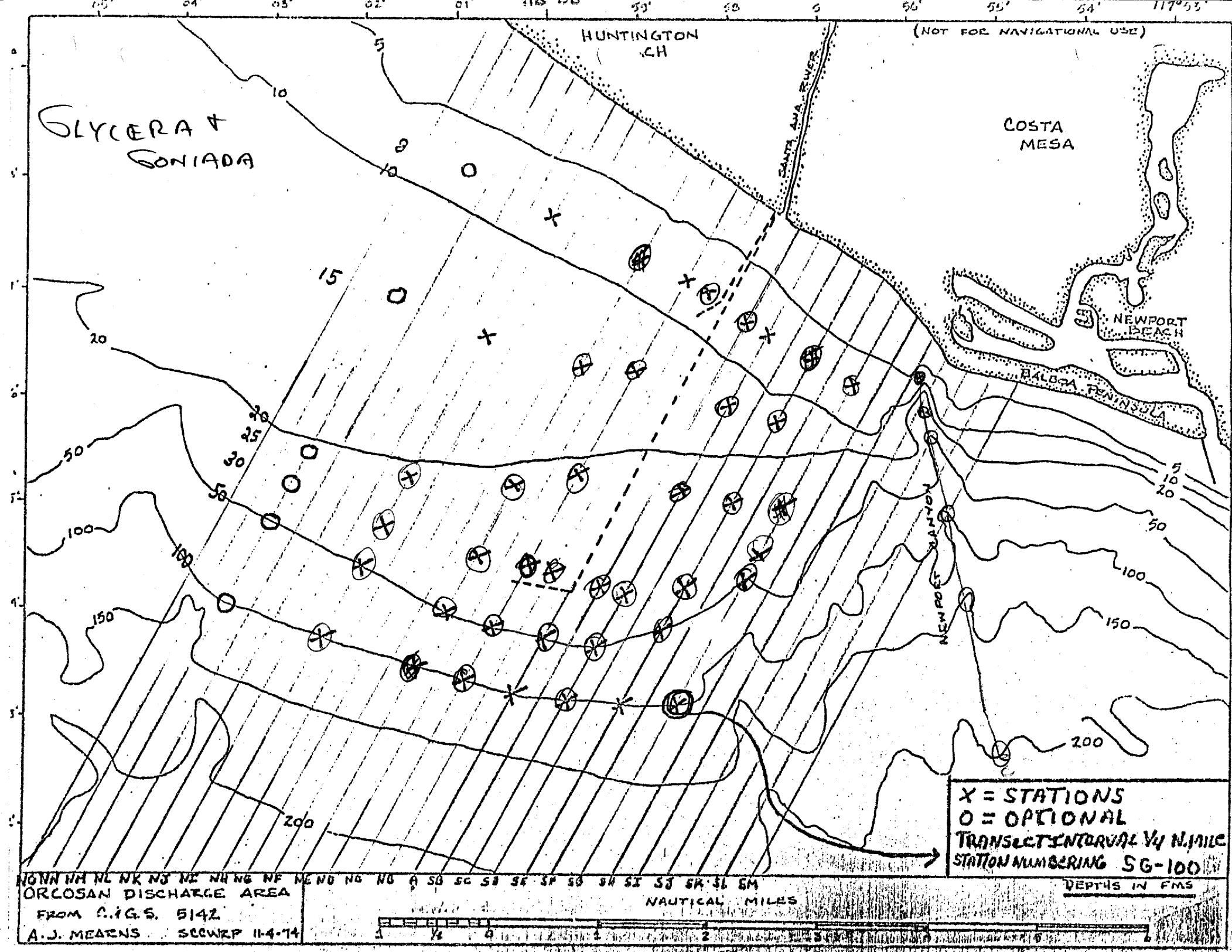
05' 04' 03' 02' 01' 118°00' 59' 58' 5 56' 55' 54' 117°53'

(NOT FOR NAVIGATION)



ORCOSAN DISCHARGE AREA
FROM C.I.G.S. 5142
A.J. MEARNS SCCWRP 11-4-74

1 1/2 0 1 2 3 4 5 6



05' 04' 03' 02' 01' 118°00' 59' 58' 5 56' 55' 54' 117°53'

