



How To Know

MARINE ISOPOD CRUSTACEANS

George A. Schultz

Hampton, New Jersey

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INTRODUCTION

This book was undertaken as an attempt to bring together for the amateur and the professional what is known of the systematics and distribution of marine isopod crustaceans from North America. There are 174 genera and 444 species recorded in the waters off the coasts of North America, and they are with few exceptions all included here. The book presents a key to all the genera and most species from North America. No such attempt has been made since 1905, or over 60 years ago, when the first "Monograph on the Isopods of North America" was made by Richardson. An attempt here is made to include all species found in North America in the book. Of course this is almost impossible in a book of this kind because some species are ill defined, others are not always made distinct from related species and certain species names are invalid. To straighten out the taxonomic tangles involved in such an assortment of species is difficult in itself so the book contains a treatment of the species as they are now defined and only the most obvious mistakes of the past are corrected. This should not reduce the usefulness of the book since it does provide a place to begin solving such problems.

An illustration of each species included is present with only a few exceptions. Species in very early times in the history of the study of animals were much more broadly defined than today's species are defined so when specimens are being compared to the illustrations discrepancies may be found. Also the places and depths at which specimens were captured were not as definite as is necessary for really good identifications, so range and depth differences must be revised by more work. Many of the old records are accurate and helpful today even though they are limited and without detail.

The taxonomic arrangement of species in the book is based on the latest available revisions in the literature. The list of genera included in the work can be considered a checklist of those genera found in North American waters, but the species list might not be complete. A known genus that is not included in the present work was perhaps found to be a synonym or not to be really present in North American waters, because of the work of later we revers. Many authors of genera during the first years of the study of isopods had

^{*}See Reference of Menzies, R. J. and P. Glynn, p. 348.

a broad idea of what constituted a genus, and today one finds several genera under what was one old genus. Many New World species were considered to be in broadly defined Old World genera, and through revision by later workers they were placed in new exclusively New World genera.

The book contains what is essentially a key to the genera of marine isopods. The characters on which the genera were defined are used wherever possible in the key so that all species in a given genus will key out to that genus and thusly be separated from other genera. The generic key then should be valuable for other geographic regions aside from North America. The species for the most part have been only found in North America, and must be keyed out on the species characters that they have so there is only local significance to the species part of the key. It must be remembered that a species recorded from a certain place is likely to be found in the same area. It also, however, can be found north or south of the original location or anywhere that is similar to the original location in ecology even if the place is at a distance from the original location. Since a species usually can be expected to be found in or near the area where it has been found before, location can be a useful aid for the identification of a species. However, it is always necessary to consult some work which contains a morphological discussion of the species in question for a positive identification

The author's wish was to construct a book useful for both the amateur and professional marine biologist and invertebrate ecologist. Most of the literature since 1905 on isopods was reviewed, and the newly described species and the range extensions and some new biological data were recorded and are presented here. Some species descriptions or records were perhaps overlooked by myself so it is necessary for any serious student to search the literature himself before concluding that everything is known about the species in the group that he may have under study.

There are mistakes in this book for which the author makes no apology. They are the normal mistakes and oversights of other authors that have been repeated here since the book is not intended to be a complete revision of marine isopod crustacean systematics. They will be pointed out wherever possible. There are, however, other mistakes that were points overlooked by the author, for which he is willing to take full blame. However, the book should be very useful in spite of these limitations. The work was done using the library facilities of the American Museum of Natural History and the United States National Museum.

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HOW IS AN ISOPOD DIFFERENT FROM OTHER ARTHROPODS AND CRUSTACEANS?

The phylum Arthropoda contains the largest number of species of any phylum in the animal kingdom. Arthropods include all animals that have a hard exoskeleton and jointed appendages. Insects and spiders are common arthropods and represent two classes of that phylum. The crabs and shrimp are also common arthropods; however, they are not in the same subgroups as insects or spiders. They form a group of equal taxonomic rank to the spiders and insects, and are in the class Crustacea along with many other subgroups of Crustacea including isopods. In order to place an isopod crustacean in the proper place in the phylogenetic tree of the animal kingdom, it is a good idea to briefly review the relationship between the familiar arthropods that are closely related to the crustaceans then review the crustaceans that are closely related to the isopods. The relationship shown in Fig. 1 is discussed briefly here.

Insects make up the largest number of arthropod species and they are found on land. The most primitive arthropods were the trilobites which today are known only as fossils. The trilobites, one of the earliest known inhabitants of the earth, inhabited the ancient

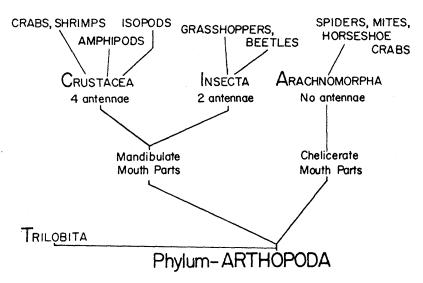


Figure 1. Evolution of the Phylum Arthropoda.

ocean bottom in deep and moderately deep water. The present day crustaceans evolved from the trilobites (Fig. 2) or from an ances-

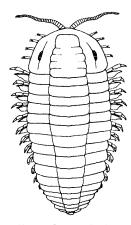


Figure 2. A trilobite, found only as a fossil. There is very little differentiation of the cephalic appendages towards being mouth parts.

tor that was very much like the trilobites in appearance. The crustacean species, which for the most part are inhabitants of the sea. also constitute a most primitive subgroup of arthropods. The spiders and insects are found almost exclusively on land today, but they also evolved from primitive marine ancestors. The crustaceans, however, for the most part remained in the sea, and many species evolved in the various ocean habitats just as many species of spiders and insects evolved in the various terrestrial habitats. Most crustaceans have remained in the sea. but there are freshwater forms apparently derived from marine species found in the near shore habitats, and which invaded the brackish water of river mouths and evolved in the habitats in rivers and lakes where they are found today.

There is a terrestrial group of isopods, probably the most successful terrestrial crustaceans other than land crabs and some tropical amphipods, that has invaded the land. They form the suborder Oniscoidea. Terrestrial isopods have become well adapted to land where they have evolved a hard, calcareous exoskeleton which protects the isopod from the drying power of air. They will be the subject, along with the freshwater species, of another "Picture-Key Nature Series" book. The crustaceans have evolved calcareous exoskeletons; whereas, the insects and spiders have evolved chitinized coverings with a wax coating that protects them from desiccation.

The spiders are more widely different from the insects than are the crustaceans. Spiders and their relatives have the cheliceral appendages of the mouth parts well developed instead of the mandibular mouth parts (Fig. 3). The chelicerae are usually developed as piercing structures, and the mouth parts as a whole are modified for piercing and sucking. In most crustaceans and insects the mandibles are well developed, and are generally modified for biting, so that even if it were not for the other obvious differences, the isopods would not be confused with the insects.

The crustaceans and insects each have specialized sensory appendages, the antennae, which are lacking in the spiders. The insects have only one pair of antennae and the crustaceans have two pairs. The type or principal mouth parts of the animals and the absence or presence of antennae are not the only differences that

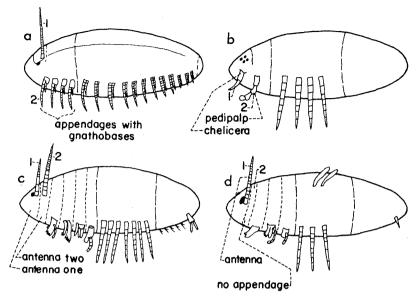


Figure 3. The cephalization of some Arthropoda compared. a. Trilobita. b. Arachnomorpha. c. Isopoda. d. Insecta.

set the spiders apart from the insects and crustaceans, nor are the presence of one pair of antennae only in the insects and two pairs in crustaceans the only difference between the groups. These differences and similarities are what are called key differences of key characters—differences that can be used to "key out" or identify the different groups. More will be said about key characters and other characters later (p. 41).

HOW IS AN ISOPOD DIFFERENT FROM OTHER CRUSTACEA?

There are many different kinds of crustaceans and isopods are only one large group of a large number of groups of crustaceans. The isopods are generally considered to be in the subclass Malacostraca, or the higher crustaceans, rather than in the lower crustaceans (several different subclasses). The Malacostraca are supposed to be composed of animals like the crabs and shrimps and isopods that are phylogenetically more advanced (they exploit more habitats and have more defined species) than the lower crustaceans. The crabs and shrimps (the Decapoda—key character 10 legs) are considered phylogenetically the highest developed superorder of Malacostraca—the Eucarida. The other superorders of

Malacostraca that are most closely related to the Isopoda are the Syncarida, the Pancarida and the Peracarida which are generally considered to be related according to the phylogenetic arrangement shown in Fig. 4.

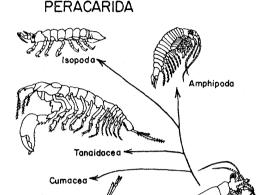


Figure 4. Evolution of the Malacostraca (After Siewing).

Only the Peracarida will be discussed briefly here because the other groups are only indirectly related to the isopods, each being on its own branch of the phylogenetic tree. The Peracarida, the superorder that contains the order Isopoda, is characterized by animals that have no carapace or only a small carapace. They have oostegites which are structures which enclose the cavity in which the eggs are laid and which form a brood pouch in the female where the young are hatched and reared until they are able to fend for themselves. The animals with the characteristic peracaridal characters are divided into five orders including the Isopoda, Mysidacea, Cumacea, Amphipoda and Tanaidacea. The isopods are considered to be phylogenetically the most advanced order of Peracarida. The phylogenetic relationship within the superorder is shown in Fig. 5 which also contains an illustration of a species of each order.

The animals that have been most frequently confused with the isopods are the amphipods which are frequently caught in the same habitats as isopods. However, the body of most amphipods is laterally compressed and the appendages are quite different. Amphipods are easily told from isopods in the fields because they normally have a smooth body that is curved as well as being later-

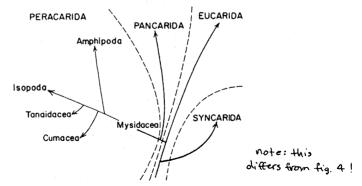


Figure 5. Evolution of the superorder Persesrids.
Malacostraca

ally compressed. Amphipods frequently hop and have a characteristic swimming motion which is rarely associated with isopods. Other animals less frequently encountered with isopods are the almost exclusively planktonic marine mysids (comparatively long animals with a shrimp-like appearance and large stalked eyes), the bottom dwelling cumaceans (comparatively rare animals, in the near-shore habitats at least, which have a characteristic appearance, the anterior half being much longer than the thin tail-like posterior half) and the tanaids. The tanaids resemble the isopods so closely that they were formerly classified in the same order, and it was not long ago that they were separated and placed in a separate order near the isopods.

The order Tanaidacea which contains the tanaids is characterized by species which have the cephalon or head and the first thoracic or peraeonal appendage fused to form a small carapace. The species have a true chelate first appendage and multiarticulate uropodal rami (Fig. 6). These characters are never found in

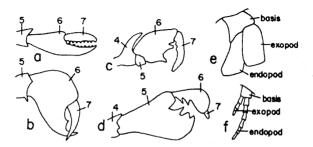


Figure 6. Types of chelipeds and uropods in some crustaceans. a. True chelipeds (Decapoda). b. True chelipeds (Tanaidacea). c, d. Subchelate peraeopods I (Isopoda). e. Uropods with rami of single segment (Isopoda), f. Uropods with rami multiarticulate (Tanaidacea). (See Fig. 29 also).

isopods. There are other differences, but these three if kept in mind will serve to separate the most closely related group, the tanaids, from the isopods. The tanaids are very abundant in near-shore habitats and are also found on the bottom in the deep-sea. They live, for the most part, in tubes and burrows and are many times

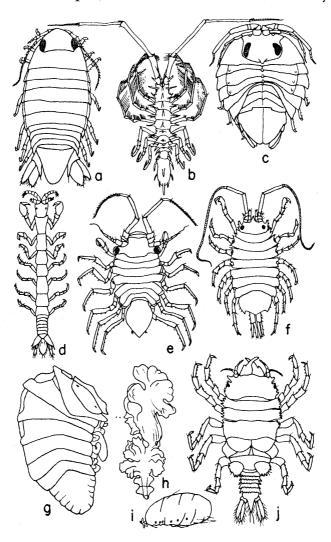


Figure 7. Examples of Isopods. a. Cirolana (Flabellifera), b. Arcturus (Valvifera). c. Serolis (Flabellifera). d. Anthurid (Anthuridea). e. Munna (Asellota). f. Janiralata (Asellota). g. Bopyrina (Epicaridea). h. Cancrion (Epicaridea). i. Faba (Epicaridea). j. Gnathia (Gnathiidea).

abundant in areas occupied by many different species of isopods, especially the anthurids. At times they are known to share the same burrows, and for the most part their general ecological preferences are known to be similar to those of isopods. It is no wonder that they were formerly classified with the isopods.

As is immediately apparent from a look at Fig. 7, the isopods are quite different from one another in general appearance although close inspection will show that they all have the same three body tagmata—the cephalon, the peraeon and the pleon. The three divisions are not apparent in the highly modified endoparasites, the entoniscids, and the highly modified cryptoniscids (Fig. 7h, i), but for the entoniscids the body is divisible into three body regions (Fig. 559). The tagmata are quite different in appearance in the different species, but they each have a very similar basic structure. Fig. 8 contains a picture of an archetype or primitive hypotheti-

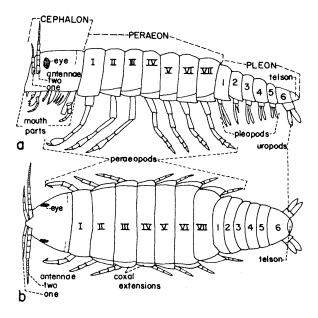


Figure 8. The archetype isopod with nomenclature of parts, b. Lateral view. b. Dorsal view.

cal isopod. It most nearly resembles a cirolanid, not a present day cirolanid, but what an early or primitive one might have looked like. The figure is a generalization or an extrapolation of what the ancient isopod might have looked like arrived at by determining the most primitive structures or forms of structures of many present

day isopod groups.

The cephalon, or head, of the ancient species was apparently composed of fused segments. On trilobites and other primitive lower crustaceans the fusion of the cephalic parts had not taken place yet, and it was only after the fusion took place that the crustacean could begin to be called an isopod. Primitively the seven peraeonal segments with appendages were without any major differences, and the six pleonal segments plus a telson were free and without major differences also. As we shall see later some anthurids and bopyrids still retain many of the primitive features of the archetype species. With the exception of the sixth pleonal segment fused to the telson, the other general features of the archetype corresponds closely to the cirolanid isopod type. Terrestrial species are derived from a cirolanid-like ancestor and resemble the archetype or primitive isopod also. The phylogenetic tree of the isopods is shown in Fig. 9, and it serves to illustrate some fundamental

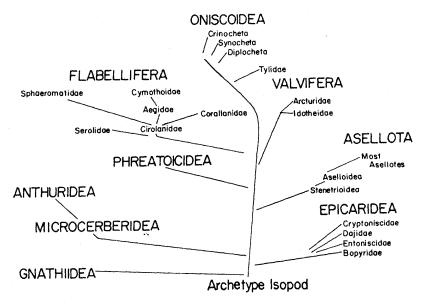


Figure 9. The evolution of the suborders of the order Isopoda (only major subgroups or families are shown).

morphological differences and similarities among major groups of isopods and the evolutionary steps are put in a time dimension. The key to the higher suborders of isopods begins on page 47, and is in part based on the archetype figure.

GENERAL ECOLOGY AND ECONOMIC IMPORTANCE

Marine isopods are of considerable economic importance to both the economy of man and the economy of nature. The role that isopod speices play in their respective habitats is generally that of reducer organism; i. e., they take the dead and dying whole organisms and parts of whole organisms and reduce them to smaller particles (Fig. 10). The isopods as a group are said to be omnivores because they eat both plant and animal matter. There are true herbivores (plant-eaters) as well as true carnivores (animal-

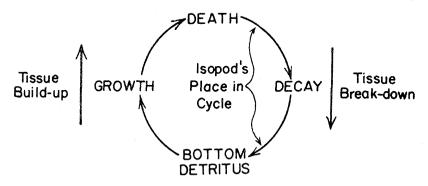


Figure 10. The place of isopods in the general scheme of organic turnover and energy flow in an ecosystem.

eaters) among the isopods, but they are few in number when compared to the total number of omnivorous (plant and animal) species.

The wood-boring species are probably the most economically important isopods to man because of their destruction of wooden marine structures that are placed in the sea and estuarian waters. The structures include wooden boats, wharves and piers and pilings that support bridges. No wooden structure is immune to isopod attack and much money is spent each year to replace the decayed pilings that once supported bridges and other marine structures and were weakened by the borings of the isopods. Much money is also spent on research on how to treat wood to prevent attack by the borers. Trees and shrubs that fall into marine water after storms and other natural causes also are soon subjected to attacks by wood-boring isopods.

The role in nature of the majority of other isopods is not so dramatic, at least not on the face of it, as is that of wood borers. However, in the overall economy of nature their combined role as reducer organisms is much greater than that of all wood borers

combined. The isopod species are in one of the most important and little understood places in the general economy of nature, that of making smaller particles out of dead or decayed plants and animals. The role of many vertebrate organisms is well known and their contribution to the energy cycle of their ecosystems is at least superficially understood. The role of invertebrate organisms (e. g., crabs, insects and isopods) is not as well known however. That isopods are very important animals in the invertebrate ecosystem is an indisputable fact once their number in population and their number in species in the various ecosystems are known. They are among the most important and dominant organisms found on the bottom in deep and very deep parts of the ocean.

Isopod species constitute some of the most important scavengers of the ecosystem. Dead fish in the intertidal zone and in any part of the ocean for that matter, are soon reduced to small particles. First the particles serve as food for the isopods: then when the isopods excrete the residue, it is utilized by smaller organisms until it is finally reduced to chemicals that are utilized by the plants and which are eaten by animals (sometimes isopods) which eventually die and are eaten by the isopods. A large dead fish can be reduced to a skeleton in a few hours by a population of isopod scavengers. The isopods do not confine their scavenger habits to dead fish, but also eat other dead organisms including other crustaceans and plants. Certain of the isopod species confine their plant eating to particular plants. Some boring isopods confine their activities to particular species of algae. Some large brown algae, for instance, are destroyed because the isopods bore large numbers of holes in their holdfasts and the large algae become free and are washed away from where they originally grew (Fig. 11).

Many isopods eat animal and plant matter that has first been broken down by other animals. This is probably true of most deep sea species. They eat the particulate matter, dead plankton and other matter, which filters down from the open ocean and accumulates in large amounts on the bottom. The bottom mud and sand is strained through either modified mouth parts or the modified an-

terior peraeopods of particular species of isopods.

The mouth parts most frequently modified for straining food are the second maxilla (Fig. 38), although other mouth parts like the maxillipeds in gnathiids (Fig. 348) or the legs of arcturids (Fig. 49) can be so modified. Some species filter sea water and others filter their food from the bottom mud and sand. If the water from the sea is filtered, not only are small organic particles filtered from the water, but live planktonic plants and animals are filtered as well. The plankton is eaten alive in the near shore habitats, but in the deep sea it arrives only after dying or having been trans-

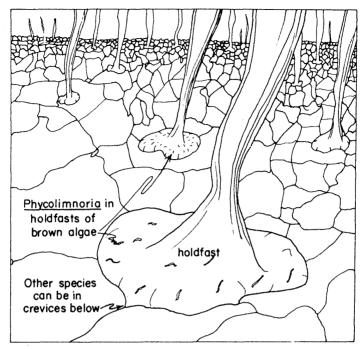


Figure 11, "Forest" of laminarian (brown) algae with large holdfasts penitrated by Limnoria (Phycolimnoria) species,

formed by other organisms into particulate matter which rains down from above and is either eaten directly or mixed with the bottom mud and sand which is then filtered by the isopods. Isopods are of course not the only filter feeders in any environment, but they are one of the most numerous and important groups of filter feeders. Some isopods live on top of the mud and sand (general benthic species), some live in tubes (tubiferous species) and some burrow in the bottom (fossorial species). Some parasitic species are also found on the bottom where they apparently spend time between bottom fish hosts.

Parasitic isopods play their part in influencing the lives of their hosts which include fish and other crustaceans including other isopods. The fish that are parasitized by the isopods include many economically important species. Some bottom fish, haddock, halibut, flounders and skates, are parasitized by one group of isopods. They (e. g., Rocinela spp.) are temporary or non-obligate parasites for the most part. They are found apparently free living on the bottom when they are not found on fish hosts. Apparently they can

easily make a transfer from one host to another since they live on the bottom with the fish hosts.

There is another group of isopod parasites of fish of the nearshore and open sea. The species are found on jacks, tuna and sharks. Their attachment to the host fish is permanent and many times they are found inside the mouth and/or the throat cavity of the host fish (Fig. 12i). Although they can live without being attached to the host, they are rarely found unattached and soon

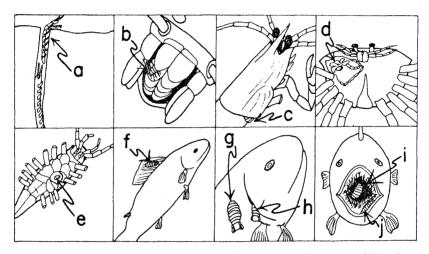


Figure 12. Different types and degrees of parasitism by isopods. a. Living in burrow of another animal. b. Living on other isopod (*lais* species on pleopods of sphaeromatid). c. Bopyrid in branchial cavity of shrimp. d. Entoniscid (a true endoparasite) in body cavity of crab. e. Cryptoniscid in marsupium of another isopod. f. Cirolanid on fin of fish. g. Ectoparasite on skin of fish (Aegid). h. in the gill slit. Parasite in throat of fish. j. Parasite on lip of fish.

reattach themselves if they are detached. The isopods never directly kill their fish host, at least not average or strong hosts, but they probably have some influence on the regulation of numbers of the host fishes.

The isopods that inhabit the body of other crustaceans also apparently influence the numbers of their hosts. However, very little is known about the parasite-host relationship in regard to numbers. Occasionally as many as 50% of some crab populations in a given location are found to be parasitized by isopods, but it is rare to find the parasites in such a large percentage and they are usually found in less than 5% of the host population. The isopods are found as ectoparasites as well as endoparasites of the other crustaceans. Isopods with little body modification are found attached to the body of their crustacean hosts. If the isopods are found within the

body of the host, mainly the internal cavity of crabs (haemocoel), they are greatly modified (Fig. 12d). The external, branchial cavity parasites, are usually indicated by a bulge or distortion of the body of the host (Fig. 12c). The endoparasites rarely distort the body of the host and are usually only found to be present by the presence of a small hole leading to the body cavity of the crab.

An interesting aspect of the isopod parasites is that they are able to affect the sex, and sexual development of their crustacean hosts. A host is of one sex and it can be changed to the opposite sex by the presence of an isopod parasite. Often the complete sexual identity of the host crustacean is destroyed when parasitized by an isopod. This is called parasitic castration. There is also evidence that the sex of the juvenile is influenced according to whether or not it remains on the host and also by the sex of other isopod parasites on the same host. The chemical or hormonal processes involved are not well understood and the subject must be studied further.

Since isopods which are parasites on other crustaceans are much more modified morphologically than isopods that parasitize fish. they are most probably phylogenetically much older than the parasites of fish. In some groups like the gnathiids only the juveniles are fish parasites. Some isopods are parasites on other isopods. Certain species are found exclusively within the burrows of other species and are apparently commensal with them (Fig. 12a). Commensal means that they share the food of the host in one way or another. Other species are found exclusively among the pleopods of larger isopods. Iais californica is thought to live exclusively among the pleopods of Sphaeroma pentodon, for instance (Fig. 12b). It almost always is found in its burrows if not on the sphaeromatid itself. Another parasitic species is found in the marsupium of the female of a free living isopod species (Fig. 12e). At first glance it looks just like another egg, but on closer examination it is an egg mass itself or really a female parasitic isopod sometimes with a male attached. Other species are attached to the host and appear to be only an egg mass (Fig. 548).

GEOGRAPHIC DISTRIBUTION OF ISOPODS AND SOME ASPECTS OF THE HISTORY OF THEIR STUDY

The geographic area covered by this book on marine isopods includes the waters around the continent of North America beginning in the north with the Arctic Sea and extending to Panama (Map, Fig. 13). The eastern boundary is Greenland and the western most of the Aleutian Islands forms the western boundary. No

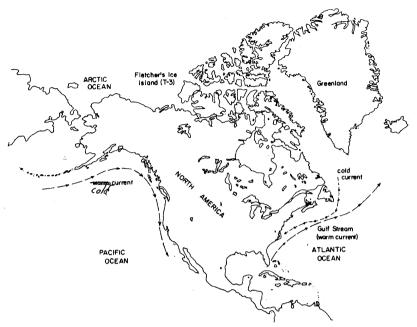


Figure 13. Map of region included in the book. Some ocean currents that influence isopod distribution are shown.

Hawaiian or open ocean species near Hawaii are included, but some species found far from the coast of western United States, Mexico and Central America are included. Species are recorded that were collected in the Arctic Sea from the ice island T-3 or Fletcher's Island, and some were collected along the northern coast of Alaska. Specimens have been collected north of Thule, Greenland, in the body of water that separates that island and North America proper. Other specimens were taken in the deep sea at about the same latitude as the southern tip of Greenland.

Specimens have been recorded from the Atlantic Ocean off North America from relatively long ago, but one of the first series of collections was made about the turn of the century by the steamer Albatross. The voyage of the Albatross included taking some specimens from the Gulf and Caribbean region. Other expeditions made collections in Bermuda, and some recent collections were made in southeastern United States.

The Albatross, in several voyages, sailed from northern United States through the newly opened Panama Canal along the Pacific coast to Alaska and the Aleutian Islands. The early collections made on the Pacific coast are thus as extensive in far from shore species as are those on the Atlantic coast. The collections were extensive

in the Aleutian Islands and the Bering Sea since the *Albatross* expedition to the region was made after the purchase of Alaska as the first floral and faunal study.