53°
39'

38'

37'

36'

35'

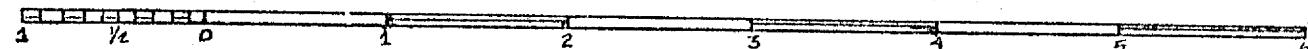
34'

33'

32'

NAUTICAL MILES

DEPTHS IN FMS



ORCOSAN DISCHARGE AREA
FROM C.I.G.S. 5142
A.J. MEARNS SCCWRP 11-4-74

HY JGTON BEACH (NOT FOR NAVIGATION)

EDWAROSIA

generally appear near shore
except south of pipe
why?

COSTA
MESA

NEWPORT
BEACH

BALBOA
PENINSULA

5
10
20
50

100
150
50

100
150

200

200

150

100

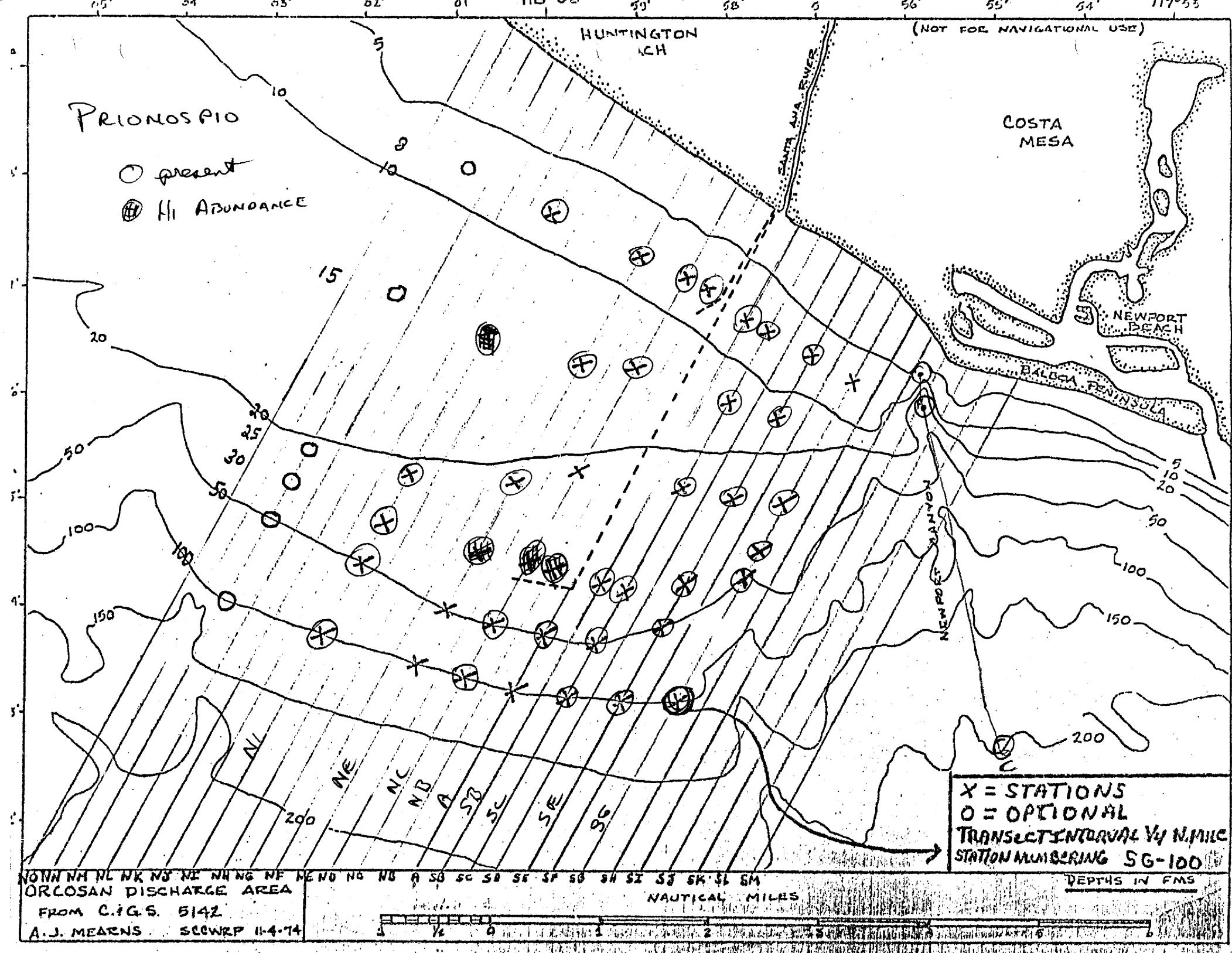
50

20

10

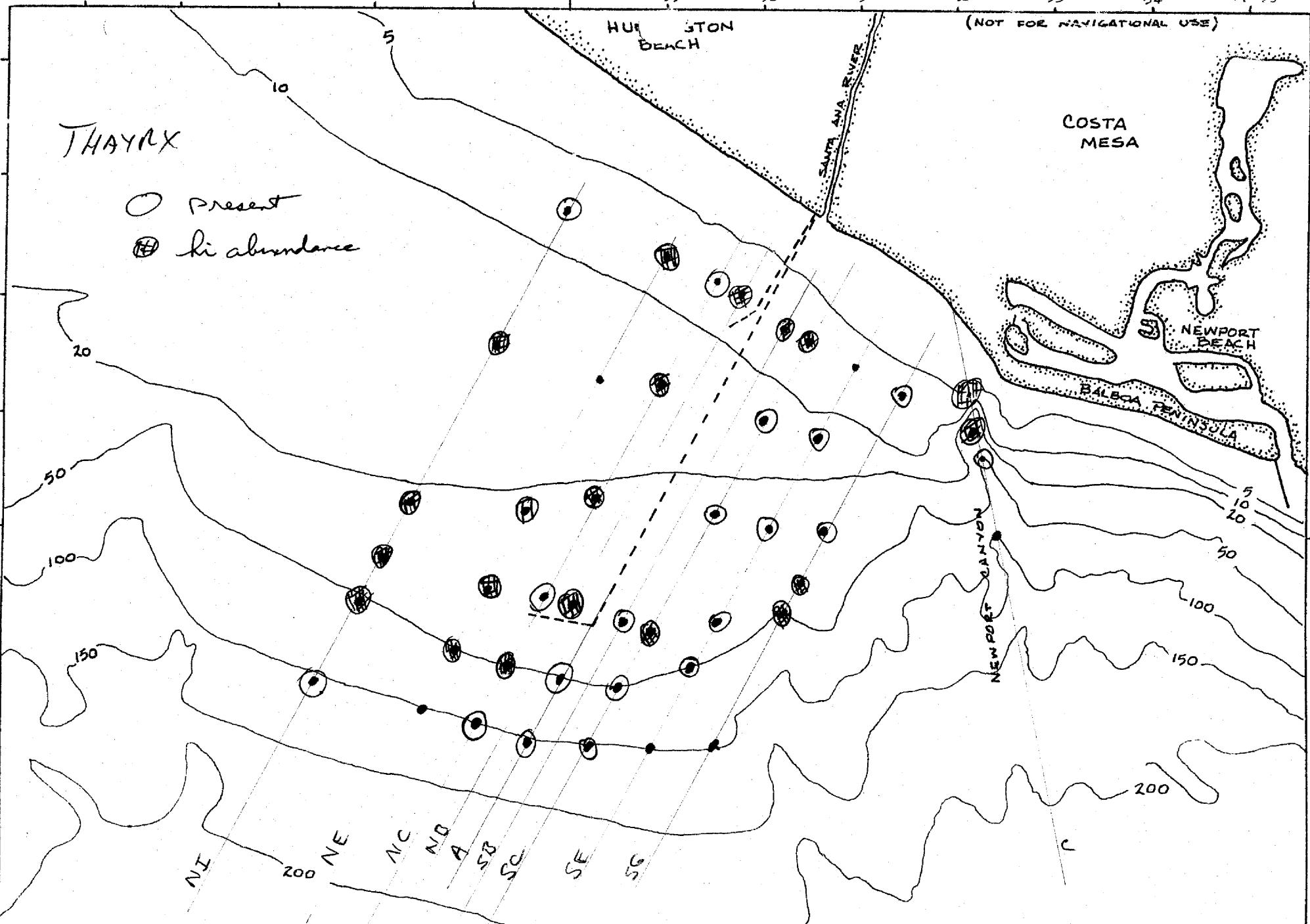
5

(NOT FOR NAVIGATIONAL USE)