Most early collections made in colonial times in North America were sent to Europe and were described by the European scientists. Several of the earliest collections made in the United States were made on the New Iersey coast at Egg Harbon, by the American naturalist Thomas Say in the early 19th century (1818). He was associated with the Academy of Natural Sciences founded in part by Benjamin Franklin in Philadelphia. The more recent near shore collection locations are found near the marine laboratories that were established. The marine laboratory established in the late 19th century at Woods Hole on the southern Massachusetts coast was the collection location for many species of isopods. Extensive work was also done at the Hopkins Marine Laboratory in Monterey Bay in the late 1800's and early 1900's. More work was done at Scripps Institute for Oceanography at La Jolla, and by the Allan Hancock Foundation, both in southern California, Other individuals made extensive collections from the near shore habitats on the midand southern California coasts in more recent times.

Many near-shore species were described from the Pacific coast and the number exceeds that of the species described from the Atlantic coast. Few collections were made in the Gulf and Caribbean regions of eastern North America. Much work remains to be done on tropical isopod distribution. The number of species recorded on the east coast of North America perhaps will exceed the number from the west coast when the eastern tropical region is studied. The southern boundary of the region covered in this book is Panama where some collections were made at about the time that the canal was opened. It extends east to Trinidad where only a few collections were ever made. Some northern South American forms, namely some from Guyana (British Guiana), are included because they are of some interest since they will probably be found in other parts of the West Indies region.

The geographic habitats where collections were made in addition to being divided into eastern and western coasts are roughly divisible into marshes, brackish water estuaries, near shore intertidal and shallow water areas, continental shelf and deep sea. There is no rigid distinction made between the regions because the people who made many of the early collections did not always make a very clear reference to the places and the depths at which their specimens were collected. More about the specific ecology of where species were found is included in the "Where to Look for Marine Isopods" section.

An interesting aspect of the distribution of isopods is the way that some marine species are distributed in freshwater habitats. Map (Fig. 13) has "X's" on it where certain members of the family Cirolanidae were found. The species are blind and found in freshwater in caves. If one draws a line connecting the points, what is geologically the boundary of an ancient sea is revealed. The species were apparently found on the edge of the sea as are most cirolanids today. When the sea retreated and the shore line changed, they remained as the inhabitants of caves that soon became freshwater habitats and there they remain today.

One of the most important things to keep in mind when studying the distribution of isopods is the local peculiarities in climate, depth and water currents and other geographic features. They distort what would be an otherwise perfect picture of north-south, low waterdeep water, etc., distributional pattern. Thus isopods, both deep water marine and near shore species are regularly found further north on the Pacific coast with its south flowing current (it originates in tropical Asian waters), than the species on the Atlantic coast with the south flowing current (the cold Labrador current). Subtropical species are found further north (to North Carolina at times) because of the warm water of the Gulf Stream on the southeastern coast of United States. Another interesting aspect of geographic distribution correlated with temperature and depth is that species found in very deep water locations in tropical regions are sometimes found in shallow water in northern locations. As one goes from the shore to deeper waters in the tropics, the water gets colder and it is ecological similar to going north from the same location. The deep sea environment is almost uniform as far as temperature, light and salinity are concerned regardless of latitude or longitude, so species from the deep sea are sometimes widespread and found in more than one ocean.

WHERE TO LOOK FOR MARINE ISOPODS

Marine isopods are found from the seashore to the deep sea floor (Fig. 14, 15 and 16). They are sometimes found in the marine

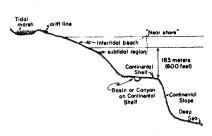


Figure 14. Profile of places where isopods are caught.

drift or debris that accumulates above the beach. Isopods from the marine drift must be adapted to withstand periods of drying, although they are usually able to crawl around and hide during the dry periods in wet vegetation and sand. They are more abundant on rocky shores and are also found on shores where the forest or dense vegetation comes down to the shore itself as in the tropics. The leaves and grass that accumulate in tidal marshes also contain certain species.

In the sand of beaches several different species of isopod are found. One species is very tiny and lives in the cavities between the sand grains. Other species burrow in the sand much like some of the mollusca and small beach crabs. Still other species are found on top of the sand. On muddy or sand-mud areas of the bottom in shallow water where the environment is at least seasonally stable, there species are found that are elongate like worms and that have more or less permanent burrows in the substrate.

Wherever there are plant and vegetable matter on the bottom of lagoons and bays TIOAL FLUX

Sphoerong lids

Limengla

Gosciliation

Firefroid

SANDY

Species

SEACH

Archar

Firefroid

SANDY

Species

SEACH

Archar

Archar

Microcorbena

Some
As allorea

Chickdotea

Mediorea

Chickdotea

Mediorea

Chickdotea

Mediorea

Chickdotea

Mediorea

Species

Sphoerong ide

Chickdotea

Mediorea

Concilian

Figure 15. Isopod habitats. a. Common encrusting organisms on and in a wooden pile. b. In and out beyond sandy beach. c. On rocky beach with tide pool and reef.

and sometimes in the ocean itself, there are species that cling to and eat the live or dead vegetation. Any dead animals found there are also eaten by other species of isopods. It is not meant to imply that isopods are the only animals that perform the aforesaid functions, it only means that they are part of the animal group of many species which perform the functions.

Isopods are sometimes encountered in the surf of beaches and in the sargassum weed (an alga) that floats for long distances and is occasionally found in abundance on the southeastern North Atlantic and the Gulf coast of the United States. Sargassum is also found washed up on the shores of many islands of the West Indies. The surf of some tropical islands is the home of a circlanid isopod that bites swimmers. The bites are not dangerous, but they can be quite annoying if many animals are biting. The open ocean, in general, either near-shore or far from land, is a very poor habitat for isopods. With few exceptions; e. g., Anuropus spp., the only planktonic isopods are parasites of fish or other crustaceans. They are mainly larval or juvenile stages of the parasitic adults. Most

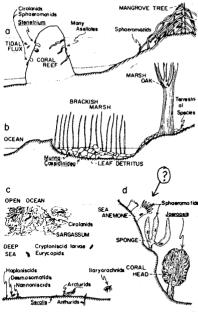


Figure 16. Isopod habitats. a. Coral reef and tropical shore. b. Woodland brack-ish marsh. c. Open ocean and deep sea floor. d. In and among hollow or colonial organisms.

isopods rear their young to juveniles in the brood sacs of the females, but the parasitic forms which undergo a great metamorphosis are free living and at times well represented in the plankton. The relations between the plankton, the isopod and the host animal are just beginning to be explored.

When man builds wharves. piers and jetties on marine coasts, he also makes habitats for many marine isopod species. Not only do the marine structures form refuges themselves to shelter isopod populations, but they also attract plants and sessile animals such as algae and barnacles that themselves become the habitat for more isopods. In fact, it will be shown that many animals common in the marine habitats. especially

those with large cavities either formed within the animal as a coelenterate or formed of colonies of animals such as sponges, can contain isopods, and in some instances the isopods are very specific in their choice of host although they are non-obligate parasites. This is also especially so in the coral habitat of the tropics and southern temperate regions of North America.

One of the most interesting isopod habitat is that which is found around and in a wooden piling. The many animals and plants that cling to the outside of the pile form habitats that contain several species of isopod. The pile itself serves as a substrate into which other species bore holes. Other species can share the burrows, and thus one wooden pile that has been in seawater for a time long enough to collect many encrusting animals and plants is likely to be the habitat of many species of isopod. On rock jetties that stick out into the marine water, there are other species of plants and clinging animals and usually other isopod species. Some species are even known to bore into concrete of the jetty, and of course into the natural stone of the right type on the marine shore. The calm water provided by the jetty and the sandy area of the protected beach are also habitats of particular species which are many times different than those of unprotected beaches.

Although isopods are rarely found in the open-sea, the deep-sea bottom is an environment where a very large number of isopods and isopod species are found. Of course it takes special equipment to collect there, but specimens are provided by an increasing number of deep-sea collecting expeditions. Knowledge of deep-sea species is essential for a complete understanding of isopod systematics and distribution, since they play such an important role in the ecology of the deep-sea bottom. As one collects further from the coast of the continent of North America, the first isopods are encountered in the near-shore habitats, next they are encountered on the continental shelf, isopods of the continental slope (the edge of the continental shelf) are next, and then the isopods of the deepsea are encountered. In each of the geographic areas there are many isopods on the bottom in the sand and mud and they are generally divided into species of the near-shore, the shelf, the slope and the deep-sea. Most of the species from beyond the shelf are blind. In the deep-sea itself there are many different habitats from the Arctic seas to the tropical seas including all the temperate region between. However, the physical factors in the deep-sea are very uniform and there is not too much difference in the "climate" 2000 meters deep off Greenland or 2000 meters deep off northern South America near Panama. The bottom however can be mud, sand, rock or any of the protozoan oozes (purely or in any combination), and there are many different kinds of habitats possible.

One of the most abundant groups of species of isopods is composed of parasitic species. Isopods are parasitic on bony fish and sharks. They are also parasites on squid and perhaps on other large molluscs such as clams and octopuses. Crustaceans, including other isopods, are one of the most extensively parasitized groups of animals that serve as isopod hosts. The isopods are both endoand exoparasites of fish and other crustaceans. They are found in the gill chambers and in the throat of fish and in the branchial (gill) chambers and among the posterior appendages of the crabs and other crustaceans.

Their presence can cause much distortion of the shape and damage to the host on which they are parasites. Many isopod parasites are also distorted in their own shape and are many times twisted along their major body axis when they are attached to or detached from their hosts. One group of parasites undergoes an even more radical change becoming almost unrecognizable as an animal let alone an isopod. They were only classified as isopods when their development and larval form were revealed. Isopods travel with the host animals; and, although most of their ancestors were marine species, they are sometimes found to be parasites on animals that move from the sea to brackish or freshwater.

A LIST OF PLACES TO LOOK FOR MARINE ISOPODS

Marine isopods are found in a wide variety of places and, except in rare instances, they are not usually abundant where they are found. One might look for many hours on a sandy or rocky beach before finding a specimen. Here is a check list of 24 places where isopods might be caught in the marine environment:

- 1. In the maritime drift along a beach or shore line.
- 2. In the sand and also between the grains of sand on a beach (the part of the beach that is continually wet by waves or covered by water at high tide).
- 3. Among and under stones on a beach, especially in tide pools.
- 4. On wooden piles and other wooden marine structures (in the encrusting plant and animal material and in the wood of the pile itself).
- 5. In eel grass (Zoraster) and other sea and marsh grass habitats.
- 6. In algae and other marine plants on rocks, jetties and other marine structures.
- 7. In and under the holdfasts of large algae (especially in southern California).
- 8. Under bark of trees that have fallen into the marine water.
- 9. On and in drift wood.
- 10. On the bottom of bays, sounds and lagoons among the debris.
- 11. In the leaf debris and dead aquatic vegetation in marine marshes.
- 12. In the burrows of other animals on mud flats.
- 13. Under shells, rocks and other debris on mud flats and marine beaches.
- 14. Under and in dead marine animals found along the intertidal shore.
- 15. In the surf on tropical beaches.
- 16. In drifting algae such as sargassum weed.
- 17. In coral reefs and other coral structures.
- 18. Among the roots of mangrove and other trees at the shore next to salt water.
- 19. In plankton (only a few are found).
- 20. In the nets of fisherman and shrimpers (look both for large free living species and for those parasitic on fish or crustaceans).
- 21. In the gill chambers and on the bodies of other crustaceans (including the marsupium of other isopods).
- 22. On fish and in the gill cavity and throat of many large fish.
- 23. In the body cavities of coelenterates, sponges and other hollow marine animals.
- 24. On the deep-sea bottom (of course very specialized equipment is needed to collect in this location!)

HOW TO COLLECT MARINE ISOPODS

Isopods can be collected with a minimum of equipment and effort for the common species at least. For species found in the deep sea and those in coral reefs, special equipment is needed which consists mainly of ship transportation to get to the collection locations. A pair of forceps and a vial of 70% ethyl alcohol is all that is generally needed, however, and the isopods are simply lifted from their hideouts of under a rock on an intertidal beach or from encrusting animals on a wharf or pile, etc., and put into the vial of alcohol. For collecting isopods it is just a matter of ingenuity as to how to seek out the species and remove them from their habitats (see section on Where to Collect Marine Isopods).

If isopods are to be found in beach sand, then much sand must be sifted through a suitable strainer. If the species are to be sought in seaweeds and/or marine plants, then the plants must be gathered with as little disturbance to them as possible and they must be placed in a container of clean sea water and washed or rinsed. The sea water then must be examined for the isopods that were hidden in the vegetation. Rocks, especially those covered with encrusting organisms, from the shore can also be washed in a bucket of clean sea water. After several rocks are washed the water can be examined for isopod specimens. Encrusting plants and animals can also be washed in a similar manner although prying tools are necessary in order to remove the hard material from large rocks and piles.

Sometimes if a small amount of formalin is added to the wash water additional animals will be freed. This should be done only after the material has been washed several times without the formalin since many animals will die still hidden in their hiding places in the washed material. The dead and dying animals many times will be washed out, although it is sometimes necessary to pick them out after separating the material. This is best done under a binocular microscope. Dead leaves and grass from marshes can also be washed. In coral reefs, a piece of coral with many holes in it can be similarly treated, but it also should be broken apart in the laboratory with a hammer. Fragments of coral fly in all directions when hit hard enough to break with a hammer so be careful.

For sand and mud habitats on the bottom of shallow water bays and estuaries, a commercial bottom grab sampler (Fig. 17) or dredge (Fig. 18) can be used. Productive regions usually can be reached by small outboard motor power boats, and the boats can be used to pull a small dredge. The grab and dredge can also be employed in a larger vessel in deeper water and in the deep sea (Fig. 19), but for the amateur this is not practical.

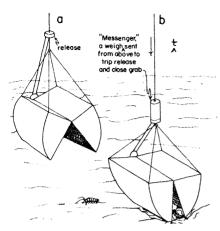


Figure 17. A grab for collecting bottom samples. a. Descending, b. Tripped on bottom by weight from above.

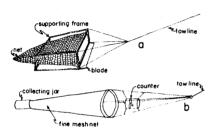


Figure 18. a. Dredge, b. Plankton net.

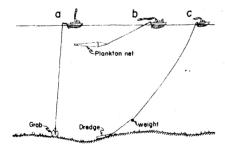


Figure 19. Research vessels. a. With grab, b. With plankton net, c. With bottom dredge.

Sometimes baiting can be employed; i. e., dead fish or other marine animals can be placed in the sand or anchored on a beach. If the bait is anchored under a wire mesh on an intertidal beach and the tide is allowed to cover the bait, then at the next low tide when the bait is exposed, it can be examined for isopods. This might be tried when the bait is exposed at night as well as in the daytime.

It is very difficult to make collection suggestions that apply exclusively to catching only isopods, but part of the fun of collecting isopods will be to see the vast array of other animals that are encountered in the search, After isopods are collected and sorted should be placed in fresh 70% alcohol for storage since the alcohol used for collecting has probably been diluted with sea water. Specimens should never be stored for any length of time in formalin since the calcium carbonate is removed from their bodies and they become soft and difficult to identify. A number written on a small piece of 3 x 5 inch index card should accompany specimens in the vial. The numwith date, place of collection, collector and a short ecological note should be recorded in a field notebook in the field so that there is never any doubt

as to the date, or where, or general ecology of the specimens collected.

THE CULTURE OF MARINE ISOPODS

The water that is used to raise marine isopods should be taken from the general area where the species to be raised was caught because it will at least be of a tolerable salinity. The plankton and small fish that are in the natural water will die causing the water to foul unless it is filtered before it is used. Ordinary chemical filter paper is suitable for the process. Estuary water should be used if the animals were taken in estuaries, and full strength sea water should be used if the species were taken from the sea Natural sea water can be diluted to the desired salinity with glass distilled water. Artificial sea water can perhaps be used, but so far its usefulness in raising isopods has not been determined. Aeriation and/or recirculation of the water are useful and at times necessary, especially for the more active species. Sometimes for tropical species a heating device to raise the temperature can be used. At other times the light and temperature can be controlled in special cabinets, but they are usually too expensive and not available to the amateur

For species that bore into wood, any common soft wood that is placed in the water with the species will soon be invaded by them. Balsam wood tied together in sheets can be used so that the sheets can be separated later revealing the burrows of the species. It must be remembered that any species except those found in the intertidal zone or shallow swamps, must be raised at a lower temperature than room temperature so some control of the temperature will be necessary for most species. It should be mentioned that the methods that are used to raise fish in salt water tanks can probably successfully be adapted to raising isopods. The number of species that can be raised at once, the particular species that can be most successfully raised and the number of individuals that can be raised in one tank must be worked out for the equipment and the species concerned. Whether species can be raised through a complete life cycle is still a question for most isopod species.

Since there has been very little concentrated effort at raising isopods, it should be an interesting and rewarding undertaking. The general rule to keep in mind is to raise the animals in artificial conditions that most nearly approximate their own natural habitats. Temperature, salinity and light are very important considerations. Other conditions such as the absence or presence of certain algae and other aquatic plants is also important. Some isopods might also require the presence of other animals which would provide a substrate or a hiding place for the species.

If the isopod to be raised occurred in or on a plant, the plants after being washed in sea water should be included in the tank where the isopods are to be raised. If an isopod were obtained from a sandy bottom, it would probably be a mistake to attempt to raise it in a tank with a mud bottom and vice versa. Many of the standard apparatus used to raise tropical fish can be applied

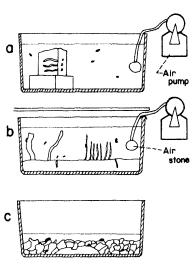


Figure 20. Aquaria for raising isopods. a. Boring isopods in a block of wood, b. Bottom and algae living isopods. c. Detritus from bottom containing isopods.

to raising isopods (Fig. 20). The things to avoid are the extremes of temperature and salinity and one must provide the right kind of biological environment. The food for the isopods should be in part produced by the environment in the tank where the species are raised; but pet fish food, especially artemia eggs and small worms, would perhaps form an acceptable additional source. Experiments must be made on choice of food, since there has been very little work done on isopod culture. Do not use too much additional food because it will cause the water to foul. With patience and an experimental approach, much will be learned about isopod culture. Good Luck!

THE MARSUPIUM AND THE DEVELOPMENT OF MARINE ISOPODS

There are three primary types of brood pouches found among the species of marine isopods. The normal, most common type is sometimes quite modified, but in most species it consists of oostegites which are structures that arise from the bases of the peraeopods on peraeonal segments I to V and form an enclosed cavity called a brood pouch or marsupium. There are generally five oostegites present, but species of *Serolis* have four and species of *Neastacilla*, which have an elongate peraeonal segment IV, have only one oostegite forming an elongate brood pouch (Fig. 55). There are other variations of oostegite number also. Most gravid females, or ovigerous females, have a conspicuous egg mass ventrally located on the peraeon (Fig. 21).

The second type of brood pouch is found in sphaeromatids (Fig. 22), and it consists of special invaginations that fold into the body cavity and form internal (not actually part of the body cavity) brood pouches which have a small opening to the outside that is covered by oostegites. The third type is found in gnathiids (Fig. 23 and 360c), and it is formed as a hollow in segments IV, V and VI and is usually stuffed with eggs and/or embryos. Oostegites are present, but they serve only to cover the openings to the brood pouches. The cavity takes up a large volume of the female body, and eggs can usually be seen through the translucent wall.

Most epicaridean, or parasitic isopods, are known only as gravid females, non-gravid females never having been recorded. The cavity that contains the eggs makes up most of the volume of the species and even houses the male. Some cryptoniscid females are composed mostly of eggs, and most of the body in non-gravid females is composed of little more than the structures forming the brood pouch. In these species there are sometimes five oostegites apparent (Fig. 492), or an undetermined number (Fig. 548).

Very little will be said here about the development of marine isopods other than to say that they have a direct embryonic development. The female lays an egg in the marsupium. Fertiliza-

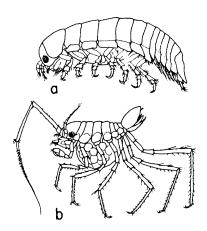


Figure 21. Gravid females (lateral views of females with marsupia). a. Limnoria. b. Munna (Both after Sars).

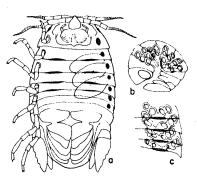


Figure 22. Special marsupium of sphaeromatids. a. Ventral aspect, b. Lateral aspect, c. Detail of marsupium (Modified after Grunner).

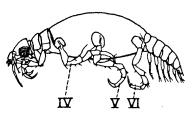


Figure 23. Gnathid female (After Grunner).

tion has already taken place if it is to take place. Some species have eggs that develop parthenogenetically; i. e., without sperm. The egg develops into an embryo and then to a juvenile without passing through an elaborate metamorphosis that is characteristic of crabs and shrimps. When the young free themselves from the marsupium, they look much like their parents, but usually they are at a larval stage and they have only six, not seven, peraeonal segments. At least two genera of anthurids remain at the six segmented stage into adulthood, but most species molt to a seven segmented stage soon after being freed from the brood pouch. They thus become miniature adults. Some species care for their young after they get out of the brood pouch, but most species do not exercise any parental care. Arcturid carry their young on their antennae too for some time after they emerge (Fig. 50).

The species continue to molt at short intervals, and even after they become sexually mature. The molt consists of two stages—first the anterior part of the body is shed then the posterior part. There can be several days between the two stages. The period between complete molts gets longer as the animals become larger. Females lose and regain their oostegites at the molts so they apparently grow larger between broods. In some species there is a change of sex at a molt stage. Some change from males to become females. Other species (especially Asellota) go through an intersex stage at which time they are neither male nor female (Fig. 366). There is much yet to be learned about isopod development, brood times and brood number and also about changes of sex during development.

HOW TO OBSERVE ISOPOD SPECIMENS FOR PURPOSE OF IDENTIFICATION

Many common isopods can be determined to species through the use of a simple binocular microscope. However, for very fine detail such as antennal flagellar article number, the structure of the male reproductive pleopod, or the mouth parts, a regular compound monocular microscope is very useful. For viewing most speci-

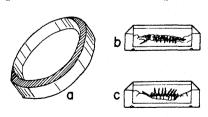


Figure 24. Watch glasses for dissecting isopods. a. Watch glass. b. Isopod for viewing dorsal surface. c. Isopod for viewing ventral surface.

mens, a small dissecting dish (a watch glass or Syracuse dish—Fig. 24a) is best because it is heavy, easy to handle and does not jar easily. If a piece of cotton is put in the bottom, and the watch glass is filled with alcohol to completely cover the specimens, then with the proper lighting the speci-

mens can be viewed from any angle using the cotton to prop the the specimen at the angle desired (Fig. 24b,c). Petroleum jelly (vaseline) can also be used as a prop for small specimens. The petroleum jelly should be put into a dry watch glass and then the specimen and the alcohol should be added. The petroleum jelly is solid enough to hold the specimen, but soft enough so that it can be positioned without injuring it. The petroleum jelly can be quickly removed from the specimen by passing it through a petroleum liquid distillate (gasoline, kerosene or what is probably best, model airplane paint thinner). The specimen should not be allowed to remain in the petroleum distillate because it will harden quickly and become brittle.

The background on which the animal is viewed can be changed by simply dying the cotton black or by placing a piece of plastic or colored cardboard under the watch glass. A piece of material colored black on one side and white on the other can be made if it is not already supplied with the microscope. White pigmentless specimens are generally best viewed with a black background; whereas, pigmented specimens are best viewed on a white background (Fig. 25).

The standard watch glass has the additional advantage of being able to be stacked using one glass as the cover of the other or others. Covering the specimens during the time that they are not under observation can be done simply by placing one glass on top of the other and the watch glasses will fit together snugly in a pile. If petroleum jelly is placed in

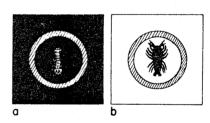


Figure 25. a. Light specimen on dark background. b. Dark specimen on light background.

between the two watch glasses, the alcohol will not evaporate and the animals can be stored for several weeks in the container thus formed.

For larger specimens larger bowls must be used, but the technique or positioning the animals for observation is about the same. Lighting can be provided by any convenient binocular microscope lamp, or, if one is not available, a reading lamp can be used or the microscope can be placed near a window. The large isopod can be manipulated and the appendages can be removed by ordinary forceps and a teasing or dissecting needle which are supplied in most general biology kits. However the fine dissecting needles described below are very useful for manipulating small specimens.

A firm support for specimens being dissected or torn apart can be made by placing modeling clay instead of cotton in the dissecting dish. The clay does not break apart in the alcohol. Of course, the dissections should take place under 70% ethyl alcohol solution (the same solution in which the specimens were collected). I say dissection should take place under the alcohol, because many people make the mistake of not covering the specimens properly and light is scattered from many wet surfaces of the specimen and fine structures cannot be properly viewed. If the specimens are completely covered with alcohol so that there is only one surface, the alcohol surface, between the viewer and the specimen, then light reflected from this surface is not reflected into the microscope and detail can be clearly seen. Under the heat of the light of the lamp, the alcohol evaporates rapidly and must be replaced to continue viewing the specimen properly and to prevent it from drying out.

Small appendages or parts of specimens that are removed can be placed on a slide in pure glycerine (available from a drug store) or glycerine jelly (available from a biological supply house), and when the cover slip is placed over them they can be viewed with a binocular microscope (lighted from above or below the specimen) or with the compound microscope with regular beneath the stage lighting. Mounting the specimens can be done on a plain slide or in a depression on a depression slide (Fig. 26).

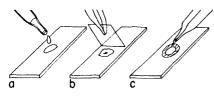


Figure 26. Preparation of specimens for viewing in a compound microscope. a. A small drop of glycerine being placed on a slide. b. Cover glass placed on after specimen added, c. Specimen being placed in a depression slide.

When the slide is viewed in the microscope with transmitted light (light coming from below the specimen), most of the details, including some internal details, will be revealed in the structure viewed. If the specimens are allowed to remain in the glycerine or glycerine jelly, in several days even more detail will be revealed, since the mounting agents are

also clearing agents which take away the dark pigments of the tissue and more detail is revealed. Do not forget to properly label the slide by attaching a standard slide label or using a colored wax marking pencil. Slides with a frosted end suitable for marking with an ordinary pencil are available and are very handy for making slides that are to be stored. The slides with their specimen parts can be stored indefinitely if the edges of the cover slip are sealed with clear finger nail polish or a commercial preparation.

For very small specimen handling, very fine watchmakers forceps are best, but they are relatively expensive. Very small dissecting needles can be made from insect mounting pins. The especially fine minuten pin used for pinning very small insects works very well if it is attached to a wooden match stick. The small needle can be pushed into the end of a slightly pointed match stick and fastened with epoxy glue (Fig. 27b). A crude but effective dissect-

ing needle can be made very easily by sticking a large insect mounting pin (or a straight pin for that matter) through the eraser of an ordinary lead pencil (Fig. 27a). A pair of teasing needles or dissecting needles that come with the standard general biological dissecting kit are very useful for



Figure 27. Dissecting needles. a. Lead pencil with insect pin (or straight pin) in eraser, b. Wooden match stick with minute insect pin in end glued firmly.

handling and sorting specimens, but they are generally too large for dissecting the average size isopod.

STORAGE AND LABELS

For storing specimens of isopods for any length of time, the best container is a wide mouth canning or freezer jar (available in most grocery stores). Cotton (available at drug stores in rolls) is placed on the bottom and the isopods are put into shell vials (available at most biological supply houses and some drug stores) and they are placed in the large jars. The storage jars can be either pints or quarts (Fig. 28). The advantages of placing the shell vials of specimens in the large jars are that species can be stored according to category—species, genus, family or location, etc.—and it is only necessary to watch the level of alcohol in one large container rather than many small ones. If the shell vials are stored in an

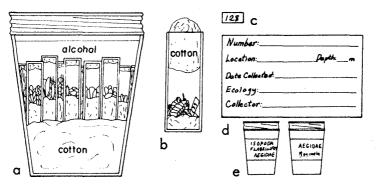


Figure 28. Storage and labeling apparatus, a. Canning or freeze: jar for storage of shell vials containing specimens, b. Shell vials with specimens, c. Notebook number to be kept with specimens, d. Label form for shell vials, e. Storage jars with labels.

inverted position in the storage jars, there is less chance that the specimens will dry out if the cover is not tight and the alcohol slowly evaporates. The cotton protects the specimens from injury when they are transported in the jars.

Of course larger specimens than average can be stored in the jars directly without first being placed in the shell vials. Sometimes a crab or fish host as well as the isopod parasite can be stored in a single large container. A large collection of specimens can be stored in the jars and the jars can be stored by the dozen in the carton in which they were purchased.

One of the most important things that must be included in every collection of isopods is the label. Specimens, no matter how exotic and interesting, have very little scientific value if they are not properly labeled. The label should be made of high quality, heavy weight, watermarked bond paper or heavy white cardboard as in a 3 x 5 inch index card. The ink used to write the label should be black waterproof and alcohol proof drawing ink. Most good quality "India" inks will do.

The label should be printed and should include the name of the species (or the smallest taxonomic category to which the specimen has been identified), the collector, the date of collection and the location where the collection was made. If possible some brief ecological data should be included such as "from beach sand," "from under rocks on stony beach," "from wooden piling," etc. For deep-sea species there is always a log of scientific events kept by the chief scientist and stored at the institution which sponsored the expedition. Information from the log is generally available to an interested person. It contains all the information as to position, depth, and physical factors at the place where equipment was lowered into the sea to catch animals including isopods. As much of this information as is possible should be stored on labels with the specimens. For parasitic isopod specimens, the name of the host should be recorded. The depth at which the species or host were taken should also be recorded.

A number should also be included, and it should have been placed in the collection jar with the specimen in the field. It should be immediately recorded in a field book with as much information as is practical added at the same time. The number placed in the collection jar in the field should stay with the specimens at all times. When the specimens are observed under a microscope, the number should be included in the watch glass and it should always accompany the specimens so that there is as little chance as possible for specimens to be confused with one another.

WHAT IS A MARINE ISOPOD?

A marine isopod crustacean at first glance might look like many other marine crustaceans that are related to it, but there are certain characters that set them apart from the other animals that the isopod might superficially resemble. A list of the characters follows:

- 1. Body usually dorsoventrally flattened.
- 2. Two pairs of uniramus antennae.
- 3. Sessile rather than stalked eyes.
- 4. Four pairs of appendages modified as mouth parts. (The maxillipeds are attached to the cephalon.)
- 5. Lateral parts of the body segments many times extended by epimeres or coxal plates.
- 6. Uniramus locomotoral (ambulatory or natatory) or prehensile peraeopods on peraeonal segments seven in number. Anterior pair or pairs can be gnathal or subchelate (never truly chelate) in structure.
- 7. Brood pouches on female peraeonal segments formed from three (one in some arcturids) to five pairs of oostegites on that many peraeonal segments.
- 8. Five appendages ventral on pleotelson or pleon; first sometimes operculate and sometimes others operculate. Second always sexually modified in male, and the others respiratory in function.
- 9. A single pair of usually biramus uropods variously modified present on pleotelson. The rami are uniarticulate.
- 10. Telson flattened and usually fused with one or more than one pleonal segment.

The above mentioned ten characters are morphological aspects of the isopod that are seen without elaborate preparation of the specimen. To each of them there are interesting exceptions, but if the ten characters are kept in mind the exceptions will become known with experience.

There are several internal anatomical characters that set the isopods apart from other related crustaceans. The heart of an isopod is in the dorsal part of the pleon. The anterior part of the digestive tract is generally equipped with a gastric mill (for grinding food particles), and isopods have a straight digestive tract usually with only two pairs of digestive glands. Isopods have a circumesophageal nerve ring (a ring of nerve tissue around the anterior part of the digestive tract) from which nerves lead forward to the eyes and antennae, and two main nerve cords lead to the posterior part of the body. A thickening or ganglionic mass

with connections between the cords is present in each segment. From the ganglionic thickenings small branches go to the appendages and other functional parts in each segment. Another so called "sympathetic" nerve system is present leading from the nerve ring in the cephalon to the posterior of the body. There are no thickenings in each segment and the function of the system is only beginning to be studied.

The body of an isopod crustacean is primitively divided into 19 segments each with its modified appendages. The 19 segments are divided into three major groups of segments or tagmata—the cephalon, the peraeon and the pleon. Only the segments of the peraeon and the pleon ever show in the dorsal view except in rare exceptions when one segment of the cephalon the maxillipedal segment is separated from the fused anterior group. Except for the occasional presence of the maxillipedal segmentation there is little to indicate segmentation in the cephalon of an isopod.

The next seven segments make up the tagmata called the peraeon. There are peraeopods or peraeonal appendages on the peraeon and they are modified in many ways, sometimes being only rudimentary and sometimes being absent, but mostly they are locomotoral in function. Seven pairs of peraeopods are usually present, but in the gnathiids there are seven peraeonal segments and only five pairs of peraeopods. The seven body segments to which the peraeopods are attached are in most instances indicated in the dorsal view of the isopod. The segments which contain the legs are called the

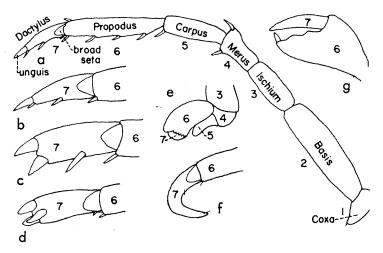


Figure 29. Peraeopod types, a-d. Ambulatory peraeopods. a. Peraeopod with segments named, b. Single unguis or claw (uniunguiculate), c. Two claws (biunguiculate), d. Three claws (triunguiculate), e-g. Prehensile peraeopods, e. Epicaridean type, f. Flabelliferan type, g. Asellotan type.

thorax in most crustaceans, but the word peraeon is coming into more use because there is much difference between the "thorax" of an isopod and the thorax containing the lungs of a vertebrate animal. The legs of the peraeon, more properly the peraeopods (Fig. 29) are modified for clinging (prehensile—Fig. 29e, f), walking (ambulatory—Fig. 29a-d) or swimming (natatory—Fig. 31).

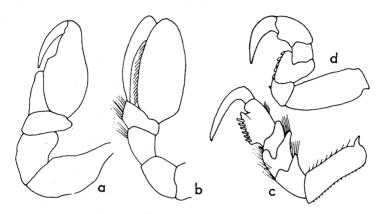


Figure 30. Peraeopod types. a. Anthurids (Cyathura). b. Sphaeromatid (Tecticeps), c. Rocinela, d. Aega,

Sometimes they are dimorphic; i. e., they (especially the first and seventh) are different in the male, female and juvenile. At times the peraeopods are modified for straining food particles (Fig. 49), and sometimes they are modified for digging (fossorial).

The oostegites that cover the brood chamber (Fig. 21) in the female are parts of the peraeopods. The isopod female is frequently longer than the male species because it is modified to carry the brood. The oostegite shape and thickness are characteristic of different

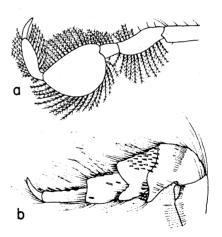


Figure 31. Peraeopod types. a. Natatory (Ilyraachna). b. Modified natatory (Cirolana).

groups, and their presence or absence on various segments are also characteristic in each group. Within the brood pouch or marsupium formed by the oostegites there are no indications that a glandular

secretion is extruded by the female (as in terrestrial isopods) to nourish the eggs and young which are hatched as juveniles within the oostegite covered pouch. They soon are released from the marsupium after they have attained suitable size.

The appendages of the pleon, the pleopods, are the sites of the breathing function or of respiratory gas exchange of the isopod. The first pleopods (more rarely the second or third) are sometimes operculate or form a covering structure to the other pleopods that are enclosed within a branchial chamber (Fig. 366). Sometimes the pleopods are fringed with long plumose setae and are said to be natatory. The second pleopod (the endopod) in the male is modified as a sexual structure in most species, and it is very characteristic of the species and can be quite effectively used in making identifications of many species.

The pleon contains a terminal structure called the telson which, in the majority of isopods, is modified by being fused to the sixth pleonal segment, and this forms a structure called the pleotelson from which the anus opens. The appendages of the sixth pleonal segment, the uropods are usually laterally or terminally located, and they are the site of many sensory setae. In the valviferans the uropods form the operculate covering to the branchial cavity (Fig. 45) which contains the pleopods. In many isopods the expanded uropods and the expanded pleotelson combined form a caudal fan which is the principal swimming and steering structure.

A much more detailed picture of the isopod follows with some account of the probable function of the structures. The cephalon has been formed by the process of cephalization of the anterior body segments of the trilobite-like primitive crustaceans. There is rarely any indication of segmentation, but the mouth parts are embryologically formed in the same manner as the peraeonal appendages or pleonal appendages. From the anterior two pairs of segments arise the sensory appendages, the antennae. Eyes also are found on the anterior segments. The anterior part of the animal receives the tactile and chemical information so that the animal knows its place in an environment and what is the chemical nature of that environment. In isopod crustaceans the antennae are uniramus; i.e., there are four antennal branches projecting from the cephalon of the isopod, not eight as in many shrimp and crayfish. There are two pairs of antennae and they are of different lengths, the second pair generally being much longer than the first pair. The second antennae are more specialized than the first to receive the tactile sensory information and the shorter first antennae are more specialized to receive the chemical information. Antenna one usually has only three peduncular segments and a varying number of flagel-

lar articles (Fig. 32b). Special broad flat setae called aesthetases (Fig. 33) are present on the apical flagellar articles and they function as the chemically sensitive sites on the antenna. Antenna two has five (in some species six) peduncular segments. On what is primitively the third although it might appear to be the second is a short segment, an antennal scale or squamma; it most probably represents what is all that remains of the primitive exopod of the second antenna since the antennae were primitively biramus appendages (Fig. 32a). It is found most frequently in species of Asellota (Fig. 34 and 35).

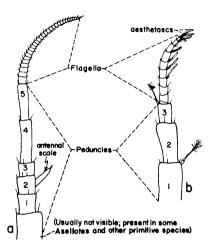


Figure 32. Antennae one and two. a. Antenna two (Sometimes the basal segment is present; e.g., in some osellotes, and the numbers begin with the primitive precoxal segment thus the antennal scale, or primitive exopod, is on segment 3 not 2 according to some workers.). b. Antenna one.

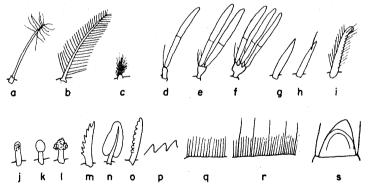


Figure 33. Setal types. a. Sensory seta. b. Plumose seta. c. Pencillate seta. d-f. Aesthetascs. d. uniramus. e. biramus. f. triramus (Microarcturus). g. Spine or stout seta. h. Stout seta with sensory tip. i. Special seta on peraeopod of Paracerceis. j-l. Coupling hook types from maxillipeds. m-o. Sensory edge of maxilliped setal types. p. Scales on sensory edge of maxilliped (Jaeropsis). q. simple fringing setae. r. Compound fringing setae. s. Broad propodal seta at aticulation of propodus with dactylus.

The mouth parts are found in what is a large buccal mass ventral on the cephalon. They are covered or limited anteriorly by the frontal lip (not a true appendage) or labrum (Fig. 36a). The

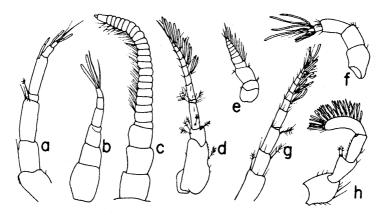


Figure 34. Antennae one types. (See Fig. 368 for Asellotan types). a. Cymothoid. b. Anthurid (female). c. Anthurid (male). d. Sphaeromatid (Paracerceis). e. Cirolana (male). f. Flabellifera (Limnoria). g. Cirolana (female). h. Valvifera (Erichsonella).

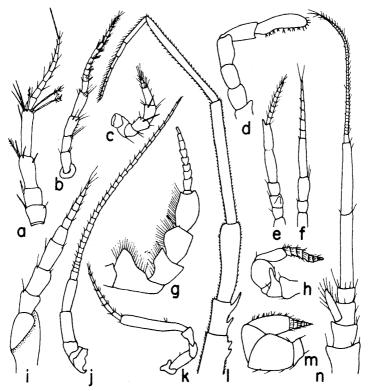


Figure 35. Antennae two types. a. Gnathiidea. b. Sphaeromatid (*Paracerceis*), c. Limnoria. d. Cleantis. e. Antennuloniscus. f. Hydroniscus. g. Saduria. h. Haploniscus. i. Anthurid. j. Cirolanid. k. Synidotea. l. Arcturid. m. Jaeropsis. n. Stenetrium.

37
Paragnoths (= lower

mandibles (Fig. 37) are strong and generally serve as biting or chewing structures although in some groups they are piercing and sucking instruments. In the gnathiids they probably serve in a secondary sexual capacity. The mandibles in

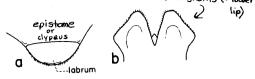


Figure 36. a. Frontal lip or labrum. b. Posterior lip or hypopharynx.

most have a sensory palp of three articles. The palp is sometimes reduced or absent as in the Valvifera and Oniscoidea. Near the incisor process (Fig. 37) or the cutting part of the mandible, but sometimes absent on one or both mandibles, is the lacinia mobilis, a moveable appendage found in most peracarideans including isopods. Just behind the mandibles and just anterior to the entrance

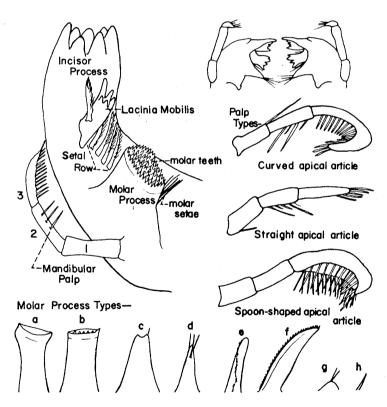


Figure 37. Mandible with palp types and molar process types. a. Broad and truncate. b. Truncate with teeth. c. Blunt pointed. d. Pointed with apical setoe. e. Spiniform (Jaeropsis). f. Blade-like (Cirolana). g. Lump with setae. h. Seto only.

of the digestive tract, there is a hypopharynx, (not a true appendage) or posterior lip, which serves as a sensory and straining structure (Fig. 36b).

The next true gnathal appendage is maxilla one which is generally composed of two lobes—a sensory endopod and a biting exopod (Fig. 38b). Maxilla two generally is composed of two lobes also,

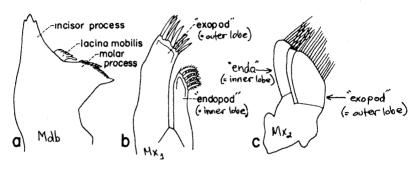
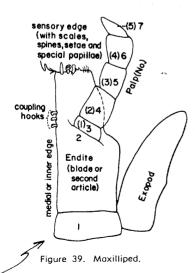


Figure 38. Mouth parts of Sphaeroma destructor, a. Mandible, b. Maxilla one, c. Maxilla two.

but one is generally double so that there are three lobes present (Fig. 38c). The second maxilla are food strainers when there are long setae present on them. Sometimes the lobes are fused into a single broad structure that has no long fringing setae (Fig. 215d), but that type is confined to some fish parasites and terrestrial isopods. Both pairs of maxillae have various other functions in the

- a aymothnic maxilla!! Not fused at all!



masticatory process, and in some species they are absent or only rudimentary structures. The maxilla are quite different in different species, but they are not used as key characters in the keys to species that follow because they are not always easy to observe.

The maxillipeds (Fig. 39) are the covering structures to the buccal cavity containing the mouth parts. The endite or second segment is generally broad and flat with coupling hooks on the medial margin. The upper edge of the endite and the palp are sensory in function. In some species where the mouth parts

his is all arong! see back fly page.

are modified for sucking and piercing, the palp is absent or reduced to only one or two articles. In some fist parasites (e.g., aegids) there are long spines or teeth on the apical article of the palp. There is much variation in the number of palp articles and the width of the palp, both absolutely and in relation to that of the second article or endite, also varies (Fig. 370). The maxilliped and its palp are very easy structures to observe in most species and can be used effectively for taxonomic discriminations, especially to help confirm the identity of a species or genus.

In the gnathiids there is an additional mouth part—the pylopod (Fig. 348). The pylopod is a special modification of the appendage of peraeonal segment I. It is operculate to the buccal cavity and has the same function as the maxilliped in other species. Apparently the true maxillipedal appendage in a gnathiid performs a food

straining function and the maxillae are rudimentary.

The seven peraeopods of the peraeonal segments are variously modified according to their function (Figs. 29-31). Generally the first pair of peraeopods are modified in some manner as food handling, grasping or secondary sexual appendages. Males of many species, especially asellotes, have the first appendage modified as a secondary sexual structure. In males it is generally subchelate and is probably as useful in grasping vegetation as in grasping meals or females. At times the first three peraeopods are subchelate as in the tube dwelling anthurids. The modified peraeopods are probably aids in food gathering. The last four peraeopods are locomotoral structures in most species. They can be either ambulatory or natatory (Fig. 480).

In some ectoparasites of crabs and other crustaceans, there is a loss of some peraeopods on one side of the asymmetrical body. Many parasites that live within the host's body cavities completely lack functional appendages. The peraeopods that are found in the ectoparasitic species are small prehensile structures. The isopods that are fish parasites have peraeopods that have very sharp claws for clinging to the body of the host (Fig. 30). Some cave species have clinging peraeopods, but they perhaps are modifications for clinging to the walls of the cave, not for clinging to a host fish. Sometimes only the first pair of peraeopods are modified for clinging; sometimes all or almost any of the first three or four pairs are modified for clinging.

The peraeopods of other species (e. g., munnids) are extremely long so that the species look like daddy long leg spiders (Fig. 21). In other species the first pair are modified for gnathal functions, the third and fourth for ambulatory, and the fifth to seventh for natatory functions (Fig. 457). In other species the peraeopods are short and fit within the sphere of the rolled up body. The species are in a family called Sphaeromatidae since they form a sphere. In still

other species the peraeopods are fossorial or modified for digging in the substratum.

There are six segments plus a telson in the pleon of the primitive isopod, and in some modified parasitic species and some tube dwelling species, a true telson with six free pleonal segments are still found. However, in most species the telson is fused in some manner with one or more pleonal segments forming a pleotelson. Many times the pleotelson is broadened as in the circlanids and with the broadened uropodal rami form a caudal fan used for swimming. The posterior segment fuses with the free pleonal segments forming a pleotelson which sometimes includes the whole pleon. In many instances only the first pleonal segment is free and there might or might not be indications of another or other segments. All segments of the pleon are fused and the pleon itself is fused with the peraeon in some species (e. g., Haplomesus).

The pleopods which are the five pairs of appendages of the anterior five pleonal segments are usually all free even when all of the segments are fused. The pleopods are also with few exceptions biramus. In two suborders, the Valvifera and the Asellota, the species have the pleopods enclosed in a branchial cavity. The number of pleopods is most always five pairs, but they are reduced in number and size in many of the parasitic bopyrids. In some species they are absent. The function of the pleopods (except for the sexually modified male pleopod 2) is sometimes natatory, but almost always they are modified for the exchange of respiratory gases or breathing. Species of the genus *Iais* utilize the pleopods of species of sphaeromatids for a home and are found almost exclusively among the pleopods of the host species.

The uropods are modified in many ways in addition to being valves or branchial cavity covers in the species of Valvifera. They can be either uniramus or biramus (Figs. 40, 41 and 42). They most

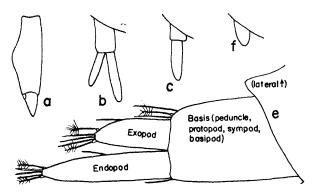


Figure 40. Uropod types. a. Valve or operculate branchial cavity cover (Valvifera). b. Simple biramus. c. Simple uniramus. d. Basis or peduncle only. e. Biramus uropod with setae.

probably are sensory structures, and are well developed and specialized in species that back into holes (anthurids). They are at times almost completely absent or absent in some species. Frequently they are broken off during the collection process so that their absence might be mistaken for normal absence. The margin of the telson, the last body segment, is generally fringed with sensory setae and the anus opens beneath it.

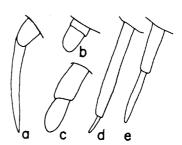


Figure 41. Uniramus uropod types. a. Ischnomesus. b. Haploniscid. c. Ianirella. d. Macrostylid. e. Acanthocope.

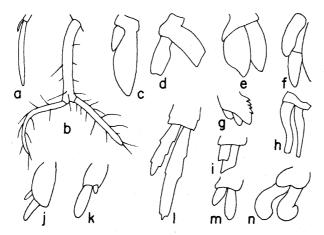


Figure 42. Biramus uropod types. a. Asellota. b. Acanthomunna. c. Cassidinea. d. Sphaeromatid (Paracerceis). e. Cirolanid. f. Anthurid. g. Jaeropsis. h. Gnathiidea. i. Munnid. j. Ilyarachnid. k. Serolis. l. Stenetrium m. Eurycope. n. Epicaridea.

KEY CHARACTERS AND OTHER CHARACTERS

It is important to know about key characters and other characters since all identifications in the book are based on the use of key characters in a key. Key characters are observed and then discriminate judgments are made that will lead to the identification of isopod specimens under study. Key characters should be easy to observe and consistently present (or absent) on almost every member of the taxon under study. Many times there are, in addition to key characters, other characters that are peculiar to the taxon or a species under study that are definitive, but the additional characters do not appear in a key to the species. There are illustrations

of structures in this book (e.g., maxillipeds) that many times in themselves can be used to identify a particular isopod to a particular taxon, but they should not be used by themselves until more than just a general knowledge of isopod morphology is obtained.

A major difference between insects and crustaceans is the difference between the structure used by the animals in the respiratory exchange of gases with the environment; i. e., the breathing structures. Insects with very few exceptions have spiracles and a tracheal system that functions to exchange gases in an environment of gases, the atmosphere. The crustaceans which are usually found in an environment of water, have various structures on their bodies that function in respiratory gas exchange—the general thin body wall, thin membraneous appendages, gills or specialized pleonal appendages (the structures used by isopods). Crustaceans never have tracheae or any structure that are really like true tracheae of insects.

The structures used in respiratory exchange represent a fundamental difference between insects and crustaceans, but they are rarely used in a key made to discriminate between insects and crustaceans because they are not simple easy to observe structures that can be observed with simple equipment. Internal characters are not used for the same reason.

It is important to get to know the difference between what is a key character and what is a major difference between species, but not easy or useful to use in distinguishing animals. A key character is merely the most useful character from a set of characters to use when distinguishing animals from one another. This applies to any level of distinction from the phylum to the species, and is why a description of the different kinds of characters is included here. It is also important to know that an isopod crustacean is defined on much more than a single or a few characters. It is an animal that can be defined on a whole set of characters, but key characters are used to make distinctions between groups because the lists of differences between two widely divergent animals can be quite large. Many times several key characters can be used, but it is best to use a small number for ease in making the series of distinctions that finally lead to the correct name and identify the isopod specimen at hand. Additional characters can be used as a check on the main key characters, but the key (with the figures for assistance) are the best characters to use.

There are two types of keys recognized by taxonomists that are used to distinguish animals—a natural key and an artificial key. If evolution took place and all animals are related to each other as is represented by a phylogenetic tree, then the distinctions that are used to identify different taxa in a key are the same ones that are used to construct the phylogenetic tree itself. The branches of

the tree are dichotomous just as the couplets in a key are dichotomous. In practice this is generally done as much as is possible, but still there are many perfectly valid characters that are obscure to immediate observation or are internal and only can be considered after dissection that are used in forming the phylogenetic tree pattern of animal evolution. Key, or artificial characters then, not natural characters are used to make the distinctions between taxa. The distinctions are then mostly artificial, because they are chosen without regard for the evolutionary importance for the animal keyed. Hence, any really useful key will be composed of natural and artificial characters, and there is really no practical reason to argue the merits of either one any further.

Completely artificial keys are sometimes constructed when the fauna from a particular geographic region are studied and identified. The animals are distinguished with little regard to their place in the phylogenetic tree of isopod evolution. The larger the geographic region studied, the more a key can be composed of natural or phylogenetic distinctions related to the evolution of the animals included. The key in this book represents a very large geographic region—the waters around the continent of North America—so the key at least in higher categories can be somewhat of a natural key. Many species that would fill in the branches of a phylogenetic tree are not represented in the geographic region included in the book so the key is not a true natural key. It is hoped that the key will serve to identify most of the isopods of the New World to genus. It will serve as a key to all of the suborders except the Oniscoidea, the terrestrial species, and the Phreatoicidea, a specialized group in Australia, New Zealand and South Africa. Any species except in those two suborders will key to suborder on a world and species of the suborder on a world and species except in the suborder of the s many will be identified to the family level. Only where there is overlap in distribution will Old World genera and species be included and there are many species that are common to the two regions, the New and the Old World.