05' 04' 03' 02' 01' 118°00' 59' 58' 5 56' 55' 54' 117°53'

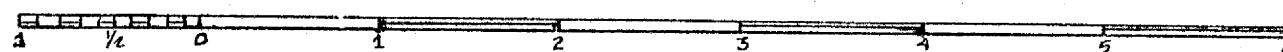
(NOT FOR NAVIGATIONAL USE)

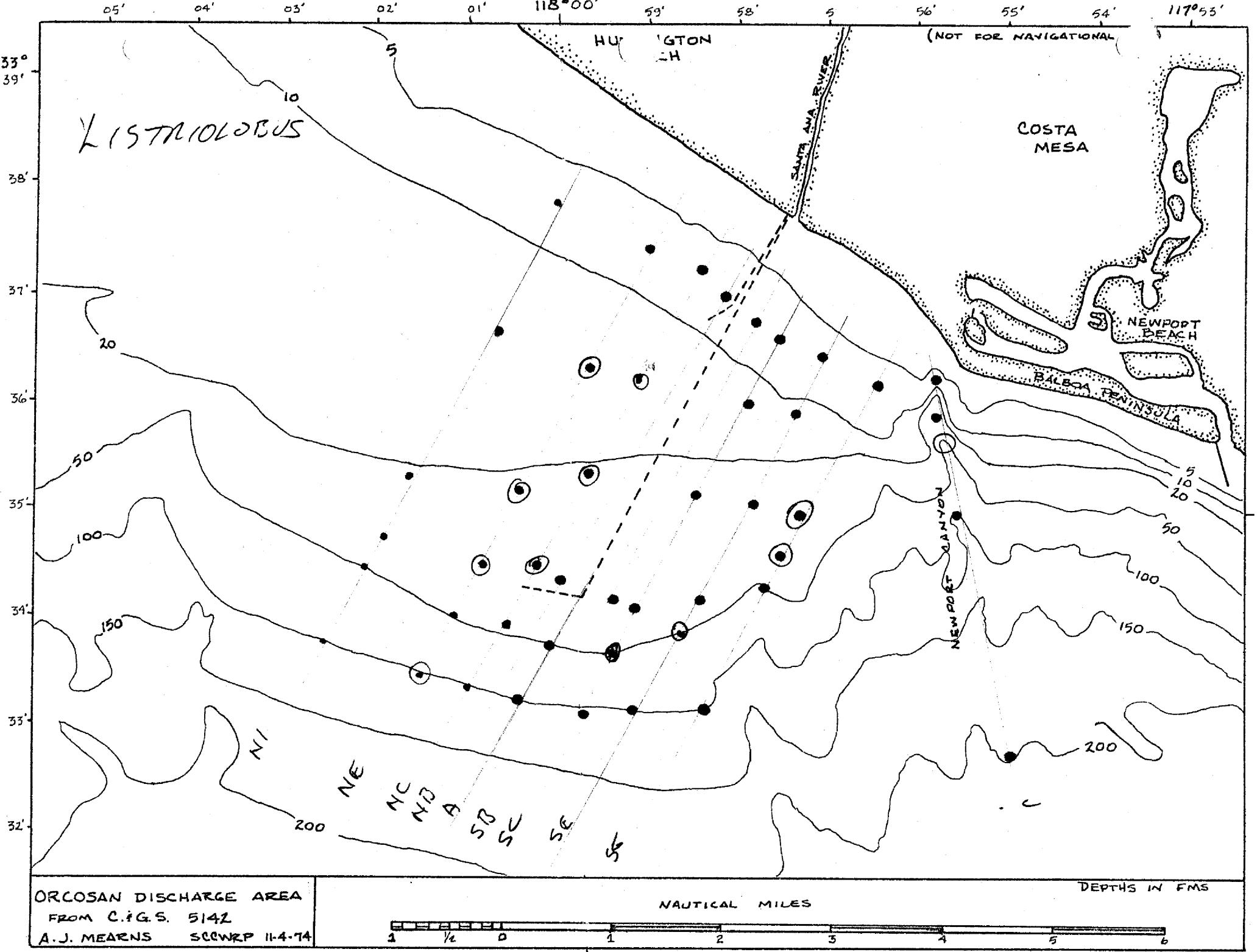


ORCOSAN DISCHARGE AREA
FROM C.I.G.S. 5142
A.J. MEARNS SCCWRP 11-4-74

NAUTICAL MILES

DEPTHS IN FMS





05' 04' 03' 02' 01' 118°00' 59' 58' 5 56' 55' 54' 117°53'

(NOT FOR NAVIGATION)