HOW TO USE THE KEY AND NOMENCLATORIAL CONSIDERATIONS

The key used in the book is of the dichotomous type. Each couplet is composed of two lines, an a line and a b line. Each couplet describes a character or set of characters which are then compared to the same characters of the specimen under the microscope. The specimen is identified if it has or lacks the structures indicated in one couplet of the key. Reference to another couplet is then made, and further distinctions between two choices of characters

are made. The first couplets are used to identify the specimen to suborder and the page number where the account of the suborder is found is given. Couplets leading to family identification (with page number given) are then encountered. The last set of couplets leads to genus and species, where there is a figure of the species given.

At the species level there will be found a reference to figure number, a species name, the author of the species (in parentheses even if he originally described the species in the same genus) and the length in millimeters of a large specimen of the species. The geographic range and usually the depth in meters where the species was taken follow. The depth is generally given as a range to include the various geographic sites and depths where it was found. A small paragraph of explanatory matter then may or may not follow. Species recorded for a particular depth at a particular geographic location will probably be reencountered at that place and depth, but one should not be surprised if a particular species is encountered at a different depth and a different geographic, but nearby, location because they were recorded only once or a very few times before.

Almost any marine isopod that the reader will casually encounter in the field will be contained in the book; it would be very rare indeed for an inexperienced person to find an isopod that would not fit or be keved to suborder or family. A specimen that did not fit the species key, would be most probably encountered. However, the probability for any specimen that an amateur collector finds to not fit the key is slight. If the specimen seems not to fit the key, then the person using the key must reconsider all the choices that he has made. It has been the author's experience to discover that a specimen which a student was keying was not even an isopod, and of course the probability of its not even being an isopod must also be kept in mind by the beginner. If a student finds a new higher taxon, he has probably not used the key properly. Specimens taken from North American waters where few collections were made in the past, however, should be examined carefully because it is possible to find a new species of isopod.

The correct use of the key is based on a knowledge of the vocabulary and the figures. For most of the descriptive words used in the key, the illustrated glossary index has a definition or there is a reference to a page on which the definition is found in words or pictures. If the word definition is combined with an illustrated definition, then a clarification of the term and its meaning as applied to the specimen under the microscope should be apparent. The meaning of the words given in this book are definitions pertinent to the study of isopods, although the words might also be used in other fields of taxonomic carcinology. Unfortunately, the terminology and the nomenclature of parts is not standard throughout the invertebrates, nor is it standard throughout the study of crustaceans for that matter. That is another useful function of this book—to standardize the nomenclature or at least provide a place where the common terms applicable to isopods are located and defined. The illustrations serve in part to help make the meaning of the words clear, and in some of them there are two or more names for some structures. The preferred term for the structure will be the one best defined in the illustrated glossary section; i. e., the one to which all alternate definitions are referred.

The term cephalon is used for the head, peraeon for the thorax and pleon for the abdomen. Antennae and mouth parts are names used for the appendages of the cephalon. Peraeopods are appendages of the peraeon whether they are locomotoral—ambulatory, natatory and prehensile—or secondary sexual in function. Pleopods and uropods are pleonal appendages. It is not necessary to elaborate, but the use of invertebrate terms for invertebrate structures is preferable to the use of invertebrate terms for invertebrate structures.

A WORD ABOUT THE ILLUSTRATIONS

The illustrations of species that are used in this book are not original except for a very few. They are for the most part line drawings taken from the originals, and are either enlargements or reductions of the original drawings of other authors or persons who revised the work of an original author of a species. Except where indicated, the scale on the lower left hand side of the main figure represents one millimeter of length of what is a large specimen of the species illustrated. That is to say, if a species were stated by its author to vary from 8 to 12 mm in length, then the scale at the side of the illustration represents one millimeter of the length of the isopod illustrated or 1/12 of its length. Where the line represents less than one millimeter, it is so stated below the scale. A generalization to keep in mind is that the longer the line representing 1 mm is in the illustration, the smaller the specimen is; and the reverse, the smaller the line, the larger the specimen. The cymothoid isopod illustrated on page 157, figure 236 is very large, indeed, 45 mm long. The munnid illustrated on page 293, figure 465 is only about 1 mm long since the line is about as long as the illustration of the specimen.

At times a whole figure of the animal has not been given. It is because it was not illustrated by the original author of the species, or it differs very little from other species in the genus. When accessory structures are illustrated in addition to the main figure, they are rarely on the same scale as the main figure generally being

enlargements. Since isopods molt and grow all during their lives, it would be misleading to place a scale next to the illustration of an accessory structure. The structure might be of a larger or smaller size, but of the same configuration in another larger or smaller member of the same species. Size comparisons in invertebrate animals in general, and in crustaceans in particular, are very misleading since the animals do not reach a particular size and stop growing within a very narrow range that is characteristic of many common vertebrate organisms. The illustrations of accessory structures, unless otherwise indicated, are of mature individuals. At times the original author might not have known that his specimen was an immature individual, so there will probably be some errors made if all illustrations are thought to be of mature specimens. Where there are fundamental differences between the species illustrated here and that illustrated by the original author, then further work demonstrated that the original author was in error.

The illustrations can be used as a starting point for the identification of an isopod specimen. If a specimen under the microscope looks very much like an illustration in the book, then the key can be used in a backward direction to discover the identity of the specimen. The identity should be confirmed however by starting again at the beginning of the key so that all alternative considerations are eliminated.

KEY TO ISOPOD SUBORDERS FOUND IN NORTH AMERICAN WATERS

- 1a. Body symmetrical or at least with traces of bilateral symmetry; peraepods distinct (Fig. 7, except h and i) _______2
- 1b. Body asymmetrical or not apparently based on bilateral symmetry; peraeopods not present or not easily seen (Fig. 7h and i); parasitic on Crustacea...... Epicaridea (in part) (p. 338)
- 2a. Uropods lateral, or lateral and operculate, or uropods absent (Fig. 43)3

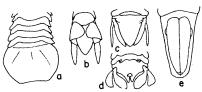


Figure 43. Lateral, absent (dorsal view) or ventral uropods. a. Absent (Anuropus). b. Anthurid. c. Exosphaeroma. d. Paracerceis. e. Valvifera (Uropods are valves or covers of branchial chamber).

2b. Uropods terminal (occasionally dorsal), or if present they would be terminal (Fig. 44)

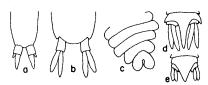


Figure 44. Terminal Uropods. a. Micro-cerberidea. b. Asellota. c. Epicaridea. d-e.

- 3b. Uropods lateral or absent 4

- 5b. Blind or with eyes; body length less than 7 times width; usually longer than 2 mm 87 6x./ Plestelson "I"-shaped or triangulate; uropods with short base,
- 6a./ Plegtelson "T"-shaped or triangulater propods with short base and long rami (Fig. 44d-e) Chathidea (p. 217) 6b. Not as above
- 7b. Body usually not symmetrical; pleon never with branchial cavity and with pleopods various; antennae always rudimentary or very small; parasitic on Crostacea...... Epicaridea (p. 309)

SUBORDER VALVIFERA

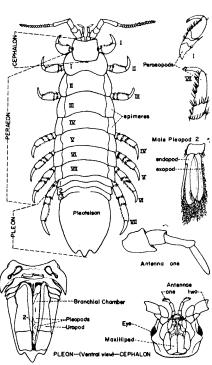


Figure 45. Valvifera (General no menclature).

Three characters are always present in the Valvifera found in North American The uropods form waters. valves which fold over the branchial chamber as a cover. The flagellum of antenna one is always formed of a single / fused segment and the mandible has no palp. The pleon always has several fused segments and the frontal margin of the cephalon rarely has a long rostrum. The North American species are divided into the two families-Arcturidae and Idotheidae. The arcturids are found in the deep sea, and indeed are filter feeders characteristic of that extensive habitat. They are found in deep water off Florida and in tropical regions, but in the northern part of their range they are found in shallow water. The idotheids are common in the near shore habitats along the margin of the continent and are especially abundant in the algae that is found there.

All species of Valvifera are highly calcified. Arcturids have a tendency to have very large spines on their bodies,

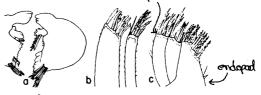


Figure 46. Mouth parts of Valviferan species. a. Mandibles. b. Maxilla one. c. Maxilla two.

and idotheids have smooth bodies but some species have spines. Spines are in part a character that shows the maturity of the specimen and in part a secondary sexual character generally being absent or smaller in females. There are other differences between the two families as will be seen. There are about **X* recorded species in North American waters. They are divided in addition to the two families into 18 genera.

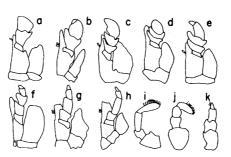


Figure 47. a-h. Maxillipeds of Valvifera, a. Synidotea, b. Cleantis, c. Idotea, d. Idotea, e. Penidotea, f. Saduria, g. Pleuroprion, h. Arcturus, i-k. Antennae one of Valviferan species, i. General, j. Penidotea, k. Cleantis.

KEY TO VALVIFERAN FAMILIES*

note: There are la valviterau families, but Idoteidae & Arcturidae (= Actuallidae) occur in No. America (or Europe).

ARCTURIDAE

Specimens of arcturids are small to large in size. Small and medium size species are found in the near shore habitats and large species are found in the deep sea. The flagellum of antenna one in all of them is of a single article. Antenna two has the last two articles of the peduncle elongate and there is a short flagellum of few to many articles. Eyes are present and generally located laterally or dorsolaterally on the cephalon. The mouth parts are all well developed and frequently peraeopod I is modified as a small gnathal appendage located directly behind (or beneath) the maxillipeds. Generally peraeopods II to IV are modified by the presence of long setae and serve as food straining structures (Figs. 48 and 49). Peraeopods V to VII are ambulatory. The pleon usually has all segments fused, the anterior segments being only indicated in the lateral and sometimes dorsal surfaces (Fig. 50). Frequently the cephalon is fused to peraeonal segment I.



Figure 48. Ventral view of food straining first peraeopods of Arcturid (After Grunner).

an vegeniculate but wluestigal



Figure 49. Peraeopod modified for food straining in Arcturus.

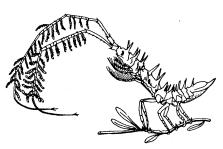


Figure 50. Arcturid with young (After Schmitt).

KEY TO GENERA AND SPECIES OF ARCTURIDAE

- 1a. Body geniculate; flagellum of antenna two more than three articles and usually longer than half length of last peduncular segment (Fig. 381) 62.

2a. Body cylindrical and of nearly uniform diameter; fourth peraeonal segment greatly longer than others; pleon with one free segment; pleotelson with posterior margin conically produced

Astacilla

Fig. 51. Astacilla lauffi Menzies and Frankenberg (5mm)

Range: Coast of Georgia (43 to 134 m)

Peraeonal segment IV of the male is about twice as wide as that of the female. Apparently this is characteristic of all species in the genus, but the females do not always have peraeonal segment IV as wide as it is in this species. One free pleonal segment is present.

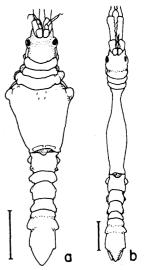


Figure 51: a. Astacilla lauffi (female). b. Male.

Fig. 52a,b. Astacilla grandulata (Sars) (16 mm)

Range: Newfoundland and Grand Banks (13 to 1170 m)

Fig. 52c. Astacilla caeca Benedict (9.5 mm)

Range: South of Martha's Vineyard (3337 m)

This species is blind and has a small rostrum on the frontal margin of the cephalon.

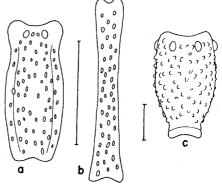


Figure 52. a-b. Astacilla grandulata (a. female, b. male). c. A. caeca.

 3a. With one free pleonal segment; peraeonal segment I not fused to cephalon; epimeres obscure in dorsal view Pleuroprion

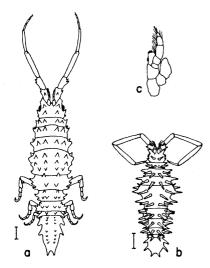


Figure 53. Pleuroprion. a. P. murdochi. b. P. intermedium. c. Maxilliped, P. intermedium.

Fig. 53a. Pleuroprion murdochi (Benedict) (12 mm)

Range: Off Pt. Frankland, Alaska

The pleotelson of this species is longer than wide and there are short spines on it.

Fig. 53b. Pleuroprion intermedium (Richardson) (7 mm)

Range: Kyska Harbor, Aleutian Islands (30 m)

The pleotelson of this species is about as wide as long and it has long spines on it.

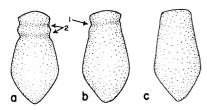


Figure 54. Pleonal sutures or indications of pleonal segments in pleotelson of Valvifera, a. Two. b. One. c. None.

4a. With two pleonal segments indicated in dorsal view of pleon (Fig. 54a); lateral lappet of peraeonal segment I not expanded beyond margin of bodyNeastacilla

Fig. 55. Neastacilla californica (Boone 1918) (6.1 mm)

Range: Southern California, Pt. Conception to Pt. Loma (Shallow water to 99m)

The species lives in sea weed. There are two pleonal segments indicated along with the conically produced pleotelson. This is the only species of the genus collected in North American waters.

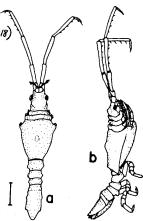


Figure 55. Neastacilla californica. a. Dorsal view. b. Lateral view.

Fig. 56. *Idarcturus allelomorphus* Menzies and Barnard_y(5.2 mm)

Range: Pt. Conception to Laguna Beach and Cortes Bank, southern California (13 to 91 m)

The species was collected in medium coarse gray sand on the shelf off southern California.

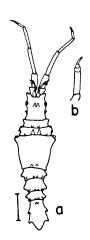


Figure 56. a. Idarcturus alle-Iomorphus. b. Detail antenna two.

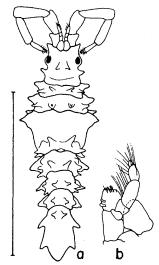


Figure 57. a. Idarcturus hedgpethi. b. Maxilliped.

Fig. 57. Idarcturus hedgpethi (Menzies /951) (1.4 mm)

Range: Tomales Bay, California (Intertidal)

This species differs from *I. allelomorphus* in that it is shorter and wider than *I. allelomorphus*. It was found on a hydroid.





Figure 58. Lateral furrows between cephalon and peraeon. a. Absent. b. Present.

- 5b. Shallow groove only between cephalon and first peraeonal segment (never laterally sutured) (Fig. 58a) Antarcturus

Fig. 59. Antarcturus floridanus (Richardson)

Range: Fernandina, Florida (494 to 499 m)

The species was originally described as a member of the genus Arcturus, but then Antarcturus was defined and the species was placed in that genus. The species differs from A. annaoides in that it has long spines on its body.

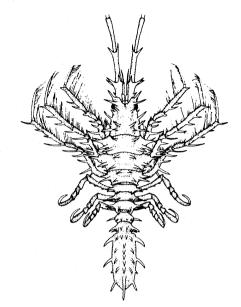


Figure 59. Antarcturus floridanus.

Fig. 60. Antarcturus annaoides (Menzies) (13 mm)

Range: Caribbean Sea, south of Jamaica (1244 m)

Antennae two are about 25 mm long or about twice the length of the body. The body is smooth; i. e., without large spines that are found in A. floridanus.

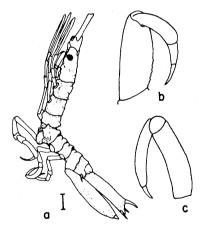


Figure 60. Antarcturus annaoides.

6a. Antennae two shorter than body; flagellum short with two to four articles; pleon never longer than last four peraeonal segments

Microarcturus

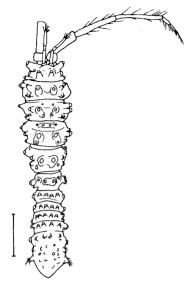


Fig. 61. Microarcturus tannerensis Schultz, v (5.5 mm)

Range: Tanner Canyon, southern California coastal shelf (1298 m)

This is the first species of this deep sea genus to be taken in North American waters. It is very much like an *Arcturus* species, but the species are blind and with short antennae two.

Figure 61. Microarcturus tannerensis.

6b. Antennae two longer than body; flagellum with more than four articles; pleon sometimes longer than last four peraeonal segments

Arcturus

GENUS ARCTURUS

The arcturids Arcturus and related genera, are among the most characteristic isopod of the deep sea. They are taken in abundance from the edge of the continental shelf to the deepest parts of the ocean. Food is probably obtained by filtering mud from the bottom where the isopod lives. The food particles are caught in the filter mechanism that is formed by folding the front legs together (Fig. 48).

The species live in cold and cool water. It is interesting to note that species caught off Florida are taken from the cold water at great depths; whereas, those sometimes of the same species taken off Greenland are taken from relatively shallow, but cold water. There are many species in the genus Arcturus and representatives are found in every ocean of the world. The genus Antarcturus was recently split from the parent genus Arcturus, and it is necessary to examine both old and newly described species in order to make sure that they are placed in the correct genus.

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7a.	Posterior margin of pleotelson notched (dorsal view)					
7b.	Posterior margin of pleotelson conically produced and without notch (dorsal view)9					
8a.	Body without spines					
8b.	Body with spines					

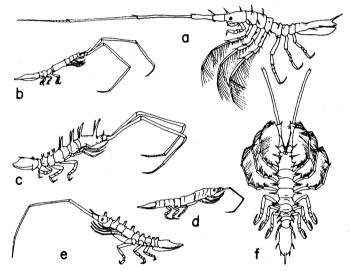


Figure 62. Arcturus. a. A. purpureus. b. A. glaber, c. A. longispinus, d. A. beringanus, e. A. baffini, f. A. caribbaeus.

9a.	Body with spines10
9b.	Body without spines
10a.	Pleotelson with long medial terminal spine projecting beyond

posterior margin 11

Fig. 62c. Arcturus baffini (Sabine) (40 mm)
Range: Greenland and northeastern Canada (9 to 274 m)

A subspecies without the large spines on the body has been called A. baffini tuberosus because it has tubercles instead of spines on the body.

- 11b. Large spines not present on pleonal segment 2. A. purpureus Fig. 62a. Arcturus purpureus (Beddard) (18 mm) Range: Sombrero Island, West Indies (813 m)

A character of er! IDOTEIDAE

Species of idoteids rarely have spines or especially long peraeopods. The flagellum of antenna one is of a single article. Antenna two has a very short or rudimentary flagellum or a long one of many articles. The mouth parts of the cephalon are always well developed and they are chewing and biting types. The peraeonal segments frequently have distinct coxal plates visible although different species have different numbers of them visible and some species have none visible. Peraeopods I to III are frequently different from peraeopods IV to VII and they never are specifically modified for food gathering or straining although they can be subchelate. There is generally one free pleonal segment and sometimes two or three. Frequently lateral indications of other segments are present, and the pleotelson is well defined. Antenna 2 with peduncle of 5 distinct articles.

KEY TO GENERA AND SPECIES OF IDOTEIDAE

1a. Eyes dorsally located; cephalon with notched anterolateral lobes; first three peraeopods subchelate; last four ambulatory

- 1b. Eyes laterally or sometimes dorsolaterally located; cephalon without anterolateral lobes; front of cephalon usually emarginate 8

Fig. 63a-c. Saduria entomon (Linaeus) (30 mm)

Range: Circumpolar, south on west coast to mid-California (On beach—north—to 813 m —south)

This species grows to about 30 mm or more and is a characteristic resident of the continental shelf in the Arctic regions. It is also taken from the mud and sand of the bottom in small bays and inlets in brackish and occasionally freshwater. Occasionally it is found on the beach buried in the sand or under rocks. This species is generally taken on sand and gravel bottoms and beaches. The specimens from mid-California were taken in

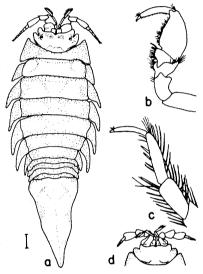


Figure 63. Saduria. a. S. entomon. b. Peraeopod I. c. Peraeopod VI. d. S. sabini.

the cool deep water, and it is limited in depth probably more by temperature than pressure. The genus has been called *Mesidotea*.

Fig. 63d. Saduria sabini (Kroyer) (20 mm)

has Ant 2 peduncular Range: Circumpolar (On beach to 139 m)

This species is blind and found on muddy bottoms and in the mud of beaches. It is possible that the species has lost its eyes because of lack of light in its muddy habitat. Its eyed elative S. entomon lives in a sand and gravel habitat. This species has been divided into a number of subspecies, but they are disregarded here because many of them are found outside of the range represented in this book.

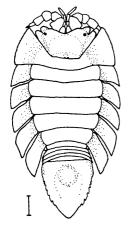


Figure 64. Saduria sibirica.

Fig. 64. Saduria sibirica (Birula) (8.8 mm) Range: Siberia to northeastern Alaska near Canada (7 to 50 m)

GENUS CHIRIDOTEA

The species of the genus are common on the sandy bottoms off the Atlantic coast of North America. They are occasionally found several centimeters below the sand on intertidal beaches. The palp of the maxilliped has three articles and the mandible lacks a molar process. The type-species of the genus of six species is C. coeca.

Burnow along just under the surface, leaving a trail of raised sediment.

KEY TO SPECIES OF CHIRIDOTEA

3a.	Prominent :	anterolateral	groove,	indentation	or	cut	on	cepha-
	lon							4

 Fig. 65a-c. Chiridotea stenops (Menzies and Frankenberg) (2.3 mm)

Range: Off Georgia coast (104 m)

This species was shown by Watling of Maurer (1975) to be a jr. synonym of <u>C</u>. <u>arenicola</u> (a juvenile stage)

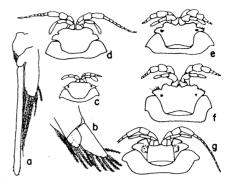


Figure 65. a-c. Chiridotea stenops. a. Pleopod 2, male stylet. b. Apex of uropod (endopod and exopod). c. Cephalon. d. C. almyra. e. C. nigrescens. f. C. arenicola. g. C. tuftsi.

Fig. 65g. Chiridotea tuftsi (Stimpson) (6.5 mm)

Range: Gulf of St. Lawrence to Long Island Sound (Low tide to 55 m)

Fig. 65d. Chiridotea almyra (Bowman) (5.8 mm)

Range: Cape Cod to Georgia (Shallow water)

This species has eyes, but they are difficult to see in preserved specimens. The species is exclusively found in brackish water.

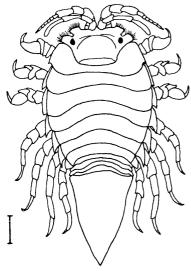


Fig. 66. Chiridotea coeca (Say) (13 mm)

Range: Nova Scotia to Florida (Surface to 31 m)

Figure 66. Chiridotea coeca.

7a.	Antenna one extended slightly beyond end of peduncle of an tenna two; one or more setae on anterior lobe of frontal margin of cephalon
	Fig. 65e. Chiridotea nigrescens (Wigley) (10.5 mm)
	Range: Southern and western shores of Cape Cod (Near surface to 1.5 m)
7b.	Antenna one not quite reaching to end of peduncle of antenna two; no setae on anterior lobe of cephalon
	Fig. 65f. Chiridotea arenicola (Wigley) (7.5 mm)
	Range: Georges Bank (27 to 66 m)
8a.	Flagellum of antenna two long and well developed (Fig. 67)
8b.	Flagellum of antenna two rudimentary to 3 articles (Fig. 102c)
9a.	Palp of maxilliped with four or five articles; various number of coxal plates visible in dorsal view
9b.	Palp of maxilliped with three articles; no coxal plates visible in dorsal view

GENUS SYNIDOTEA Harger 1878

This genus contains many species all of which are superficially alike and can be told apart, if at all, by making observations of minute or qualitative characters. The species are mostly found on the west coast of North America south to southern California, but they are circumpolar in distribution and are found on the east coast south to Cape Cod. see Menzies & Miller (1972) for a key to the California Synidotea, a a review of world species.

KEY TO SPECIES OF SYNIDOTEA

10a.	Pleotelson with posterior margin notched
10b.	Pleotelson with posterior margin rounded or pointed
lla.	Two small spines or two spines with ridge base mediolaterally placed on frontal margin of cephalon12
11b.	Cephalon without spines on frontal margin
12a.	Spines on frontal margin of cephalon on ridges
12b.	Spines on frontal margin of cephalon not on ridges14
13a.	Pleon about as wide as long; cephalon about 2/3 as wide as body

Fig. 67. Synidotea bicuspida (Owen) (25 mm)

Range: West Coast of Alaska and Bering
Sea (6 to 148 m), Originally described as

Idoteo bicuspida. Synunomized by Hanger
in 1879. Circum polar; S. to Bodega Bay.

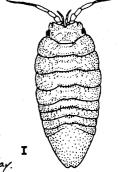


Figure 67. Synidotea bicuspida.

13b. Pleon shorter than long; cephalon less than 2/3 width of body S. marmorata

<u>Synidotea media</u> Iverson, 1972. California

<u>Synidotea spinosa anadyrensis</u> Gurjanova 1955. Bering Sea.

<u>Synidotea berolzheimeri</u> Menzies & Miller 1972. Morro Bay to Sanoma Co.

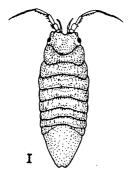


Figure 68. Synidotea marmorata.

Fig. 68. Synidotea marmorata (Packard) (19 mm)

Range: Laborador and Grand Banks (22 to 236 m)

This species and the smaller S. nodulosa are the only two species of Synidotea on the east coast of North America.



Figure 69. Synidotea ritteri.

Fig. 69. Synidotea ritteri Richardson 1 (6 mm)
Range: San Francisco, California (Shallow water) to

Vancower Ls



Fig. 70. Synidotea pettiboneae Hatch (10.5 mm)

Range: Puget Sound, Washington (27 to 55 m), to San Mateo Ca (Calif.)

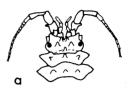




Figure 70. a. Synidotea pettiboneae, anterior part. b. Posterior part.

16b. With no medial dorsal tubercles on dorsum of pleon; lateral edges of peraeonal segments broadly roundedS. pallida

Fig. 71. Synidotea pallida Benedict $_{\Lambda}$ (12

Range: Chirikof Island, Alaska (1271 m)

Arctic; Bering Sea

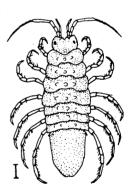
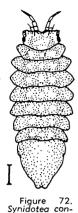


Figure 71. Synidotea



solidata.

= S. pettiboneae (See menzies & Miller, 1972:25)

Fig. 72. Synidotea consolidata (Stimpson) (8 mm) Range: Pacific Grove, California (Shallow water)

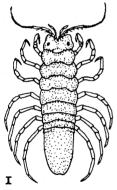


Figure 73. Synidotea

Fig. 73. Synidotea erosa Benedict A (21 mm)

Range: Sannakh Island, Alaska (11 to 59 m) (869 m)

1897

S. nebulosa

Fig. 74. Synidotea nebulosa Benedict A (12 mm)

Range: Aleutian Islands, Alaska (11 to 59 m)

Bering See to Washington State

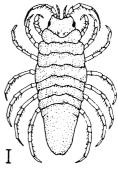


Figure 74. Synidotea

Fig. 75. Synidotea laticauda Benedict $_{\Lambda}$ (17.5 mm)

Range: San Francisco Bay, California (Shallow water) Presently restricted to
San Francisco Bay, although claims to
have collected it in Japan. Probably

synonomous with 5. harbordi.

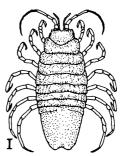


Figure 75. Synidotea

20a. Cephalon with shallow medial frontal groove; body with continuous lateral margins tapering towards pleon S. hartfordi

Fig. 76. Synidotea hartfordi Benedict (16 mm)

Range: Southern California to northern Baja.

Galifornia

- Iverson has collections from Japan
- Menzies of Miller report it from Jayon
- We have collections from Pt. Chivato & Guaymas.

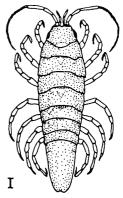


Figure 76. Synidotea

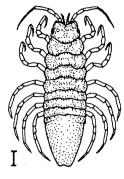


Fig. 77. Synidotea angulata Benedict A (12 mm)

Range: Washington State (57 to 69 m)

Figure 77. Synidotea angulata.

21a. Pleotelson with posterior margin rounded
22
21b. Pleotelson with posterior margin pointed
23
22a. Body margins almost parallel; peraeonal segments all of

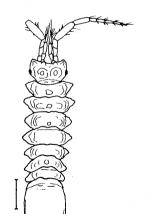


Fig. 78. Synidotea calcarea Schultz & (6 mm)

Range: Tanner Canyon on shelf off southern California (813 m)

Santa Rosa (yn. (?) 54 m

, pleutelson actually subacute

Figure 78. Synidotea alcarea.

22b. Body margin widest at peraeonal segment III; peraeonal segment III both longer and wider than peraeonal segment VII

S. magnifica

Fig. 79. Synidotea magnifica Menzies and

Range: Pt. Conception to Oceanside, California (55 to 91 m)



Figure 79. Synidotea magnifica.

or wo

Barnard v (6 mm)

23a. Body with short spines or large tubercles 24, 25

23b Bodywith mashort spines or large tubercles 1110 1250

Fig. 80. Synidotea muricata (Harford) (22 mm)

Range: Icy Cape, Greenland (46 m)

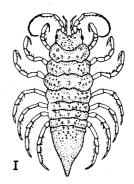


Figure 80. Synidotea muricata.

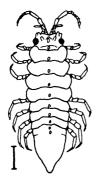


Figure 81. Synidotea nodulosa.

Fig. 81. Synidotea nodulosa (Krøyer (7.5 mm)

Range: Circumpolar, south to Georges Bank and British Columbia (11 to 203 m)

Sometimes the parasitic isopod *Clyptoniscus* meinerti (p. 339) is found in the marsupium of the female.

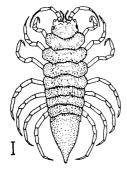


Figure 82. Synidotea laevis.

Fig. 82. Synidotea laevis Benedict (13 mm)

1897

Range: Bering Sea near Pribilof Islands (54 to 66 m)

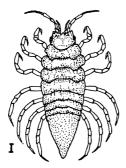
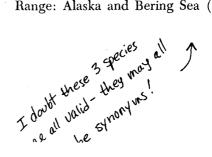


Figure 83. Synidotea picta.

IST Fig. 83. Synidotea picta Benedict, (16 mm)

Range: Alaska and Bering Sea (9 to 37 m)



MARINE ISOPOD CRUSTACEAN

Key to the Species of Idoteidae Know

1. Pleon composed of more than one dis

KEY TO BOREAL SPECIES
PLEOTELSON N

Note: these 4 species are all ymous, at least picta, not

- 1. Anterior part of pleon witubercles or spines on permargin of cephalon with do of Ant. 2 with more than
 - Anterior part of pleon witubercles or spines on pervery low and weak; frontato modest incision or not than llarticles
- 2. Median tubercles of pereo region of cephalon with t 2 flagellum with more tha
 - Median tubercles of perec posterior region of cepha antenna 2 flagellum with
- 3. Median tubercles of perec about 3 times width; from incision or notch; antennand extended to middle of
 - Median tubercles of peres body length equal tao abo of cephalon without a dis may be concave; antenna extended to middle of per

on drifting seaweeds throughout the Atlant occurrance in the northeast Pacific is not t



Fig. 81. Synidotea nodulosa (Krøyer (7.5 mm)

I doubt valid synonym

VI to VII in Colidater

	Key to the Species of Idoteidae Known from the Gulf of California
_	Pleon composed of more than one distinct segment
2.	Pleon composed of 4 segments, plus 1 pair of partial suture lines; flagellum of antenna 2 uniarticulate Cleantioides occidentalis
	Pleon composed of 3 segments, plus 1 pair of partial suture lines; flagellum of antenna 2 multiarticulate
3.	Maxillipedal palp of 4 articles Idotea (Idotea) urotoma
	Maxillipedal palp of 5 articles
4.	Eyes transversely (dorsoventrally) elongate and narrow; maxilliped with 1, 2, or 3 coupling hooks Idotea (Pentidotea) stenops
_	Eyes not transversely elongate and narrow; maxilliped with 1 cou-
	pling hook 5
5.	Posterior border of pleotelson strongly concave; frontal process
	extended beyond frontal lamina 1 Idotea (Pentidotea) resecuta
	Posterior border of pleotelson convex, with small median lobe;
	frontal process not extended beyond margin of frontal lamina 1 6
6.	Length less than 3.7 times width; eyes reniform; males with distinct
	tufts of setae on pereopods Idotea (Pentidotea) wosnesenskii
-	Length more than 3.7 times width; eyes circular; males without
_	tufts of setae on pereopods Idotea (Pentidotea) aculeata
	Flagellum of antenna 2 of a single article
•	Flagellum of antenna 2 multiarticulate
٥.	Lateral margins of pleon expanded posteriorly; pleon without suture lines Erichsonella cortezi
	Lateral margins of pleon smooth and gently convex; pleon with one
_	pair of partial anterolateral suture lines
9	Maxillipedal palp of 4 articles; cephalon with a mediodorsal hump;
٠.	coxal plates of pereonite 6 distinct in dorsal view
	Eusymmeri s antennatus
	Maxillipedal palp of 3 articles; cephalon without mediodorsal
	hump; coxal plates of pereonite 6 NOT distinct in dorsal view
	Parasymmerus annamaryae, n. sp.
10.	Maxillipedal palp of 4 articles; posterior margin of pleotelson acu-
•	minate
_	Maxillipedal palp of 3 articles; posterior margin of pleotelson con-
	cave Synidotea harfordi

Note: The Atlantic species *Idotea metallica* has not previously been reported from the northeastern Pacific; we have, however, a record of a single specimen from the Gulf of California and a second individual, collected by M. Ninos, from Catalina Island, California. This species is commonly found on drifting seaweeds throughout the Atlantic Ocean and elsewhere, and its occurrance in the northeast Pacific is not totally unexpected.

[1957] clarification of the genus Edotea in this regard). Hence, it is apparent from the literature that various workers have interpreted the concept of a "coxal plate" in a variety of ways, some considering a slightly swollen coxa as representing a coxal plate, others assuming lack of an obvious suture line meant the plate had fused with its respective pereonite (when in fact the

Mackey for the collection of important materials used in this paper, and other studies; Melinda Thun, for illustrating the dorsal views of S. harfordi, P. annararyae, and C. occidentalis; Bob Setzer and Dr. pavid Young, for algal identifications; S. Luke, for assistance in obtaining pan materials from Scripps Inditution; and Ruth Toyama, for typing the manuscript.

This research was supported, in part, by grants from the National Science Foundation. This is contribution No. 368 of the All Hancock Foundation.

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- 26a. Coxal plates visible in dorsal view on peraeonal segments II to VII or on only IV to VII; maxillipedal palp of four articles

GENUS PENTIDOTEA

The medium to large species of this genus are fairly common and are frequently found between the tides on rocks and in seaweed, kelp, eelgrass and other marine vegetation. Apparently their color is changeable and varies according to the background on which they are located. The species of the genus have been considered as a subgenus (*Pentidotea*) of *Idotea*, but since there is a consistent difference in maxillipedal palp article number between species of *Pentidotea* and those of *Idotea*, it is here considered a full genus.

KEY TO SPECIES OF PENTIDOTEA

- 27a. Posterior margin of pleotelson with single medial protuberance 28

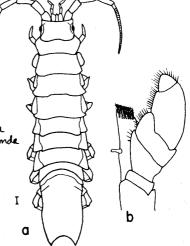
Fig. 84. Pentidotea resecta (Stimpson) (36 mm)

Range: Southeastern Alaska to Punta southern Galifornia (Surface Banda to 18 m)

- Records so, of Pt. Banda are uncommon but it apparently drifts on scaweed a lot to how been collected from La Paz & the

mouth of the Gulf, as well Figure 84. a. Pentidotea resecta. b. Maxilliped.

as some of Wexicos offshore islands



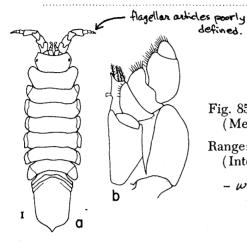


Fig. 85. Pentidotea schmittii (Menzies) (30 mm)

Range: Dillon Beach, California (Intertidal) to Ponta Banda

- we also have a record from Washington State

Figure 85. a. Pentidotea schmittii. b. Maxilliped.

28b. Cephalon moderately to deeply set into peraeonal segment I; eyes laterally placed on cephalon 29

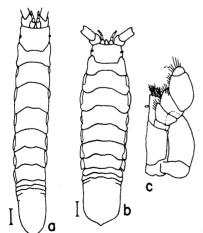


Fig. 86. Pentidotea montereyensis (Maloney) (14.3 mm)

Range: Washington to Monterey

Bay California (Intertidal)

Figure 86. a. Pentidotea montereyensis (male). b. Female. c. Maxilliped.

29b. Body margins somewhat convex and pleon noticeably narrower than widest body part; lateral ends of peraeonal segments slightly to moderately separated at ends; posterior margin of pleotelson moderately to strongly produced 30

30a. Pleon as narrow or narrower than cephalon with posterior margin strongly produced P. aculeata

Fig. 87. Pentidotea aculeata (Stafford) (23 mm) No. Calif. to lower Gulf & Range: Central to continue, California (Tide pools)

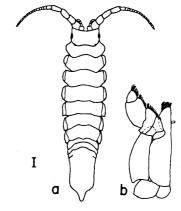


Figure 87. a. Pentidotea aculeata.

30b. Pleon wider than cephalon with posterior margin moderately produced 31

31a. Lateral edges of free pleonal segments separated and moderately pointed; posterolateral margin of pleon rounded P. wosnesenskii

Fig. 88. Pentidotea wosensenskii (Brandt) (32 mm)

Range: Western Aleutians to San Francisco Bay (Surface to 16 m)

> La Paz (USNM #69575) Gulf of Alaska (rody littoral)-common

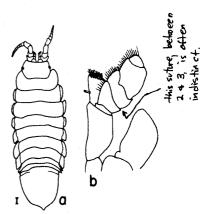


Figure 88. a. Pentidotea wos-nesenskii. b. Maxilliped.

Pentidotea Kirchanskii Miller & Lee, 1970 Central Calif. (N. of San Francisco) Resemble c 7 newlesta

31b. Lateral edges of pleonal segments more or less continuous with lateral body margin; posterolateral margin somewhat quadrate P. stenops

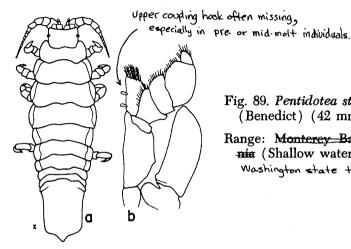
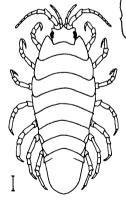


Fig. 89. Pentidotea stenops (Benedict) (42 mm)

Range: Monterey Bay, California (Shallow water) Washington state to La Paz

Figure 89. a. Pentidotea stenops. b. Maxilliped.

32a. Coxal plates visible only on peraeonal segments V to VII Colidotea



Dorsal coxal plates exist on II-VII, but anterior ones are small and situated ventrally, visible in lateral view; II-VII or III-VIII only visible indorsal view.)

Fig. 90. Colidotea rostrata (Benedict) (12 mm)

Range: San Pedro, California (Shallow water) to Punta Eugenio.

This species is the type and only one in the genus. No 7

C. edmondsoni Hiller 1940 (Hawaii)

C. findleyi Brusca & Wallerstein 1977

conversal or thousand occurreting (mest wexico) .C. wallowsteini Brusca 1983 (NW nexico)

32b. Coxal plates visible on peraeonal segments II to VII

GENUS IDOTEA

The species of this genus are among the most common isopods found in the algae in near shore habitats. Many of them are very widely distributed since they are easily transported by ships. The species are of economic importance because they are frequently encountered in the stomachs of commercial fish.

KEY TO SPECIES OF IDOTEA

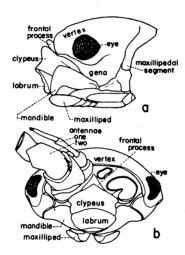


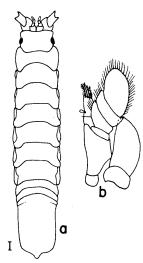
Figure 91. Nomenclature of cephalon of Valvifera. a. Lateral view. b. Frontal view.



Figure 92. Idotea phosphorea.

Fig. 92. *Idotea phosporea* (Harger) (21 mm) Range: Gulf of St. Lawrence south to Cape C

Range: Gulf of St. Lawrence south to Cape Cod (Surface to 33 m)



(Idotea)

Fig. 93. Idotea fewkesi Richardson (42 mm)

Range: Alaska to southern California (Shallow water)

This species is very common and has frequently been described as a new species.

Figure 93. a. Idotea fewkesi. b. Maxilliped.

Fig. 94. Idotea balthica (Pallas) (20 mm)

Range: Atlantic coast of North America from Gulf of St. Lawrence to Rio de Janeiro (Surface to 218 m)

This species is common in floating or attached seaweed, in eelgrass and in sand and gravel in the bays and inlets along the entire coast of North America (and Europe). It is sometimes taken in quantity from the stomachs of fish, especially smelt, so it is apparently of some economic importance and also imporant in the ecosystem. The species comes in many color forms sometimes being quite brightly colored.

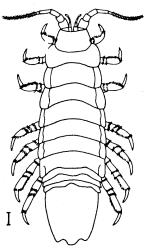


Figure 94. Idotea balthica.

36b. Epimeral sutures obliquely cut acrossed anterolateral margins of peraeonal segments II, III and IV; lateral margins of pleotelson concave

I. ochotensis

(Idotea)

Fig. 95. Idotea ochotensis Brandt (42 mm)

Range: Northwestern North America to San Francisco Bay (Surface to 33 m)

This species is perhaps a synonym of I. fewkesi.

- I. ochotensis aluetica Gurjanova
- <u>I. ochotensis ochotensis</u> Brandt (includes <u>I. (I.) derjugini)</u>



Figure 95. Idotea ochotensis.

- 37a. Posterior margin of pleotelson truncate or ending in small posterior protuberance or obtuse angle 38
- I. obscura Rafi, 1972 Queen Charlotte Is. N. to Prince

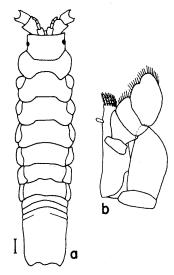


Figure 96. a. *Idotea rufescens*. b. Maxilliped.

(Pentidotea)

Fig. 96. Idotea rufescens (Fee) (15 mm)

Range: British Columbia to central California (Intertidal to 82 m)
(So. of Carmel, Monterey Co.)

= I. resecuta by Menzies & waidzunas (1948), but not by Menzies (1950)

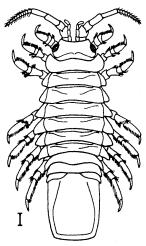


Figure 97. Idotea metalica.

Fig. 97. Idotea metallica (Bosc) (18 mm)

Range: Greenland, Nova Scotia to Straits of Magellan (South America) (Surface to 166 m)

This species is common and widespread and is frequently found on floating seaweed and is also taken from attached seaweeds along the east coast of North America and Europe.

USUM has a specimen (1) from the Gulf of California, & we have I from Catalina Island! 39a. Posterior margin of pleotelson ending in obtuse angle.....

.....I. rectilinea

Fig. 98. Idotea rectilinea (Lockington) (17 mm)

Range: Mid-California to northern Baja California (55 to 73 m)

This species has been taken on sandy and muddy bottoms.

= <u>I. urotoma</u> (see Menzies, 1950: 166)

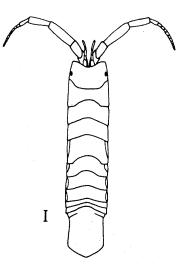


Figure 98. Idotea rectilinea.

39b. Posterior margin of pleotelson ending in small protuberance 40

Fig. 99. Idotea urotoma Stimpson (17.5 mm)

Range: Puget Sound, Washington to southorn California (Intertidal)

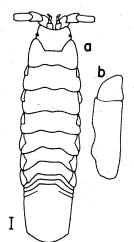


Figure 99. a. Idotea urotoma. b. Uropod or



gracil-

Idotea

lima

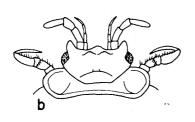
Fig. 100. Idotea gracillima (Dana) (11 mm)

Range: Coast of California

= <u>I</u>. montereyensis (see Meuries, 1950: 185)

of 1 large article & 1-2

Antenna two with



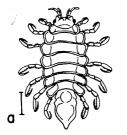


Figure 101. a. Edotea montosa. b. Detail anterior part.

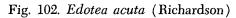


Fig. 101. Edotea montosa (Stimpson) (9 mm)

Range: Long Island to Nova Scotia (15 to 46 m)

The posterior margin of the pleotelson is produced in this species.

= E. triloba



Range: Off New England coast (188 m)

This species was taken from the stomach of a cod. It has knob-like anterolateral corners on the cephalon.

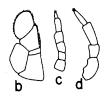
= E. triloba



Fig. 103. Edotea triloba (Say) (7 mm)

Range: Maine to New Jersey (Surface to 1 m) S.C.

This species was found in muddy water and eelgrass. There are no indications of a second pleonal segment in the species.



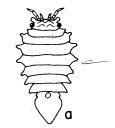


Figure 102. a. Edotea acuta. b. Maxilliped. c. Antenna two. d. Antenna one,

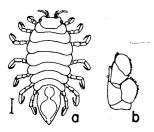


Figure 103. a. Edotea triloba. b. Maxilliped.

Fig. 104. Edotea sublittoralis (Menzies and Barnard) (4 mm)

Range: Pt. Conception, California to northern baja (14 to 64 m)

This species has two small tubercles on the frontal margin of the cephalon.

Barry & Crnie collected E. sublittoralis at Compo Poewee - summer, 1979 (4 specimens)



Figure 104, Edotea sublittoralis.

41b. Antenna two longer than antenna one; flagellum of one to three articles; palp of maxilliped with four articles 42

42a. Pleon with 2 to 3 free segments with indications of the presence of 3 or 4; flagellum of two or three articlesCleantis

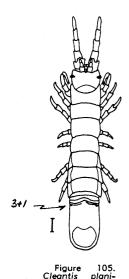


Fig. 105. Cleantis planicauda (Benedict) (15 mm)

Range: Georgia to Florida also Pureto Rico (Beach to 44 m) of tropical E. Pacific (see Brusca & wallerstein, 1979)

This species has been collected on a sandy bottom on the Georgia coast. It has a broadly rounded posterior margin on the pleotelson.

The genus Cleantis has:

- 1 mab lacks palp
- @ wroped of 1 or 2 articles
- 3 Mxp. palp of \$ # 5 articles



caudata.

plani-

Fig. 106. Cleantis heathii (Richardson) (8 mm)

Range: Monterey Bay, California (Shallow water)

There are posterolateral corners on the pleotelson of this species.

= Idotea urotoma (see Mouzies 1950:166)

Figure 106. Cleantis heathii.

Fig. 107. Cleantis occidentalis Richardson (7.5 mm)

Range: Magdalena Bay, Baja California (22 m)

The posterior borders of the pleotelson are rounded in this species.

= <u>Cleantioides</u> <u>occidentalis</u> (Rich. 1899a) by Kensley & Kaufman, 1978



43a. Pleotelson smooth and rounded; indications of one pleonal segment Eusymmerus

Fig. 108. Eusymmerus antennatus (Richardson) (8 mm)

Range: Abreojos, Baja California to (10 m) So, Mexico

This is the type and only species in the genus.

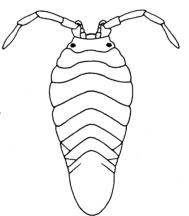


Figure 108. Eusymmerus antennatus.

The correct authorship is:

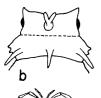
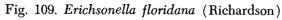




Figure 109. a. Erichsonella floridana. b. Detail anterior part.



Range: Florida (Below low tide)

This is the only species of the genus with spines on the dorsum.

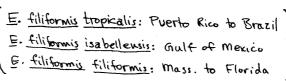


Fig. 110. Erichsonella filiformis (Say) (8 mm)

Range: Massachusetts to Florida to Texas including Bahama Islands (8 to 33m)

The species is common in the eelgrass beds of the shallow bays and estuaries within its range. The subspecies, E. filiformis filiformis (Menzies) and E. f. isabelensis (Menzies), have been described. E. f. išabelensis is much narrower and found further west than E. f. filiformis.

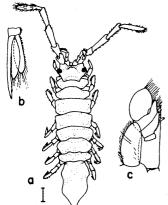


Figure 110. a. Erichsonella filiformis. b. Male pleopod 2. c. Maxilliped.

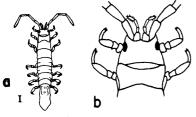


Figure 111. a. Erichsonella attenuata. b. Detail anterior part.

Fig. 111. Erichsonella attenuata
(Harger) (12 mm)
Range: Connecticut to North
Carolina (Below low tide)
The species is taken in eelgrass (Zostera) and in coral. Its
body is much broader in proportion than that of E. filiformis.

AHF has a specimen from Corpis
Christi (Texas) collected by m.K.
wicksten; from Thallassia.

= Stenulata: Holotype (d), no 5 -s on percon.

MARINE ISOPOD CRUSTACEANS

Erichsonella cortezi Brusca & Wallerstein 1977

Fig. 112. Erichsonella crenulata (Menzies) (23 mm)

Range: Newport Bay, California (Shallow water)

The species was taken from eelgrass (Zostera). We have a specimen from Bahía San Quintin with small lobes on the legs as in E. pseudoculata.

All 3 species appear very similar (= siblings).

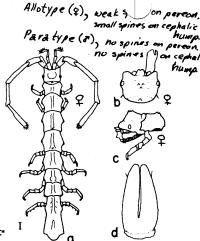


Figure 112. a. Erichsonella crenulata. b. Detail, cephalon. c. Lateral view cephalon. d. Penis.

Fig. 113. Erichsonella pseudoculata (Boone) (9 mm)

Range: Pt. Conception to La Jolla, California (Surface to 18 m)

The species was first taken from eelgrass at San Pedro, California. Once it was the only species in the genus Ronalea, although it was originally described in Erichsonella. (Ronalea was

synonymized with <u>Etichsonella</u>; it was originally estab. on the basis of the

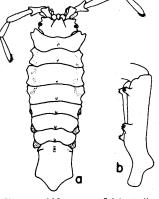


Figure 113. a. Erichsonella pseudoculata. b. Basis of peraeopod VII.

partial suture on the plean of E. <u>escudoculata</u>, a character since shown

SUBORDER ANTHURIDEA to be variable in the

ANTHURIDAE

Genus!

Species of anthurids are placed in only one family, the Anthuridae. The isopods are medium to small in size. Some species common in the near shore habitats can reach to about 25 mm long for mature individuals, but most specimens average about half that size. Species found in the deep sea can be quite small. Anthurids live on the bottom where they are generally found in tubular burrows. Mostly the burrows are of their own construction although they are at times found in the tubes of other animals, especially those of marine worms. They are also frequently collected from

encrusting animals and algae on piles or rock jetties, but they are usually found only on the side where the water is relatively calm. There are some species that have been taken exclusively in near shore caves, and they are blind like their deep sea relatives.

At times the two sexes have been described as different species. The anthurid species are different from other isopods in that they have extremely elongate bodies and a peculiar kind of caudal fan, both of which are adaptations for living in holes in the mud or sand

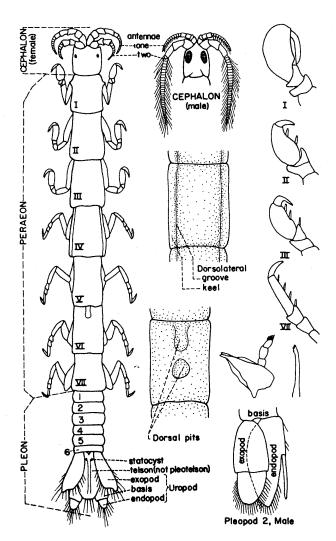


Figure 114. Anthuridea (General nomenclature).

of the bottom. Specimens of anthurids are most frequently females, males being relatively rare. Apparently all animals are female at first (protogynous) then some change to males. The male can be identified by the very long, sometimes quite bushy antennae, and by the presence of much larger eyes than those of the female. Some males have more than one set of eyes also.