NUCULANIDAE

HYDROGRAPHIC
ANCHOR

COSTA
MESA

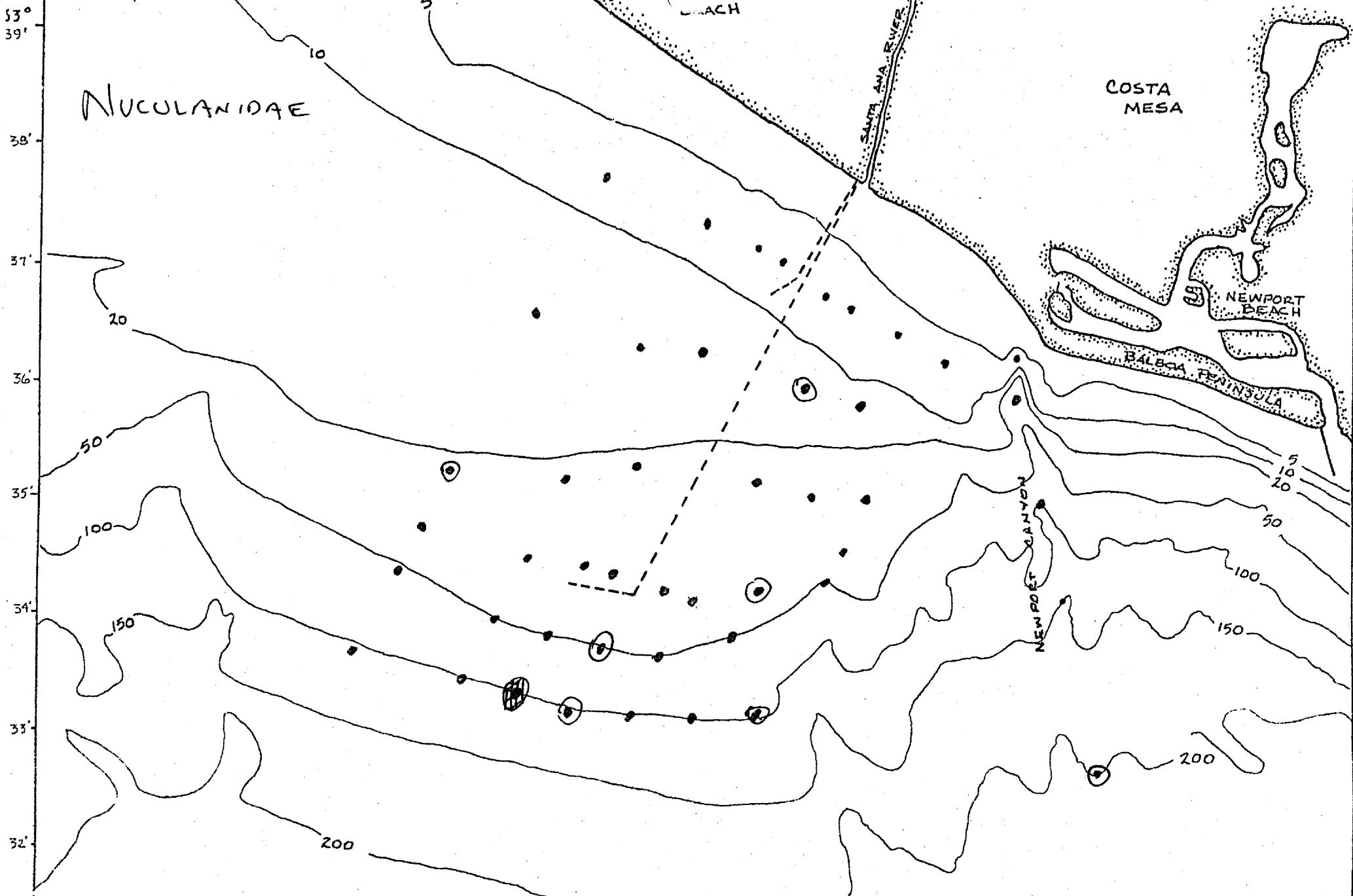
NEWPORT
BEACH

BALBOA
PENINSULA

5 10 20 50 100 150 200

5 10 20 50 100 150 200

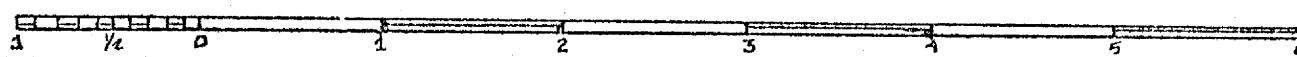
5 10 20 50 100 150 200

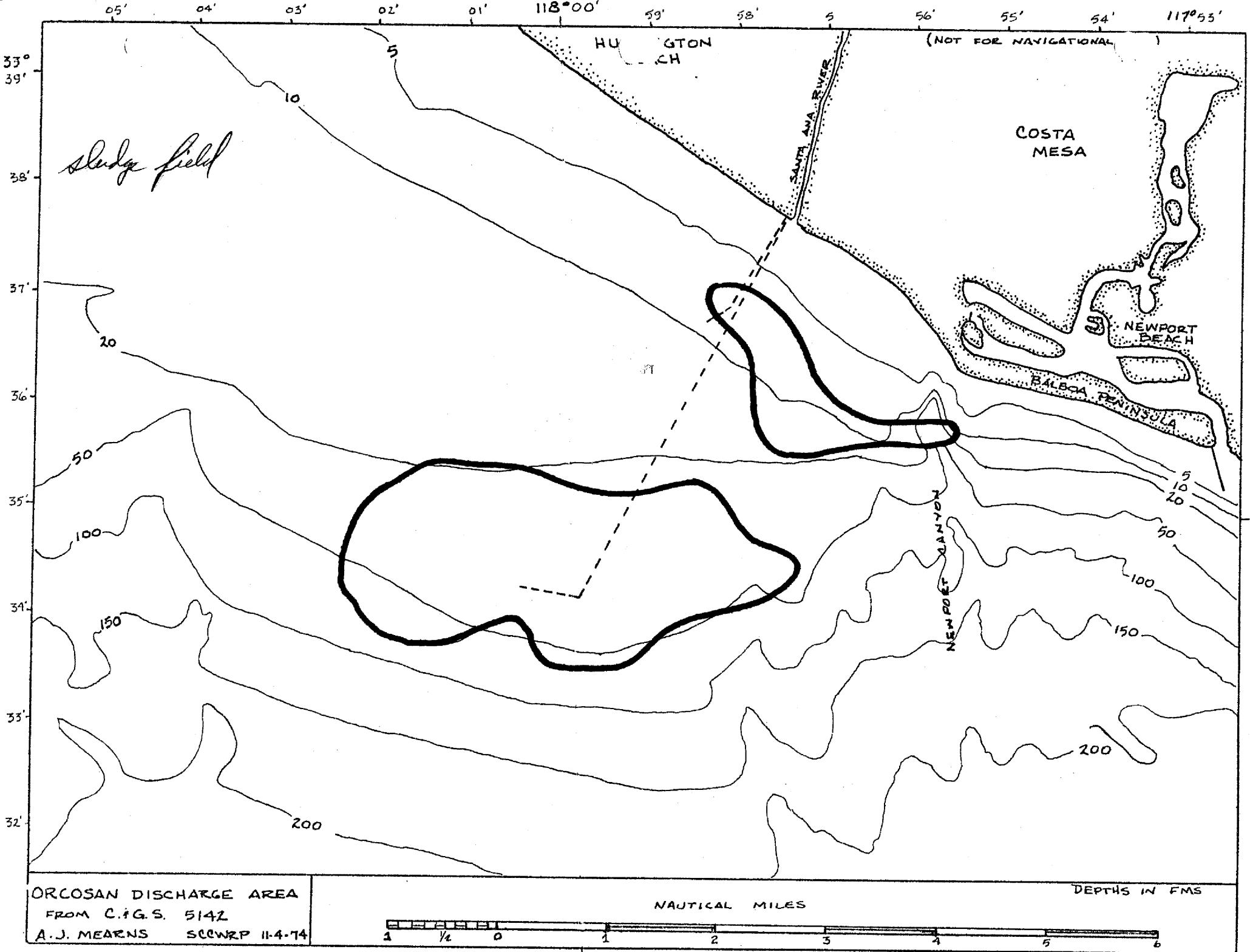


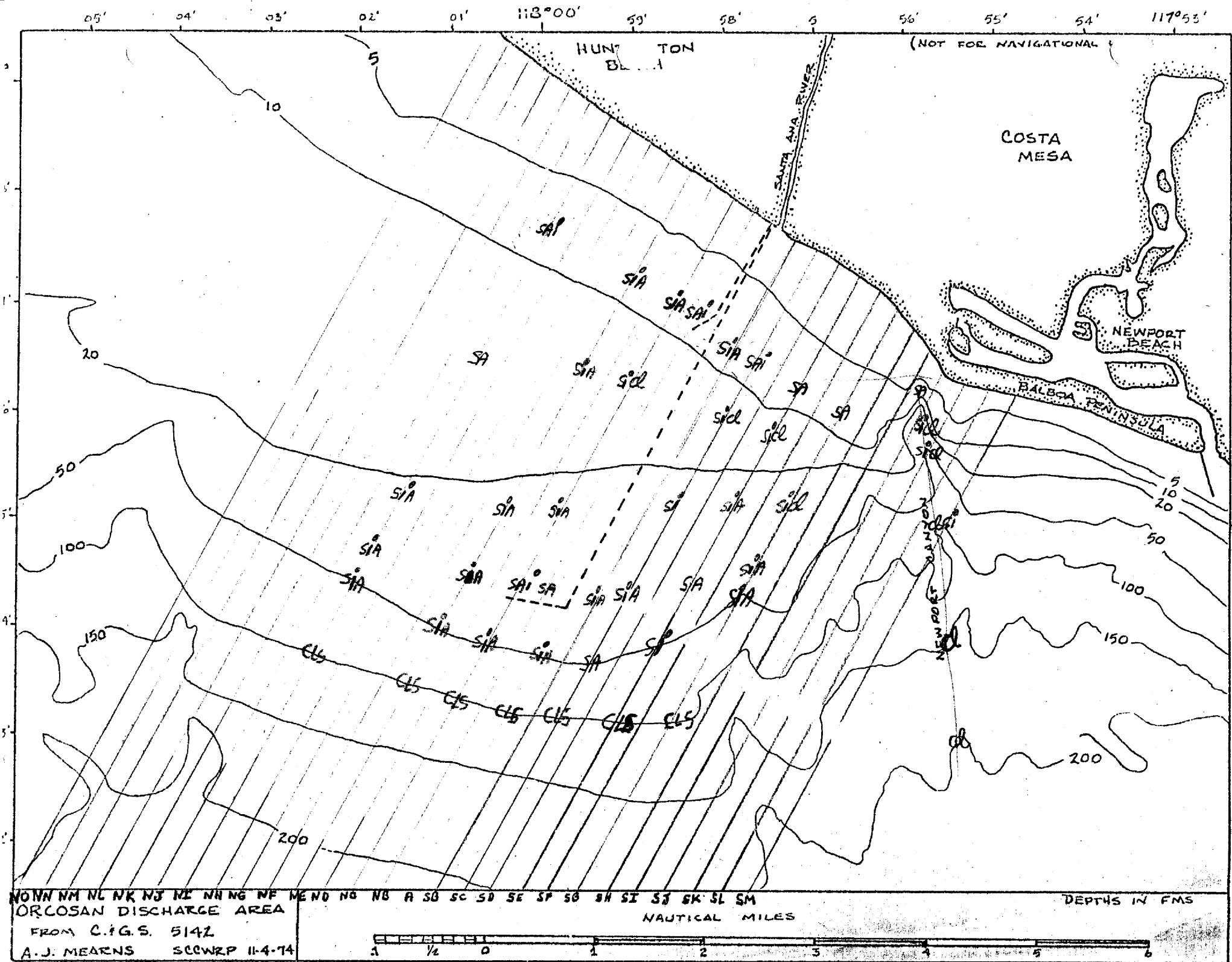
ORCOSAN DISCHARGE AREA
FROM C.I.G.S. 5142
A.J. MEARN'S SCCWRP 11-4-74

NAUTICAL MILES

DEPTHS IN FMS







CLAY

CL>SI

SI>CL

SILT

SI>SA

SA>SI

SAND

5 10 15 20 25 30 35 40 45 50

60

100

FATHOMS