The number of articles in the flagellum of the antennae can vary from few to many according to species as well as sex. The mandible and mandibular palp are generally normal although there is a loss of mandibular palp articles or complete loss of the structure in some species. The structure of the maxilliped is almost specific to genus (Fig. 115), but there are several genera that cannot be distinguished by the maxillipedal configuration alone. The number of palp articles varies from none to five. Species with piercing

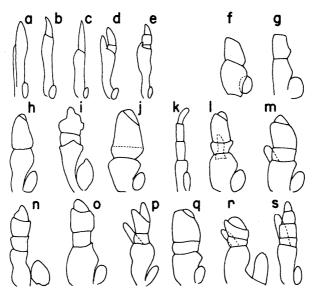


Figure 115. Maxillipeds of Anthuridea. a-e. Piercing and sucking. a. Neoanthura. b. Leptanthura. c. Paranthura. d. Calathura. e. Accalathura. f-s. Biting. f. Anthura. g. Ptilanthura. h. Cyathura. i. Xenanthlura. j. Haliophasma. k. Eisthistos. l. Anathura. m. Apanthura. n. Holoanthura. o. Mesanthura. p. Hyssura. q. Skuphonura. r. Bathura. s. Panthura.

and sucking mouth parts have pointed maxillipeds without or with only a few articles. Those with normal or biting mouth parts generally have one to five palp articles, the usual number being less than five.

The shape of the peraeonal segments is in keeping with the general body configuration and it is long and thin in most instances, although at times peraeonal segment VII is quadrate or shorter than wide or does not show at all in the dorsal view. The peraeopod number is usually seven when there are seven peraeonal segments, and the anterior peraeopods are subchelate. In many instances peraeopod I is strongly subchelate and all other peraeopods. especially I to III, are less so until peraeopod VII which is more or less ambulatory. Occasionally in some species the male will have a very large subchelate peraeopod I and the female will have peraepod I only slightly modified. On the peraeonal segments there are sometimes very strongly developed lateral ridges or grooves. At times they are called keels or lateral leels and sometimes they are called dorsolateral grooves. In many species they are reduced in size or absent. There are dorsal pits on the dorsum of the peraeonal segments of some species and they can be either located medially (generally anterior on the segment) or paired (generally posterior on the segment). The peraeonal segments are never fused to the cephalon, the pleon or to each other. Oostegites are present on the female on either segments II to V or III to V.

The pleonal structure varies both according to species and according to sex. In some species all segments including the sixth are distinct; in others there are only marginal sutures and in others there are no external indications of segments. The pleopods also vary there being two general types. In the first type the anterior or first set of pleopods is indurate and operculate to the others. In the second all pleopods are about the same there never being an operculate set of pleopods. The second male pleopod is sexually modified and is a good guide to species although it unfortunately has not been well described for most species. Pleonal segment six in all instances has two large generally oval and sometimes spoonshaped rami, the uropods. The telson itself is generally elongate or oval, and, indeed, the posterior margin and the cross sectional view of the telson has many different shapes including one with a mediodorsal ridge. Generally long non-plumose marginal setae are present on the exopod, endopod and telson, and they apparently serve to anchor the specimens in the tubes in which they dwell. There are special organs called statocysts and found only in anthurids present on the anterior half of the telson in many species. They are places where muscles are attached and perhaps have some sensory function in regard to body position. A pair of them is most often present, but in some instances they apparently fuse on the midline to form a single statocyst.

The anthurids are one of the most economically important isopod groups and the species serve a function in their environment not unlike that of the earthworm in its terrestrial habitat. At times

FOR KEY TO CAMPTITE ANTWINIS

SEE Schultz (1977). Keys to

world genera see Poore 89 (A.B., 1987)

they are found in great abundance and frequently many square miles of bottom will be covered by their burrows. Man is only beginning to understand the near-shore ecological habitats of which the anthurids are an important part. They deserve much more study. There are about 20 genera and 37 species in North American waters. Some are not too well defined. They can be easily confused with species of the order Tanaidacea from which they differ in the type of peraeopod I (in tanaids they are truly chelate, not subchelate—Fig. 6) and in the configuration of the pleotelson.

KEY TO GENERA AND SPECIES OF ANTHURIDAE

- 1a. Six large peraeonal segments visible in dorsal view of mature specimen (+ minute 7th segment, w/o legs) 2
- 1b. Seven large peraeonal segments visible in dorsal view of mature specimen 3

GENUS COLANTHURA

The species of this genus and the single species of the genus *Neoanthura* differ from other anthurids in that there are six rather than seven elongate, distinct peraeonal segments visible in the dorsal view. Peraeonal segment VII is minute and located between the pleon and peraeonal segment VI. The mouth parts of the species of both genera are modified for piercing and sucking.

Schultz (1977) synonomized this with Paranthura infundibulata from Bermuda.

Fig. 116. Colanthura tenuis (Richardson) (4 mm)

Range: Bermuda (Shallow water)

This species is the type species of the genus. It is very much like the Pacific coast species of the genus from which it differs only in the shape of the peraeonal segments.

note: Schultz (1977) supressed the name

Colanthura, replacing it with Califanthura.

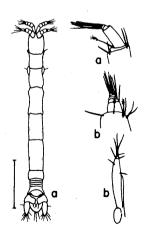


Figure 116. a. Colanthura tenuis. b. Antenna one. c. Antenna two. d. Maxilliped.

This was in error and was reported by Poore (1980). But, take a press) is now resurrecting california are well be back to be

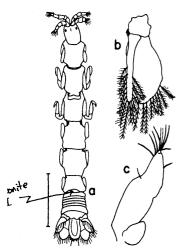


Figure 117. a. Colanthura squamosissma. b. Male pleopod 2. c. Maxilliped.

Fig. 117. Colanthura squamosissima Menzies (4.4 mm)

Range: Coastal shelf, Pt. Conception to Mexican border (18 to 91 m)

Also in Gulf of California.

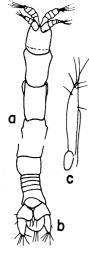


Figure 118. a. Neoanthura coeca, anterior part. b. Posterior part. c. Maxilliped.

Fig. 118. Neoanthura coeca (Menzies) (3.8 mm) Range: Caribbean Sea south of Jamaica (1244 m)

The species is the type and only one in the genus. The species is blind and pigmentless and all pleonal segments are distinct. The maxilliped consists of the endite only.

Colanthura tonvis (Caribbean species) occurs in

Fig. 119. Skuphonura laticeps (Barnard) (7 mm) Range: Lesser Antillies (7 to 37 m)

The broad cephalon sets the species apart from other species of anthurids. The species is the type and only one in the genus. Dorsolateral keels and dorsal pits are absent. The segments of the pleon are fused and without lateral sutures. The telson is shorter than the pleon, thin, dorsally flattened and with small paired statocysts (Fig. 120a). Pleopod one is operculiform. The maxillipedal palp has three articles (Fig. 115g).

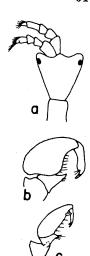


Figure 119. a. Skuphonura laticeps, dorsal view of cephalon. b. Peraeopod I. c. Peraeopod II.

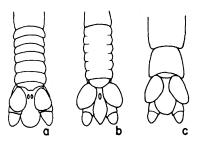


Figure 120. Pleon and statocyst types. a. Segments distinct and paired statocysts. b. Marginal sutures only present and statocyst single. c. No segments or statocysts distinct.

 5b. Pleopod 1 not operculate (Fig. 121b); telson acutely pointed and long or concave and shorter than exopod and endopod ... 6

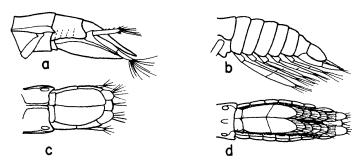


Figure 121. Pleopod I types. a. Operculate. b. Not operculate. c-d. Ventral views.

- 6a. Blind; maxillipedal palp of three articles; mandibular palp of three articles; telson long and pointed (Fig. 127)

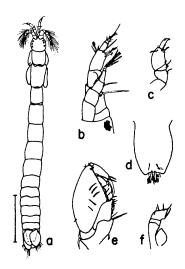


Figure 122. a. Xenanthura brevitelson (male). b. Antenna one and two, female. c. Peraeopod VI, male. d. Telson. e. Peraeopod I, male. f. Maxilliped.

Fig. 122. Xenanthura brevitelson (Barnard) (4.5 mm)

Range: Off Georgia Coast and West Indies (18 to 133 m)

The species is remarkable in that each body segment is distinct and there is no marked difference between the peraeon and the pleon except in the length of the segments. The species is the type and only species of the genus. The female has shorter antennae than the male and only one pair of eyes.

7a. Exopods of uropods with spines on inner margins (Fig. 123)

Horoloanthura

Fig. 123. Horoloanthura irpex (Menzies and Frankenberg) (3.7 mm)

Range: Off Georgia coast (48 to 77 m)

This blind, pigmentless species is the type and only species in the genus. It has biting mouth parts.

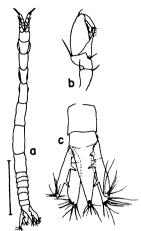


Figure 123. a. Horolcanthura irpex (female). b. Peraeopod I. c. Detail posterior part.

7b. Exopods of uropods with no spines on inner margins ... Hyssura

GENUS HYSSURA

The species of this genus are blind with very pointed, indurate telsons. No dorsolateral keels or dorsal pits are present. The maxillipedal palp has three articles, and pleopod 1 is never operculate.

Fig. 124a. Hyssura producta (Norman and Stebbing) (6.5 mm)

Range: North Atlantic-56° N X 37° W (2642 m)

This species has the posterior border of the telson much rounder in cross section than that of *H. profunda*.

Fig. 124b. Hyssura profunda (Barnard) (10 mm)

Range: North Atlantic (No specific place or depth given)

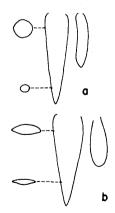


Figure 124. a. Hyssura producta, telson and endopod. b. H. profunda, telson and endopod.

8a. Mouth parts modified for piercing and sucking; maxillipedal palp with one or two articles; dorsal pits never present9

8b. Biting mouth parts; maxillipedal palp of three articles; dorsal pits absent or present11

9a. Maxillipedal palp of one article; no statocysts present; eyes present Paranthura

GENUS PARANTHURA

The genus contains a heterogeneous assemblage of species that are best separated here on the basis of shape of the telson.

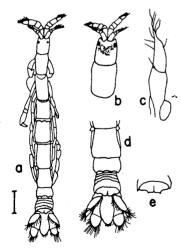


Figure 125. a. Paranthura elegana. b. Detail anterior part. c. Maxilliped. d. Detail posterior part. e. Pleonal segment 6.

Fig. 125. Paranthura elegans Menzies v (8.6 mm)

Range: Mid-California to northern Baja California (18 to 55 m)

The telson is (lanceolate) in this species.

Also in Gulf of Colifornia

Paranthura longitelson Wägele, 1983 Sea of Cortez (midriff area) Fig. 126. Paranthura infundibulata (Richardson) (7 mm)

Range: Bermuda and West Indies (Shallow water)

As the illustrations show, the male has a concave posterior border on the telson which has subparallel borders; the female has a straight posterior border on the telson. The male and female were originally described as separate species. A third species *P. antillensis* Barnard (5.5 mm long, from St. Johns and St. James Islands in the West Indies at 29 m) has been included in the genus. It has never been adequately illustrated, but is said to differ from the above species in that the telson is ovate (Fig. 127c).

This is a senior syonyon of <u>Colorithum tenuis</u> by Schultz (1977), but not according to Kensley & Poore (see letter from Kensley 7/20/72).

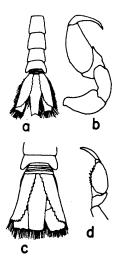


Figure 126. a. Paranth u ra infundibulata (male). b. Peraeopod I, male. c. Posterior part female. d. Peraeopod II, male.

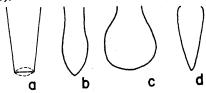


Figure 127. Telson types. a. With subparallel margins, posterior border (convex or concave). b. Lancelolate. c. Ovate. d. Pointed.

9b. Maxillipedal palp of two articles; single or fused statocysts present; eyes absent or present _______10

10a. Eyes usually present; peraeonal segment VII shorter than broad; pleon short but with distinct segments Accalathura

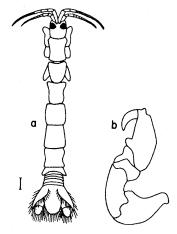


Figure 128. Accalathura crenulata (male). b. Peraeopod I, male.

Fig. 128. Accalathura crenulata (Richardson) (18 mm)

Range: Bahamas, West Indies and Yucatan (38 m)

This species has been found in relatively shallow water and also in gulfweed—sargassum.

Accalathura crassa (Barnard) (8 mm) from St. Johns, West Indies, has also been described, but never well illustrated. It differs from other species in the genus in being blind and lacking a produced posterolateral corner on peraeonal segment VII. It needs to be redescribed to be properly placed in the anthurids.

GENUS LEPTANTHURA

The species of the genus have a single fused statocyst (Fig. 120b). About 10 species are included in the genus, but only two are from North American waters.

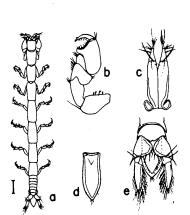


Figure 129. a. Leptanthura tenuis. b. Peraeopod I. c. Maxillipeds, d. Telson. e. Detail posterior part.

Fig. 129. Leptanthura tenuis (Sars) (14 mm)

Range: North Atlantic (near Portugal) (274 to 1313 m)

The specimen was taken just outside of the range of most species included in this book, but it is illustrated because L. thori Barnard (7 mm) from the North Atlantic (957 m) found within the North American waters has never been illustrated. L. thori resembles L. tenuis, but the telson is widest at the base narrowing very slightly to a rounded apex which bears a tuft of setae not set into a notch. The endopods of the uropods are also more pointed in L. thori.

- 11a. Eyes present or absent; operculate pleopods 1 (Fig. 121a, c) never fused 12
- 11b. Eyes present; operculate pleopods 1 fused Eisothistos

Fig. 130. Eisothistos atlanticus (Vanhoeffen) (5 mm)

Range: Cape Verde Islands and St. Thomas, West Indies (9 m)

Since the specimens that represent the species are from two widespread localities, they could represent two species. They must be examined more closely.

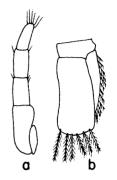


Figure 130. a. Eisoth is tos atlanticus, maxilliped. b. Pleopod 2, female.

12a. Eyes present; maxillipedal palp of four or five articles; dorsal pits absent, but dorsolateral keels presentPananthura

Fig. 131. Pananthura formosa (Menzies and Frankenberg) (4.5 mm)

Range: Off Georgia coast (77 to 90 m)

This eyed, small species has fairly long antennae and is the first species of the genus to be recorded from North American waters.

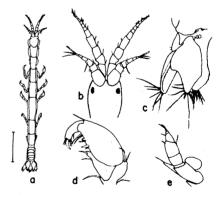
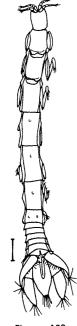


Figure 131. a. *Pananthura formosa*, b. Detail, cephalon. c. Detail, pleon. d. Peraeopod I, female. e. Maxilliped.

- 13a. Blind; dorsal pits and dorsolateral keels present and conspicuous; telson slightly longer than pleon with paired statocysts, dorsomedial groove and truncate posterior margin fringed with setae

 Bathura



Cortezura penasco ensis Schultz 1977

Fig. 132. Bathura luna Schultz (21 mm)

Range: Tanner Canyon on the shelf off southern California (783 to 812 m)

The species was taken from the green mud on the bottom of the canyon. The species is the type and only one in the genus. AHF has only the Hobtypes, No subsequent records are known, not even from the BLM study. Kensley (18) remained this species to Anthelura Norman & Stellings thus leaved the genus Parish a housen

Figure 132. Bathura luna.

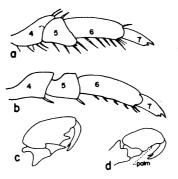


Figure 133. Peraeopod VI and palm types. a. Carpus underriding propodus. b. Carpus not underriding propodus. (4-merus; 5-carpus; 6-propodus; 7-dactylus) c. Palm notched. d. Palm not notched.

GENUS APANTHURA

Only two species of this very large genus are recorded from North American waters.

For a key to the world species see Krucznski & Hyers (1976). For a generic description see Miller & Musics (1952).

Fig. 134. Apanthura magnifica Menzies and Frankenberg /%6 (8.5 mm)

Range: Off Georgia coast (17 to 137 m). Also Florida.

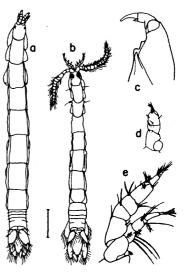
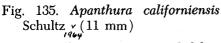


Figure 134. a. Apanthura magnifica, (female). b. Male c. Peraespod I, female. d. Maxilliped. e. Antennae two and one.



Range: Off Santa Monica, California (80 m)

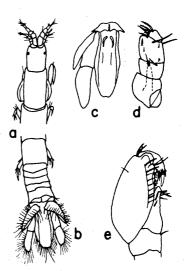


Figure 135. a. Apanthura californiensis anterior part. b. Posterior part. c. Telson and uropod. e. Peraeopod I, female. d. Map.

14b. Usually blind; dorsal pits absent or present; telson various; carpus of peraeopod VI not underriding propodus (Fig. 133b)

GENUS ANANTHURA

This genus contains a heterogeneous group of species which are not well enough described to be placed in any other genus with confidence. The palp of the maxilliped has three articles, and pleopod 1 is operculate.

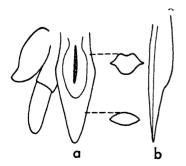


Figure 136. a. Ananthura sulcaticauda, telson and uropod. b. Lateral view of indurate telson.

Fig. 136. Ananthura sulcaticauda (Barnard) (6.5 mm)

Range: Davis Strait

This is the type-species of the genus.

Fig. 137. Ananthura affinis (Richardson)

Range: Bermuda (Shallow water)

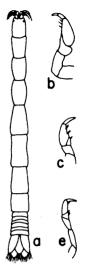


Figure 137. a. Ananthura affinis. b. Peraeopod I. c. Peraeopod II. d. Peraeopod VI.

Fig. 138. Ananthura abyssorum (Norman and Stebbing) (9 mm)

Range: Davis Strait (3200 m)

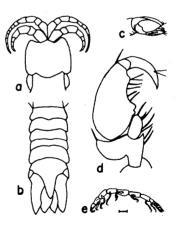


Figure 138. a. Ananthura abyssorum anterior part. b. Posterior part. c. Laterial view, posterior part. d. Peraeopod I. e. Lateral view, whole animal.

15b. Blind; telson broadly ovate widest at basal third; dorsal pits absent

Anthelura

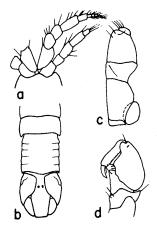


Figure 139. a. Anthelura truncata anterior part. b. Posterior part. c. Maxillipeds. d. Peraeopod I.

Fig. 139. Anthelura truncata (Hansen) (10 mm)

Range: Davis Strait (2193 to 2624 m)

There is still question as to the correct genus for this species—is it an *Anthelura* or a *Cyathura*?

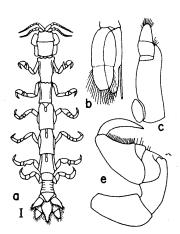


Figure 140. a. Calathura branchiata. b. Pleopod 2, male. c. Maxilliped. d. Peraeopod 1, male.

Fig. 140. Calathura branchiata (Stimpson) (26 mm)

Range: North Atlantic (37 to 831 m)

This type and only species is common being found burrowed in mud, clay and in rocky sandy bottoms from the Gulf of Maine northward. The species is the largest recorded anthurid from North American waters, but the length is approached by species of *Cyathura*.

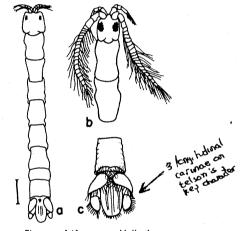
Silophasma

Fig. 141. Haliophasma geminata (Menzies and Barnard 1959)

Range: Pt. Conception to San Quintin Bay, Baja California (Shallow water to 16 m)

The species is very abundant on the continental shelf off southern California and northern Mexico. It is apparently associated with Gnathia crenulatifrons, another abundant benthic isopod (p. 224). The species has chewing mouth parts.

- 9.2 to 5/2 m, on shelf & slope.
- see schultz (1977) for generic eyonymy.



Supressed!

Figure 141. a. Haliophasma geminata (female). b. Anterior part, male. c. Posterior part, male.

- Konsley & Poore claim H. geminate fits Poore's
generic description perfectly and should remain in Haliophasma. (see lefter) Sills
106 Control of the second should remain in Haliophasma.

GENUS CYATHURA

This genus contains the largest number of species of any anthurid genus from North American waters. The species *C. polita* has been studied more than any other anthurid, and probably as much is known of its ecology as of any other isopod species. The species listed elsewhere in this book (p. 102) called *Anthelura truncata* perhaps is more properly placed in this genus. It needs to be examined more closely.

KEY TO SPECIES OF CYATHURA

19a.	Eyes present	.21
19b.	Blind	.20
	Male pleopod 2 with stylet longer than exopod	
	C. curassas	ica

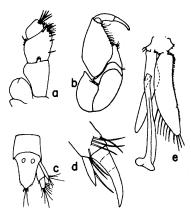


Fig. 142. Cyathura curassavica (Stock) (7 mm)

Range: Freshwater caves on Curação, Dutch West Indies

Figure 142. a. Cyathura curassavica, maxilliped. b. Peraeopod I, male. c. Detail, pleon and uropod. d. Detail, dactylus peraeopod V. e. Pleopod 2, male.

20b. Male pleopod 2 with stylet shorter than exopodC. specus

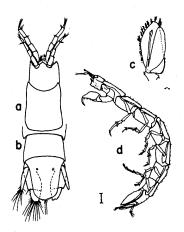
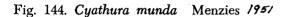


Figure 143. a. Cyathura specus anterior part, female. b. Posterior part. c. Pleopod 2, male. d. Lateral view, female.

Fig. 143. Cyathura specus (Bowman) (20 mm)

Range: Freshwater caves of northern Cuba



Range: Mid-California to Mexican border (18 to 55 m) & in Gulf of California.

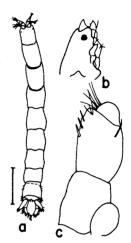


Figure 144. a. Cyathura munda (female). b. Lateral view, cephalon. c. Maxilliped.



Fig. 145. Cyathura crucis (Barnard) (7 mm)

Range: St. Croix, West Indies (7 m)



Figure 145. a. Cyathura cruis, telson a n d uropod, male. b. Telson and uropod, female.

23a. Sixth p	leonal segn	nent fused to telson
& A .	, market	Fig. 146. Cyathura polita (Stimpson) (25 mm)
		Range: Maine to Louisiana (Tidal marshes and shallow water)
b		The species lives in mud and muddy sand in unlined burrows of its own con- struction, usually in places where there is a continual movement of water and an inflow of freshwater from a stream or spring. The isopods have a high tolerance
C/	77	for the changes in the physical factors and salinity changes that take place in their shallow water habitats. At times
		they are found in enormous numbers. The species form a very important part of the ecosystem of the bays and marshes that border the sea where they eat and

Figure 146. a. Cyathura polita (male). b. Maxilliped. c. Detail, posterior part of pleon, e. Peraeopod I, male,

22b. Dorsolateral keels feebly or strongly indicated 23

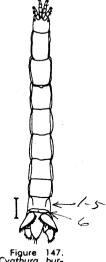
The species form a very important part of the ecosystem of the bays and marshes that border the sea where they eat and are eaten by other animals in the ecological food web. They thus have an important role to play in the flow of energy through the near shore ecosystem. Their role on the bottom of the shallow water is somewhat like that of earthworms on

land. The composition of the mud in which they live is constantly changed through the chemical and physical actions of their living processes. A detailed study of the life processes of the isopod is rewarding because the environment in which they live is one of the richest in terms of biological productivity of any environment on land or in the deep sea.

- 23b. Sixth pleonal segment separated from telson 24
- 24a. Medial cephalic projection rounded or obtusely pointed, as long as or slightly shorter than anterolateral cephalic pro-

Fig. 147. Cuathura burbanki Frankenberg (12 mm)

Range: Coast of Georgia (16 to 77 m)



24b. Medial cephalic projection pointed and as long as or slightly

Fig. 148. Cyathura carinata (Kroyer) (15 mm)

Range: Northwestern North America and Greenland (Surface to 19.5 m)

It is possible that this species is really only found in Europe, and its inclusion in North American records is in error. If it is present, it is found only in northern waters because the more southern locations where it has been reported to occur are inhabited only by C. polita. The species is the type-species of the genus. All animals of the species begin life as females, become sexually mature, and then some change to become males; i. e., they are protogynous.

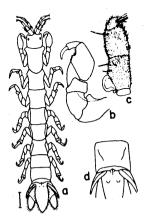


Figure 148. a. Cyathura carinata. b. Peraeopod. c. Maxilliped. d. Detail, posterior part of pleon.

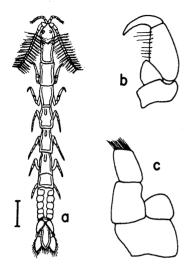


Figure 149. a. Ptilanthura tenuis (male). b. Peraeopod I, male. c. Maxilliped.

Fig. 149. Ptilanthura tenuis (Harger) (8.5 mm)

Range: Bay of Fundy to Long Island Sound (Surface to 35 m)

This species is the type-species of the genus. It was taken on muddy bottoms in deep water and on stony bottoms in shallow water.

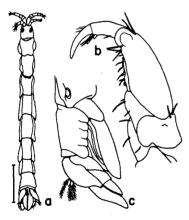


Figure 150. a. Ptilanthura tricarina (female). b. Peraeopod I, female. c. Lateral view, pleon.

Fig. 150. Ptilanthura tricarina (Menzies and Frankenberg) (5.8 mm)

Range: Off Georgia (25 to 141 m)

There is no dorsal indication of segmentation in the pleon of the species as there is in *P. tenuis*.

25b. Maxillipedal palp of three articles; mandibular palp of three articles

Mesanthura

Fig. 151. Mesanthura pulchra (Barnard) (7 mm)

Range: St. Thomas and St. Johns, West Indies (18 to 40 m)

Only a single crude figure of this species is available, but the dorsal pigment pattern is characteristic of the species.



Figure 1. Mesanthura pulchra.