STATION	8 Fm	15 Fm	25 Fm	30 Fm	50 Fm	100 Fm	150 Fm	200 Fm	STATION
Sg-8	SA (1.5cm)		Si → CL (3.5cm)						Sg-8
Sg-25				Si → SA (5cm)					Sg-25
Sg-30					SP → SA (1.5cm)				Sg-30
S-50						CL → Si (10cm)			Sg-50
S-100	SA (1cm)								Sg-100
SE-8	SA (1cm)	Si → CL (2cm)							SE-8
SE-15		Si → SA (4.5cm)							SE-15
SE-25			Si (0.75)						SE-25
SE-30				Si (0.5)					SE-30
SE-50					CL → SP (7.5)				SE-50
SE-100	SA → Si (1.5)								SE-100
SC-8		Si' → CL (6)							SC-8
SC-15			Si (2.0)						SC-15
SC-25				Si → SA (2.5)					SC-25
SC-30					SP (1.5)				SC-30
SC-50						CL → Si (6.5)			SC-50
SC-100	Si → SA (6)								SC-100
SB-8			Si → SA (1.5)						SB-8
SB-30				Si → SA (1.1)					SB-30
A-50					CL → Si (5)				A-50
-100	SA → SL (1cm)								A-100
NB-5			SA (1)						NB-5
NB-30									NB-30
NC-8	Si → SA (3.5)	Si → CL (5.5)		SA → SL (2)					NC-8
NC-15			Si → SA (3)		Si → SA (4cm)				NC-15
NC-25				SA → SL (2)					NC-25
NC-30					CL → SP (11.5)				NC-30
NC-50									NC-50
NC-100	Si → SA (1.5)								NC-100
NE-8		Si → SP (3)		Si → SP (4)		Si → SA (3)			NE-8
NE-15			Si → SP (3.5)				CL → Si (11)		NE-15
NE-25				Si → SP (4)					NE-25
NE-30									NE-30
NE-50									NE-50
NE-100	SA → Si (1.5)								NE-100
NI-8		SA (1cm)	Si → SA (2.5)	Si → SA (3.5)	Si → SA (3)				NI-8
NI-15									NI-15
NI-25									NI-25
NI-30									NI-30
NI-50									NI-50
NI-100						C4 → Si (11.5)			NI-100

(Xcm) = VOLUME 48.5 X 36 X (Xcm)

CL = CLAY
Si = SILT
SA = SAND

6
12.0
 (18×3)
11.0
10.5
✓
10.0
0
L
9.0
U
8.5
M
8.0
N
7.5
7.0
D
6.5
e
6.0
P
5.5
t
5.0
H
4.5
4.0
(Cm)
3.5
3.0
2.5
2.0
1.5
1.0
0.5

5 10 15 20 25 30 35 40 45 50

100