Mesanthura nubitera wägele, 1983
Sea of Cortezintertidal
(midriff region)

Fig. 152. Mesanthura occidentalis Menzies and Barnard, 1959

Range: Pt. Conception to San Quintin Bay, Baja California (Shallow water to 18 m)

This species has a characteristic body pigmentation. It is found in live and dead algae on the bottom in shallow water. And in Gulf of California

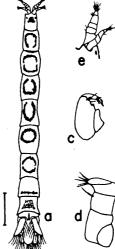


Figure 152. a. Mesanthura occidentalis (female). b. Antennae one and two. c. Peraeopod I, female. d. Maxilliped.

SUBORDER FLABELLIFERA

The suborder Flabellifera contains the largest number of species that any amature collector will probably encounter of any isopod

suborder. It includes most of the common near shore species and also many economically important species. Generally the species are medium to large in size, but the largest as well as some of the smallest isopods are included within the suborder. The species are abundant in most near-shore habitats and are also common as fish parasites. They are rarely encountered in the deep sea. For most species the mouth parts are biting and strongly developed, but the parasitic species have mouth parts that are modified for their peculiar parasitic type of feeding. Most species have eyes although cave forms have lost their eyes, and certain of the more parasitic species have rudimentary eyes or no eyes at all. The antennae are generally well developed, but parasitic forms for the most part have small antennae.

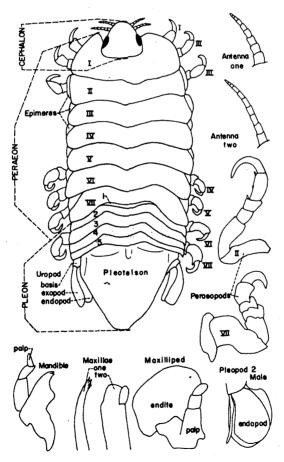


Figure 153. Flabellifera (General nomenclature).

The peraeopods are ambulatory and prehensile and frequently a combination of the two types are found on one animal. The pleonal segments are generally well defined although they are sometimes not all distinct in the dorsal view. Lateral indications are generally present when pleonal segments are fused. Pleopods are always present, and they are never found within a branchial chamber. In some species they are well developed and are said to be natatory. The uropods generally have a flattened exopod and endopod and together with the pleotelson serve as part of the caudal fan which is used in swimming. Since there are many well defined families in the suborder, further discussion of the flabelliferan species will be found there. There are many subtile differences between groups and many exceptions to general rules of structure among the species. Species of ten families are known from North American waters.

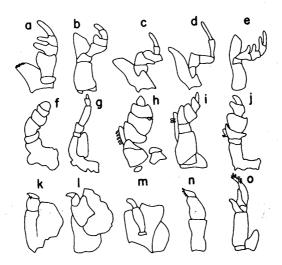


Figure 154. Maxillipeds of Flabellifera (for Cirolanides see Fig. 274). a. Tecticeps. b. Cassidinidea. c. Sphaeroma quadridentatum. d. Sphaeroma destructor. e. Exosphaeroma (E. rhomburum). f. Alcirona. g. Excorallana. h. Bathynomus. i. Excirolana. j. Cirolanides. k. Anilocra. 1. Meinertia. m. Serolis. n. Nerocila. o. Aega.

KEY TO FAMILIES OF FLABELLIFERA

1a. Pleon with at least free pleonal segment 2

1b. Pleon with only one pleonal segment indicated mediodorsally Cirolanidae (genus Colopisthus only-p. 168)

2a. Pleon composed of four or five free segments plus pleotelson (a pleonal segment sometimes is hidden under peraeonal segment VII)

2b.	Pleon composed of three or less visible free segments plus pleotelson
3a.	Pleon composed of only one free segments plus pleotelson; uropod with one ramus fused to basis; generally able to roll into ball
3b.	Pleon of three free segments plus pleotelson; uropods with both rami movable; body greatly compressed dorsoventrally, never able to roll into ball
4 a.	Uropods present5
4b.	Uropods absent
5a.	Uropods with both exopod and endopod flattened6
5b.	Uropods with one or both with hook-like ramiLimnoridae (p. 138)
6a.	All peraeopods prehensile (modified for clinging) or all peraeopods ambulatory, never mixed; with when the location we have the company of th
6b.	Peraeopods both prehensile and ambulatory (or modified natatory—Fig. 31b) on same individual 5 Mdb. w/a lacinia mebils
7a.	All peraeopods prehensile; maxillipedal palp of two articles, distal article with hook-like setae
7b.	All peraeopods ambulatory; mandibles with lacinia mobilis and well developed peculiar tooth-bearing molar process (Fig. 37f)
8a.	Maxillipedal palp of two or five articles, distal article with stout hook-like setae
8b.	Maxillipedal palp usually of five articles, distal article with no hooks only fringing setae
9a.	Antepenultimate article (palp article three) of maxillipedal palp longer than broad (Fig. 155a) Excorallanidae (p. 205)

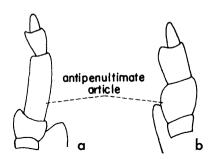


Figure 155. Antipenultimate maxillipedal palp articles compared. a. Excorallana, b. Corallana.

SPHAEROMATIDAE

The species in the family sphaeromatid, are not only among the most common groups of isopods, but they are in one of the most taxonomically confused groups as well. Species of many genera are found in the near shore environments. Some species are sexually dimorphic; others are not. They are generally small to medium in size and are very good swimmers, but there are no special flattened natatory peraeopods on the specimens. Instead, peraeopods with very long natatory setae are found (Fig. 31b). The species are frequently taken from under rocks and debris found on the beaches of the world. They are abundant in tidal pools and are frequently taken in and among encrusting animals and plants on wharves, piles and rocks. Many times they are found in burrows of other animals including the boring species of Limnoria. There are some species that will bite if they are caught in the bare hand. Their bites are harmless, however. One species of the family is known to destroy wooden pilings by boring into them just as the species of Limnoria do. They then are of considerable economic importance to man.

There are several different ways that the sphaeromatid are classified. One of them, used mainly for the European species, is based on the structure of the pleopods. Some pleopods, usually the last two or three, are "fleshy" or membraneous and others are more or less translucent. Sometimes all pleopods are translucent. There are either one or two segments in the rami of the pleopods (Fig. 180), and fringing setae (usually plumose setae) are absent or present. Since the particular pattern of the characters of the pleopods are

not known for all North American genera, it will be used here only in part.

Some member of the sphaeromatids are sexually dimorphic, the male being much different than the female. The female is usually "plain" or generalized and does not differ much from other females either of the same or of different genera. The species generally can only be firmly defined and differentiated on the differences found in the males. Species of Paracerceis are good examples of sexually dimorphic species (p. 120).

Many sphaeromatid species are able to roll into a tight ball like their terrestrial cousins the pill bugs (Oniscoidea). Indeed, that is where the name sphaeromatid comes from since the isopod roll into spheres. Sometimes there are dorsal projections or the uropods are modified so that they form very spiny balls when rolled up. The five pleonal segments are fused so that there are only lateral indications of the segments. The pleotelson forms a caudal fan with the uropods and it serves as a swimming structure. The basis of the uropod and the endopod are fused and the exopod is freely articulate and lateral to the fused basis and endopod. The epimeral plates are fused to the lateral margins of the peraeonal segments in such a manner as to appear continuous with them, except of course those of peraeonal segment I.

KEY TO GENERA AND SPECIES OF SPHAEROMATIDAE

1a. Uropods uniramus, only long endopod presentAncinus

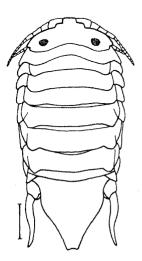


Figure 156. Ancinus depressus.

Fig. 156. Ancinus depressus (Say) (6.3 mm)

Range: Massachusetts to Texas (Surface to 3 m)

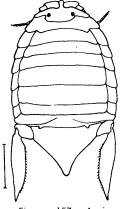
The species is rarely found in water over 3 meters deep. Occasionally in North Carolina the species is taken in wave washed beach sand. The species of the genus are common, but rarely are found in large numbers.

Fig. 157. Ancinus granulosus (Holmes and Gay) (8 mm)

Range: Southern California (6 to 10 m)

The species was found in two places in southern California, At Pt. Conception it was found in medium-coarse, gray sand. It differs from the east coast species of the genus in that the pleotelson is more pointed and other characters. (see Schultz, 1973)

Ancinus seticomvus Trask is a jr. synonym of A. granulatus



157. Ancinus dellanae

- 1b. Uropods biramus (remember the endopod and basis are fused)
- 2a. Uropod with exopod very short and set within margin of long endopod

Fig. 158. Cassidinidea lunifrons (Richardson) (4 mm) Charleston (Stono River)

Range: Great Egg Harbor, New Jersey to Georgia (Surface to 1 m)

The species is found in the mud and dead leaves in the shallow water of estuaries and inland embayments and brackish marshes. The coxal extensions are indicated by a groove; they are not actually separated from the peraeonal segments as in species of Ancinus.

Also: Cassidinidea Euberculata Rich 1912 from Madre, Mexico. (pleotelson roundeds pleon tuberculate).

Fig. 159. Cassidinidea ovalis (Say) (3.8 mm) Range: South Carolina to Florida (Verv shallow water)

It is still an open question if this species is the same as C. lunifrons. The two species overlap in range, but there still are differences in the uropodal structure so that they

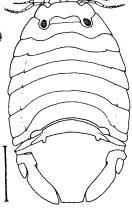


Figure 158. Cassidinidea lunifrons.

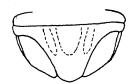


Figure 159. Cassidinidea ovalis, posterior part.

note: H.E. Hendrickx has sent me specimens of a Cassidinea from Mozatlan!

must be considered separate species until more specimens from southern locations are obtained.

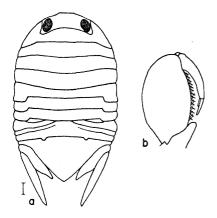


Figure 160, a. Tecticeps alascensis, b. Peraeopod I.

Fig. 160. Tecticeps alascensis (Richardson) (16 mm)

Range: Bering Sea and Aleutian Islands (16 to 194 m)

The posterior margin of the pleotelson is pointed in this species.

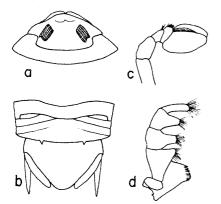


Figure 161. a. *Tecticeps convexus*, anterior part. b. Posterior part. c. Peraeopod I. d. Maxilliped.

Fig. 161. Tecticeps convexus (Richardson) (11.5 mm)

Range: Monterey Bay, Califor nia (9 m) Ph Conception to Crewent City.

This species is known to roll into a ball and the exopod of the uropod forms a protective spine. There is a rounded posterior margin on the pleotelson in the species.

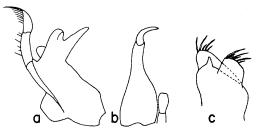


Figure 162. Mouth parts of *Tecticeps convexus*, a. Mandible, b. Maxilla one, c. Maxilla two.

Figs. 162 and 163. Tecticeps pugettensis (Hatch) (10.1 mm)

Range: Whidbey Island, Washington (13 m)

The configuration of the posterior margin of the pleotelson and the position of the eyes distinguish this species from other North American members of the genus.

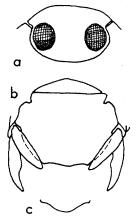


Figure 163. a. Tecticeps pugettensis, cephalon. b. Pleotelson. c. Posterior marain of pleotelson.

- 3b. All peraeopods ambulatory; maxillipedal palp structure vari-
- 4b. Posterior medial border of pleotelson not deeply cleft or with moderately deep medial groove (deep medialateral clefts can be present causing the medial portion of the posterior border of the pleotelson to be produced into a point-Fig. 309)
- 5a. Exopod and endopod about same length with exopod capable of folding under endopod; medial cleft in posterior border of pleotelson flanked by two produced sections of posterior border which can be closely adherent at tips causing triangulate hollow in pleotelson (Fig. 173)

- 6a. Cephalon produced into three anterior lobes; uropods with tips of exopods turned outward (in males); no teeth within shallow notch on posterior border of pleotelson *Dynameniscus*

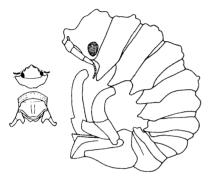


Figure 164. a. Dynameniscus carinatus, dorsal view. b. Lateral view.

Fig. 164. Dynameniscus carinatus (Richardson)

Range: Coast of Georgia (805 m)

One small male specimen is all that represents the typespecies of the genus.

6b. Cephalon not produced into three lobes; uropods with tips of exopods turned inward (in males); teeth within notch on posterior border of pleotelson

Paracerceis

GENUS PARACERCEIS

This genus with a complicated taxonomic history is further made more difficult to study because the males are quite different from the females. Males are usually greatly ornamented with long caudal process extending from the pleotelson or it might be said a deep indentation usually with internal teeth in the medial border of the pleotelson. There are five species in the genus in North American waters, but it is quite possible that some of them are the same species or synonymous. It is also just as possible that there have been several species described under one species name.

Since the male and females are so different some have been described as being in different genera. More attention must be paid to the variation in the females when collected with known males. Females are not well described and are frequently confused, not

only with females of the same genus, but with females of other genera as well. They are probably subject to some individual and regional variation, but with more complete and accurate descriptions of their morphology their true identity can be made known. Males are also subjected to age and regional variations which have never been recorded. The species of the genus are frequently found on sandy bottoms and are also taken in corals and sponges. No key is included here since males and females are not known in all species.

Fig. 165. Paracerceis caudata (Say) (9 mm)

Range: New Jersey, Bermuda and West Indies (Surface to 46 m)

The species is found among algae and grass and in coral reefs. It is probably composed of a complex of species and subspecies and deserves more study.

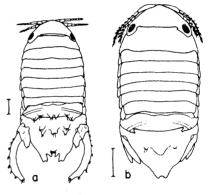


Figure 165. a. Paracerceis caudato (male). b. Female.

Fig. 166. Paracerceis tomentosa (Schultz and McCloskey) (6.2 mm)

Range: Cape Lookout, North Carolina (5 m)

The species was found in a coral head on Cape Lookout jetty. The male is unknown.

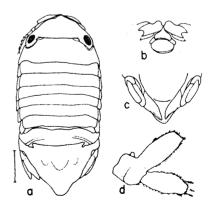


Figure 166. a. Paracerceis tomentosa (female). b. Frontal lamina. c. Pleon, ventral aspect. d. Uropods.

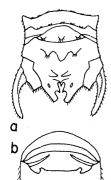


Figure 167. a. Paracerceis sculpta (male). b. Female.

Fig. 167. Paracerceis sculpta (Holmes)

Range: San Clemente Island and San Diego, California (Shallow water)

The species was found in pieces of sponges.

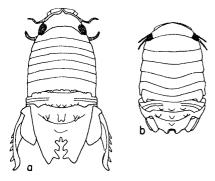


Figure 168. a. Paracerceis cordata (male). b. Female.

Fig. 168. Paracerceis cordata (Richardson) (7.1 mm)

Range: Aleutian Islands to southern California (37 to 55 m)

The species is found in corals as well as on sandy bottoms.

ie has descriptions of figs. of two new species of Paracerceis.

Fig. 169. Paracerceis gilliana (Richardson) (4.8 mm)

Range: Southern California (55 to 73 m)

This species differs from *P. cordata* only in minor features.

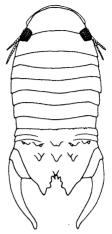


Figure 169. Paracerceis gilliana (male).

Fig. 170. Paradynamene benjamensis (Richardson) (10 mm)

Range: Atlantic coast and in Gulf Stream

The species was found in floating algae in the Gulf Stream.

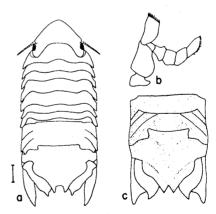


Figure 170. a. Paradynamene benjamensis (female). Maxilliped. c. Male.

8a. Antenna one with two basal segments of peduncle flattened and enlarged, generally visible in dorsal view in front of cephalon

Dynamene dilatata

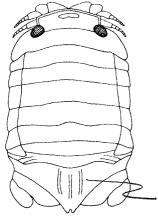


Fig. 171. Dynamene dilatata (Richardson) (3 mm)

Range: Monterey Bay, California

The true generic status of this species is still undetermined. Only one specimen has ever been collected.

flattened, w. shorel-like epistone

Figure 171. Dynamene dilatata.

one?

8b. Antenna two without expanded peduncular segments

Dynamenella and Dynamenopsis

GENERA DYNAMENELLA AND DYNAMENOPSIS

Reasonable morphological criteria for distinguishing species of the two genera are not found among the North American members. Also immature males look like adult females and can only be distinguished by the differences found in pleopods 2. The two genera are defined on the character of the pleopods and it is not known what those structures are like in North American species.

- 9a. Tubercles on dorsum of pleotelson in trapezoidal or square pattern ______10

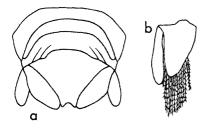


Fig. 172. Dynamenella glabra (Richardson) (3 mm)

Range: Monterey Bay, California (Surface)

Figure 172. a. Dyna menella glabra, posterior part. b. Pleopod 2, male.

Fig. 173. Dynamenopsis dianae
(Menzies) (4 mm)

Range: Northern Baja California
(Shallow water)

This species closely resembles
Dynamenella perforata from Puerto
Rico. It is found in large numbers
in San Quintin Bay, Baja California.

Also reported from Puerto Kico 4

Eniwetok Holl. 4 No Carolina

Figure 173. Dynamenopsis dianae.

Fig. 174. Dynamenella sheareri
(Hatch) (3.3 m) (formerly Typhamene)
Range: Coos Bay, Oregon (Shallow water)

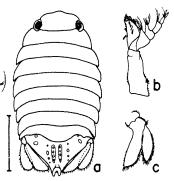


Figure 174. a. Dynamenella sheareri. b. Maxilliped. c. Uropod.

11b. Posterolateral margins of pleotelson convex along entire margin 12

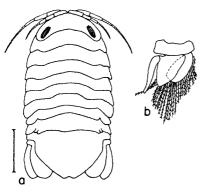


Figure 175. a. Dynamenella perforata, immature male. b. Pleopod 2, male.

Fig. 175. Dynamenella perforata (immature male)

Range: Bermuda to Puerto Rico (Intertidal)

The species has been recorded to be present clinging to the bottoms of chitons. Chitons are common mollusks of the tropical intertidal zone (see Fig. 176).

13a. Deep cleft of pleotelson opening into large cavity

D. perforata

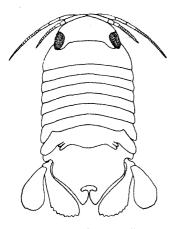


Figure 176. Dynamenella perforata.

Fig. 176. Dynamenella perforata (Moore) (4 mm)

Range: Puerto Rico, Culebra Island and Bermuda (Intertidal)

The species is found on the roots of mangroves and is the same species as D. moorei which was the name given to immature males and females of the species (see also Fig. 175).

 Fig. 177. Dynamenella angulata (Richardson)

Range: No Name Key, Florida

No males were ever described and the species might turn out to be the female of another related species.





Figure 177. a. Dynamenella angulata, posterior part. b. Maxilliped.

14b. Posterior margin of uropodal rami roundedD. benedict

Fig. 178. Dynamenella benedicti (Richardson) (3 mm)

Range: Monterey Bay, California (Shallow water)

Both males and females are known for this species.

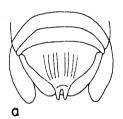




Figure 178. a. Dynamenella benedicti, posterior part. b. Pleopod 2, male.

15b.	Surface of body not densely granulated	17
16a.	Male with deep mediolateral clefts and endopod mu than exopod; pleotelson separated from pleonal (which are feebly indicated) by straight line	segments

15a. Surface of body densely granulated 16

GENUS DISCERCEIS

Only two species are now contained in this formerly large genus. Other species have been removed and put into separate genera all of which are included here.

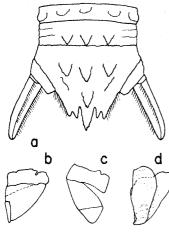


Figure 179. a. Discerceis grandulosa, posterior part. b. Pleopod 2, male. b. Pleopod 3. d. Pleopod 4.

Fig. 179. Discerceis granulosa (Richardson) (9 mm)

Range: Cerros Island, Baja California (37 m)

This species is the type-species of the genus. It is not well described and *D. linguicauda* is also not well described.

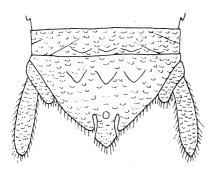


Figure 180. a. Discerceis linguicauda.

Fig. 180. Discerceis linguicauda (Richardson)

Range: Cape Catoche, Yucatan, Mexico (44 to 46 m)

16b. Male and female with exopod and endopod well developed; pleotelson separated from remaining part of pleon, not by straight line (sutures of pleonal segments strongly indicated at least laterally)

Cymodoce

Fig. 181. Cymodoce japonica (Richardson) (17.5 mm)

Range: Japan, but introduced to west coast (Surface to 1547 m)

The species was introduced to North America most probably on oysters. It thus has economic importance.

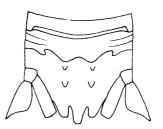


Figure 181. Cymodoce japonica.

Fig. 182. Cymodoce faxoni (Richardson (5.5 mm)

Range: Florida to Texas

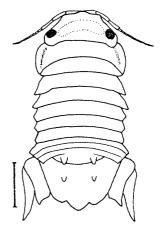


Figure 182. Cymodoce faxoni.

GENUS SPHAEROMA

Four species of this genus are common and widespread on the east coast of North America, but there is only one species on the west coast. The species are remarkably well pigmented, and there is much variation of color in the different populations and different regions. The species are frequently taken from the burrows of Limnoria species, the wood-boring isopod, and they apparently form an association with them. One species, S. destructor, has been

recorded to be itself a borer of wooden pilings in southern and southeastern United States.

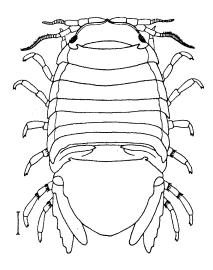


Figure 183. Sphaeroma quadridentatum.

Fig. 183. Sphaeroma quadridentatum (Say) (10 mm)

Range: Massachusetts to Key West, Florida (Surface to 1 m)

This species is very common on the east coast of North America where it is found on algae, marine pilings and occasionally on rocky, shelly bottoms. The dorsum of the pleotelson of this species is relatively smooth when compared to that of S. destructor.

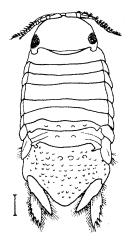


Figure 184. Sphaeroma destructor (for mouth parts see Fig. 38).

This sp. was made a terebrands

This sp. was of S. terebrands

This sp. was of S. terebrands

This sp. was made (acon) Pillary

Rate Lebbing (acon) Pillary

(acon), exc.

Fig. 184. Sphaeroma destructor (Richardson) (10 mm)

Range: Georgia and Florida to Louisiana (Intertidal and shallow water) Circumtropical!

This species destroys wooden marine structures and is of considerable economic importance. It has tubercles over its entire dorsum, but they are especially noticeable on the pleotelson. Bores into preprosts of B. margle (see simberleff, et al., 1978).

Fig. 185. Sphaeroma pentodon (Richardson), 1904
(8 mm) = 5. quoyana

Range: Sausalito, San Francisco Bay, California and southward (Mud flats and in soft rock)

This species has large teeth on the outer margin of the exopod of the uropod and two rows of tubercles medially placed on the dorsum of the pleotelson. The species is unique among North American isopods in that it burrows into soft rock just as S. destructor burrows into wood. The pleopods of the species serve as the home of the species Iais californica (p. 269), a small asellote. Also reported from Australia

New Zealand & Tasmania. With long schae on PI-7.



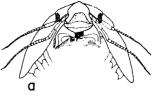


Figure 185. a. Sphaeroma pentadon, maxilliped. b. Posterior part.

Fig. 186. Sphaeroma barrerae (Boone) (13.5 mm)

Range: Cabanas, Cuba

This species has a "dome-like" pleotelson.



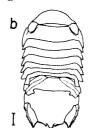


Figure 186. a. Sphaeroma barrerae, anterior part, ventral aspect. b. Dorsal view.

18a. Lateral edges of two or three pleonal segments extend to border of pleon; no "fleshy," folded pleopods

....Gnorimosphaeroma

Fig. 187(a). Gnorimosphaeroma oregonensis (Dana) (12 mm)

Range: Mid-California to Alaska (Intertidal to 22 m)

The species is common on the west coast in tidal pools and sometimes in brackish water. The edges of three pleonal segments reach the edge of the pleonal margin.



Figure 187(a). Gnorimosphaeroma oregoriensis, details of pleonal edges.

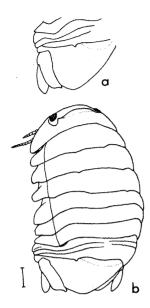


Figure 187(b). a. Gnorimosphaeroma lutea, details of pleonal edges. b. Dorsaloblique view.

Figs. 187(b). Gnorimosphaeroma lutea. Menzies 181(8.5 mm)

Range: Mid California to Alaska (Fresh and brackish water)

The species had been considered as a subspecies of *G. oregonensis*, until it was demonstrated that there were consistent physiological and ecological differences. The lateral edges of only two pleonal segments reach the edge of the margin of the pleon.

this is the ir syn of
G. insulare. see
Hoostloudt, 1977

For inform 6 nori mosphaeroma Dec.
popers by Hoestlandt, and also Menzies, 1954
1977 \$1969

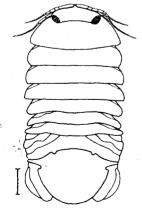


Figure 188. Gnorimosphaeroma insularis.

Fig. 188. Gnorimosphaeroma insular/s (Van Name, v (8 mm)

Range: San Nicolos Island off southern California (Freshwater)

The species differs from G. oreogonensis and G. lutea in angulation, length and relationship of the pleonal sutures.

freshwater - estuarial; Calif to Alaska.

MARINE ISOPOD CRUSTACEANS

noblei

Fig. 189. Gnorimosphaeroma nobeli-Menzies (2.9 mm)

Range: Tomales Bay, California (Intertidal) to Los Angeles.

This species can be told from other species in the genus by the configuration of the pleotelson and by the smooth dorsum. G. rayi Hoestlandt has also been described from Tomales Bay, but it only differs in color from other species in the genus. It may only be a subspecies of one of the species in the genus. Also reported from kurile Is., Sea of Othersk.

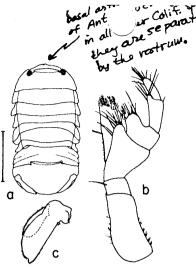


Figure 189. a. Gnorimosphaeroma nobeli, b. Maxilliped. c. Uropod.

18b. Lateral edges of only one pleonal segment extend to border of pleon; endopods of pleopods 4 and 5 "fleshy" and folded Exosphaeroma

GENUS EXOSPHAEROMA

This genus contains a large and heterogeneous assemblage of species. When they are more thoroughly examined they will undoubtedly be divided into several genera.

KEY TO SPECIES OF EXOSPHAEROMA

19a. Uropodal rami very large and broad; posterior margin of pleotelson pointed E. amplicauda

Fig. 190. Exosphaeroma amplicauda (Stimpson) (8 mm)

Range: Aleutian Islands, Alaska to 50. mid-California (Wet shore to low water)

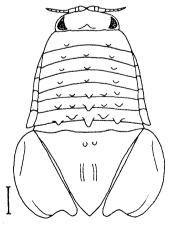
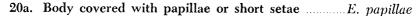


Figure 190. Exosphaeroma amplicauda.

					posterior		
telson t	runcate,	rounded	or	obtuse	ly pointed	d	20



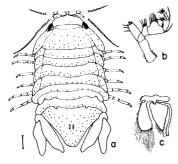


Figure 191. a. Exosphaeroma papillae. b. Maxilliped. c. Pleopod 2, male.

Fig. 191. Exosphaeroma papillae (Bayliff) (9 mm)

Range: North shore Long Island, New York (Brackish water)

20b.	Body without noticeable papillae or short setae
21a.	Cephalon not deeply or moderately set into peraeonal segment I, but extending acrossed entire width of body22
21b.	Cephalon deeply or moderately set into peraeonal segment I
22a.	Uropods set outside of general body margin
22b.	Uropods set within general body margin E. crenulatum

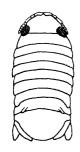


Figure 192. Exosphaeroma crenulatum.

Fig. 192. Exosphaeroma crenulatum (Richardson)

Range: Bermuda

This species according to the original author is morphologically placed between a *Sphaeroma* and an *Exosphaeroma* species.

Fig. 193. Exosphaeroma inornata
Dow /958

to Humboldt Bay,
Range: Palos Verdes A California
The isopod was found under the
holdfasts of kelp.

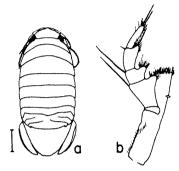


Figure 193. a. Exosphaeroma inornata. b. Maxilliped.

23b. Body margins only slightly convex; pleonal segment 3 completely separated from others E. media

Fig. 194. Exosphaeroma media George and Stromberg 1968

Range: Puget Sound, Washington (Intertidal)

The species is found in the sand in the upper part of the intertidal zone.

This will be reduced to a jr. synonym of E. inornata by & Iverson.

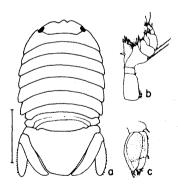


Figure 194. a. Exosphaeroma media. b. Maxilliped. c. Pleopod 4, male.

24a. Pleotelson longer than wide; uropodal rami elongate

E. dugesi

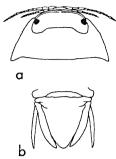


Figure 195. a. Exosphaeroma dugesi, anterior part. b. Posterior part.

Fig. 195. Exosphaeroma dugesi (Dollfus) (12 mm)

Range: Aquascalientes State, Mexico (Springs)

The species is found in regular freshwater springs and in hot springs abundant near Aquascalientes, Mexico. All peraeopods are ambulatory on the species.

- 25a. Eyes large; posterior border of pleotelson strongly truncate; two tubercles on dorsum of pleotelson E. antillense

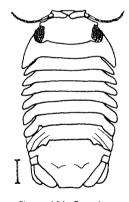


Figure 196. Exosphaeroma antillense.

Fig. 196. Exosphaeroma antillense (Richardson) (7.2 mm)

Range: Montego Bay, Jamaica (Shallow water)

25b.	Eyes of only moderate size; posterior border of pleotel	
	various; dorsum of pleotelson variously decorated or decorated	
26a.	Dorsum of pleotelson with ornamentation	27

27a. Posterolateral margin of pleotelson regularly convex E. octoncum

Fig. 197a. Exosphaeroma octoncum (Richardson)

Range: Monterey Bay, California to Tomales Head
(Marin Co.)



Fig. 197b. Exosphaeroma rhomburum (Richardson) (3.1 mm)

Range: Monterey Bay, California



Figure 197. a. Exosphaeroma octoncum. b. E. rhomburum.

28a. Cephalon deeply set into peraeonal segment I; posterior margin of peraeopodal segment I sinuate E. thermophilum

Fig. 198. Exosphaeroma thermophilum (Richardson) (4.6 mm)

Range: Near Socorro, New Mexico (Warm Springs)

Another species of the genus *E. dugesi* has also been found in warm springs. How the species reached the warm springs from the marine environment is a mystery.



rigure 198. Exosphaeroma ther-

28b. Cephalon only moderately set within peraeonal segment I; posterior margin of peraeonal segment I straight

E. diminutum

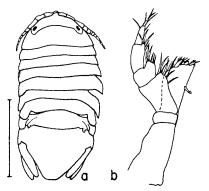


Figure 199. a. Exosphaeroma diminutum. b. Maxilliped.

Fig. 199. Exosphaeroma diminutum (Menzies and Frankenberg) (2.2 mm)

Range: North Carolina to Georgia (Intertidal and in beach sand)

SEROLIDAE

The family is composed of large dorsoventrally flattened animals that are widely distributed on the bottom of the deep sea. In the Antarctic region the species are especially abundant and are found in shallow water and tidal pools as well as in the deep sea. The specimens of Serolis species are large enough to appear in photographs taken by camera lowered to near the bottom in water 2 miles deep. The species apparently live partially immersed in the loose bottom mud and are most probably filter feeders. There are several genera in the family, but the nominal genus Serolis is well known and the only one that is represented in North American waters. Three species, two from the North Atlantic and one from the Pacific Ocean, have been recorded. All North American specimens are small when compared to those from more southern locations. Only six peraeonal segments are visible in the dorsal view in most species and the first peraeopod has a subchelate prehensile apex.

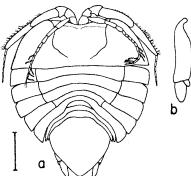


Figure 200. a. Serolis vemae, b. Uropod.

Fig. 200. Serolis vemae (Menzies) (4.3 mm)

Range: Between Martha's Vineyard and Bermuda (2682 to 5024 m)

This bottom dwelling species is blind. It was the first species of *Serolis* collected in the North Atlantic.

Fig. 201(a). Serolis mgrayi (Menzies) (4 mm)

Range: Off coast of Georgia (33 to 44 m)

This species has eyes and was taken from relatively shallow water.

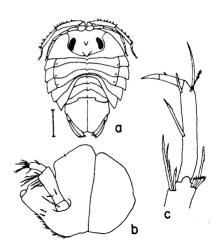


Figure 201(a). a. Serolis mgrayi, b. Maxilliped. c. Peraeopod VII.

Fig. 201(b). Serolis carinata (Lockington) (5 mm)

Range: Santa Monica to San Diego, California (13 to 55 m)

This species was taken in small numbers in relatively shallow water off the southern California coast. (See Hessier, 1972)

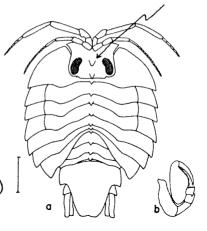


Figure 201(b). a. Serolis carinata. b. Peraeopod I.

Serdis tropica West central

America. Lacks cephalic spines. See Glynn (1976).

ANUROPIDAE

Only species of the genus *Bathynomus* from the deep sea floor are larger than isopods of the family Anuropidae. The name of the family implies that the animals have no uropods; however, uropods are present which are only superficially different from the five pleopods. Many species are found in the deep sea, but they are not bottom dwellers as are species of *Bathynomus*. They are rather bathypelagic; i. e., they are found swimming in the deep

water and apparently only rarely go to the bottom. Instead of having a caudal fan composed of broadened uropods and a flattened pleotelson, only the pleotelson which is extremely large and flattened as a swimming structure is present. In proportion to the size of the isopod, the first antennae are extremely large and the second antennae are very small. The isopods are frequently taken in deep plankton tows and are occasionally taken near the surface as well. Only three species are recorded for the family and one is present in North American waters off the coast of California.

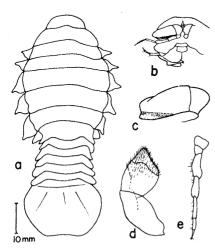


Figure 202. a. Anuropus bathypelagicus. b. Cephalon, ventral view. c. Antenna one. d. Maxilliped. e. Antenna two.

Fig. 202. Anuropus bathypelagicus (Menzies and Dow) (70 mm)

Range: Off central California (Surface to 2926 m)

The species is blind and is usually taken in very deep plankton tows, but occasionally is caught on or near the surface. At the depth where it is most frequently caught, it has no need of eyes.

LIMNORIDAE

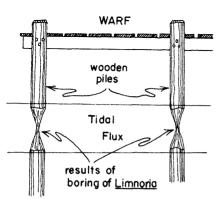


Figure 203. A wharf with a short useful life as the result of *Limnoria* borings.

Species of the family are wood and algae eaters. They are able to bore holes into the plant materials that are found naturally or placed by man in the sea and brackish water. Since much of the marine construction of man is supported on wooden pilings (Fig. 203) and many of his small boats are of wooden construction, a natural food source is supplied to the isopods and millions of dollars are spent to replace that which is eaten.

KEY TO GENERA AND SPECIES OF LIMNORIDAE

Fig. 204. Paralimnoria andrewsi (Calman) (2 mm)

Range: Caribbean region including south Florida (Intertidal and subtidal)

The species is the type and only member of the genus and is apparently widespread being first described from Christmas Island in the Indian Ocean.

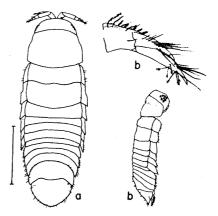


Figure 204. a. Paralimnoria andrewsi. b. Uropod. c. Lateral view.

- 1b. Branches of uropods dissimilar; exopod short with claw-like apex, endopod long with apex blunt ______2
- 2a. Mandibles without file-like ridges or rasp-like arrangement of scales (Fig. 205a)(Phycolimnoria)

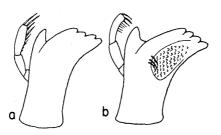


Figure 205. Mandibles of Limnoria (Limnoria) sp. and Limnoria (Phycolimnoria) sp. compared. a. L. (Limnoria). b. L. (Phycolimnoria).

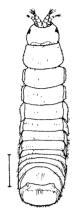


Figure 206. Limnoria (Phycolimnoria) algarum.

Fig. 206. Limnoria (Phycolimnoria) algarum (Menzies) (5.5 mm)

Range: Orgeon to southern California (Intertidal and subtidal)

The species is found in burrows that it makes in the holdfasts of the brown laminarian algae. The species is the only known North American *Limnoria* species that burrows exclusively in algae.

GENUS LIMNORIA (LIMNORIA)

Species of the genus *Limnoria* subgenus (*Limnoria*) are among the most economic important of the isopods. They attack wooden

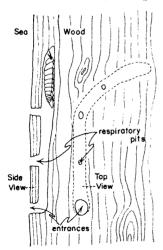


Figure 207. Limnoria species gnawing within a wooden pile. The burrows weaken the wooden structures in which they are located.

structures that are found in marine and brackish waters. This includes wooden bridge pilings, wharf pilings and any support pilings plus small boat hulls (Fig. 207). It is interesting to note that the water must be of a certain salinity for large populations of Limnoria species to flourish and become economically important. In the early history of San Francisco Bay in California, the water of the bay was composed mostly of the runoff of the rivers that flowed in and it was too brackish to support large populations of species of Limnoria. When the Central Valley irrigation projects were started and the amount of freshwater flowing into the bay decreased, the salt water of the ocean flowed into the bay. In a very short span of years the

wooden pilings of bridges and wharfs began to crumble. The higher salinity of the bay changed the environment so that large populations of *Limnoria* species were able to become established and flourish. The economic loss from the invasion of the isopods was put at several million dollars.

In the Baltic Sea, on the other hand, the water is not saline enough to support large populations of Limnoria species. (Nor was it high enough to support large populations of the mullusca Toredo which also bores into wooden structures.) A wooden ship, the Swedish warship Vesa, was launched into the brackish estuary. Because of some architectural miscalculations the ship capsized and sunk soon after the launching and for several hundred years remained at the bottom of the brackish estuary. Because the water in the several hundred years was never saline enough to support large populations of Limnoria, the ship was raised intact and can be seen today as a well preserved example of a 17th century ship.

Unfortunately most wooden ships sunk in the coastal waters of the world soon are attacked and eaten by *Limnoria* species. It has yet to be discovered if wooden ships sunk in the deep sea are subjected to *Limnoria* attacks. Sometimes a wooded ship is saved and preserved in shallow water if it is quickly covered by sand or mud and thus is immune to the isopods. However, the wrecked ships, being covered by sand and mud, elude scuba divers.

Many species of the genus are world wide in distribution and were distributed early in maritime history by establishing colonies in the hulls of wooden ships. Some species of the Old World are found in local situations in the New World where they have been introduced and live but do not spread. Much research into the choices of wood made by the species, the chemicals and paints for treating the wood and the general biology of the species has been undertaken, but the species are still able to cause many millions of dollars of damage.

In southern California the isopod species Caecijaera horvathi (p. 257) is found as a commensal in the burrows of Limnoria species. The seven species described here are all wood eaters of the subgenus (Limnoria) of the genus Limnoria. They are all found in the intertidal zone.

KEY TO SPECIES OF LIMNORIA

3a.	Pleotelson with 2, 3 or 4 tubercles on dorsum
3b.	Pleotelson with no tubercles, but with two (or one bifurcating) or no ridges on pleotelson 6
4 a.	Three or four tubercles present on pleotelson; outer margin of uropodal basis crenulate or smooth

4b. Two tubercles present on pleotelson; outer margin of uropodal basis smooth L. simulata

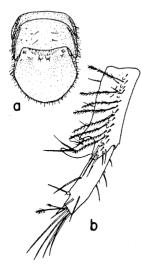
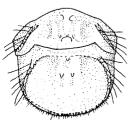


Fig. 208. Limnoria simulata (Menzies) (3 mm)

Range: Virgin Islands

Figure 208. a. Limnoria (L.) simulata. b. Uropod.

5a. Three tubercles present; outer branch of uropodal basis crenulate L. tripunctata



 α

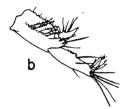


Figure 209 a. Limnoria (L.) tripunctata. b. Uropod.

Fig. 209. Limnoria tripunctata (Menzies) (2 mm)

Range: Bermuda, Gulf and Caribbean including southeastern United States; also southern California.

The species is found widespread in southern regions; whereas, *L. lignorum* is found widespread in northern regions.

5b. Four tubercles present; outer branch of uropodal basis smooth

L. quadripunctata

Fig. 210. Limnoria quadripunctata (Holthuis) (3.4 mm)

Range: Mid- to southern California

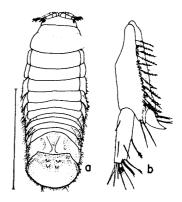


Figure 210. a. Limnoria (L.) quadripunctata. b. Uropod.

- 6a. Pleotelson with two conspicuous medial ridges or one bifurcating ridge
- 6b. Pleotelson with no conspicuous medial ridge ... L. platycaudata



Fig. 211. Limnoria platycaudata (Menzies) (2.5 mm)

Range: Caribbean Sea



Figure 211. a. Limnoria (L.) platycaudata. b. Uropod.

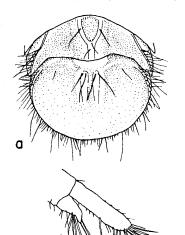
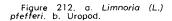
 

Fig. 212. Limnoria pfefferi (Stebbing) (3.5 mm)

Range: Indian Ocean and Miami, Florida



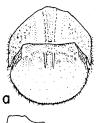




Figure 213. a. Limnoria (L.) saseboensis. b. Uropod.

Fig. 213. Limnoria saseboensis (Menzies) (3.5 mm)

Range: Sasebo, Japan and Miami, Florida

8b. Pleonal segment 5 with one medial ridge almost entire length; pleotelson with one bifurcating ridge on dorsum ...L. lignorum

Fig. 214. Limnoria lignorum (Rathke) (3.5 mm)

Range: Newfoundland to North Carolina and Alaska to California

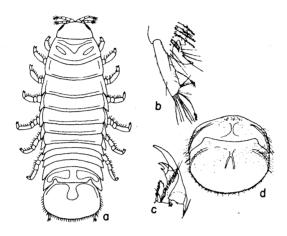


Figure 214. a. Limnoria (L.) lignorum, b. Uropod, c. Pergeopod I. d. Pleatelson.

The species is very important in the northern and temperate parts of North America and the world. It is also very common in Europe where it probably evolved as a species.

CYMOTHOIDAE

There are many species in this large important family of isopods. Most of them are fish parasites although they have been found on squid or not attached to any host at all. Some species have been shown to detach themselves from a host under unfavorable conditions and then reattach when conditions again become favorable. Members of the family get to be about 60 mm long, and they number among the largest of the isopods. The species are very much like members of the family Aegidae in general appearance, but cymothoids do not have the large eyes and symmetrical bodies that are characteristic of aegids. Cymothoids have a much greater dependence on their hosts than do aegids and this in part accounts for some of their differences. Species of the many genera of the family are found mainly as ectoparasites of fish as are aegid species, but there are other species that are found within the gill and mouth chambers of their hosts also.

The bodies of the clinging isopods are quite often much modified according to where they are found on the host. Those on the right side of the host's body can have their body axis twisted in one direction while those on the other side have their body twisted

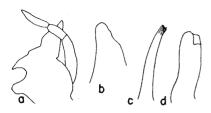


Figure 215. Mouth parts of Olencira praegustator. a. Mandible. b. Mandible without palp. c. Maxilla one. d. Maxilla two

in the opposite direction. More complex changes in appendage shape and mouth parts structure (Fig. 215) also take place. Loss or reduction in size of the eyes frequently occurs in animals that become parasites during their evolution, and cymothoid isopods are no exception. In a sense a parasite utilizes the eyes of the host to which it is attached.

The young of the species are generally much alike in appearance (Figs. 216, 217 and 218). They generally are free living, but begin

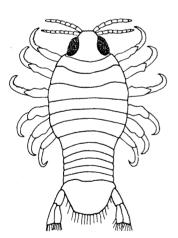


Figure 216. Cymothoa excisa, juvenile.

their parasitic life early. They have developed very large claws for attachment by the time that they reach the manca or six appendage stage. Indeed, the very young forms of many species are not known since they have not yet been found attached to a host. Attachment apparently does not take place immediately after release from the marsupium of the female. When they first attach to the host they are often males and their body is symmetrical. By the time that they are adults and have passed through several molts, they are asymmetrical and are females with brood pouches. The isopods are hermaphroditic, or more accurately andromorphic; i. e., they have the ability to change from male

to female, the male stage coming first.

Animals of different ages and sexes are found on one host fish. Generally only particular species are found on particular fish, but more likely the particular isopod parasite has several fish host species. Some described species are perhaps the young of other described species that are already known as adults. There

Cymothoid genera
of questionable validity include: Cterissa, Tetragonocephalon

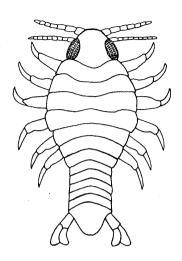
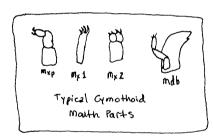


Figure 217. Agarna carinata, juvenile.

is much yet to be learned of the life cycle of the cymothoids. The fish themselves are frequently the host of one or more than one species of isopod, but usually two different species are not found on one host. The degree of specificity between host and parasitic species is much greater than that of the aegid species, probably because they are more dependent upon their host. Some species of cymothoids are found exclusively in the throat of certain fish species for instance. Since many host fish are commercially important, species of the family are included among the most important ecologically and economically important species of isopods.



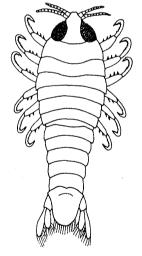


Figure 218. Mothocya nana, juvenile.

KEY TO GENERA AND SPECIES OF CYMOTHOIDAE

1a. Border between cephalon and peraeonal segment I trisinuate (with three lobes) or posterior border of cephalon produced into three lobes; cephalon never deeply immersed into peraeonal segment I

1b. Border between cephalon and peraeonal segment I broadly rounded (never trisinuate); cephalon deeply immersed into peraeonal segment I

(but check Olencira, P. 152)

2a. Posterior margin of uropodal rami with fringing setae; eyes very large; body symmetrical Aegathoa

"GENUS AEGATHOA"

The species of this genus are undoubtedly the young of other cymothoid species. Three specimens have been recorded from North American waters-two are from fish and the other is from both fish and a squid. They are not proper species and will undoubtedly be placed in other genera when their adult forms are known. The specimens are frequently found unattached to a host, hence, it is only with luck that the actual host is known.

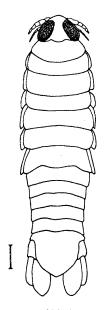


Figure 219. Aegathoa oculata.

Fig. 219. Aegathoa oculata (Say) (3 mm)

Range: Connecticut to Yucatan, Mexico, including West Indies

The species has been found as a parasite on squid and also on the fishes mullet and sand perch. There is no medial rostral projection from the frontal border of the cephalon and epimeral plates are visible in the dorsal view in this species.

Fig. 220. Aegathoa linguifrons Richardson 190/ Range: Trinidad

The rostral projection is different than that found in A. medialis.



Figure 220. Aegathoa linguifrons.

Fig. 221. Aegathoa medialis Richardson 1900 Range: Barren Island, Chesapeake Bay (5 to 45 m)

The broad rostral projection of the frontal margin of the cephalon set it apart from other species.



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Figure 221, Aegathoa medialis.

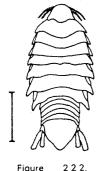
- 2b. Posterior margin of uropodal rami never with fringing setae; eyes if present, and moderately large 3

GENUS NEROCILA

The species of this genus are ectoparasites of fish. They are symmetrical and moderately large, and can be found on the skin or on a fin of their hosts.

KEY TO SPECIES OF NEROCILA

4b. Blind N. lanceolata



Nerocila

Fig. 222. Nerocila lanceolata (Say) (19 mm)

Range: Cumberland Island, Georgia

This species was collected around 1817 by the American naturalist, Thomas Say, and it has not been collected since. The host is unknown.

Nerocila excisa Rich. 1914

Fig. 223. Nerocila munda Harger 1873 (13 mm)

Range: Vineyard Sound, Woods Hole, Massachusetts

The species was found on the fin of Alutera schoepfii.

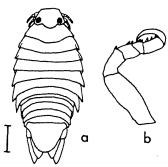


Figure 223. a. Nerocila munda. b. Peraeopod VII.

Fig. 224. Nerocila californica Schioedte and Meinert y (19 mm)

Range: Southern California to Panama & throughout Gulf of California.

The species is found on the fin of many species of fish. See &rusca(Arr).

This is now ir. syn. of N. acuminata (see Brusca, 1980)

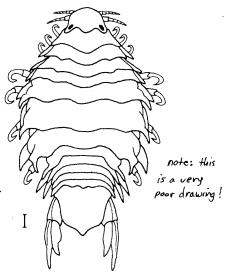


Figure 224. Nerocila californica.

6b. Posterolateral angles of last two peraeonal segments produced

N. acuminata

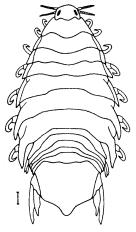


Figure 225. Nerocila acuminata.

Fig. 225. Nerocila acuminata Schioedte and Meinert v (21 mm)

Range: Virginia to Mexico and Bermuda

This external parasite has been found on dog fish and other species. Pterisopodus bartachi which was put in the family Pterisopodidae by the person who described it is probably only a small specimen of this species. Brosca (1978) claims this is the Atlantic analog of N. Californica; (1980) claims is sr. Syronym.

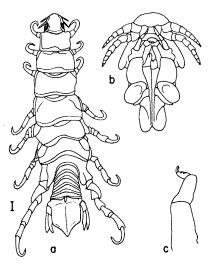


Figure 226. a. Olencira praegustator. b. Cephalon, ventral view. c. Maxilliped. (For mouth parts see Fig. 215).

Fig. 226. Olencira praegustator (Latrobe) (26 mm)

Range: New Jersey to Texas

The species is found on fish of the coastal and brackish waters of the east coast of North America. It has been found in the mouth cavity of menhaden fish, *Brevoortia*, off south and southeastern United States.

Fig. 227. Anilocra laticauda Milne-Edwards v (34 mm)

Throughout Caribbean b

Range: Maryland southward in us. probably only in Forida!

The species has been collected from the body surface of *Haemulon plumieri*, *Upeneus martinicus* and other fish. The pleotelson has an acuminate posterior margin and is widest in the distal two thirds of length.

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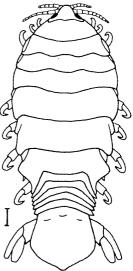


Figure 227. Anilocra

= A. mexcana, A. leachii & A. laevis!!

(not Ceratothoa laticanda milne Edwards)

Fig. 228. Anilocra plebia Schioedte and Meinert v (21.5 mm)

Range: Costa Rica and Central America

The pleotelson of this species has an acuminate posterior border that is widest at the proximal part.

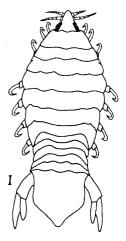


Figure 228, Anilocra plebia.

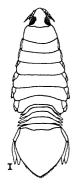


Figure 229. Anilocra acu-

Fig. 229. Anilocra acuta Richardson (34 mm)
Range: Savannah, Georgia to Louisiana

A gar pike was the host fish in Georgia. It was taken on *Lepidosteus osseus* in the western part of its range.

see bowmon, et al., 1977

Anilocra meridionalis Rich. 1914 (near Galapagos)

Anilocra laevis Miers 1877 (Peru; Mantinique)

(= A. laticaváa based on Martinique syntype; see

Brusca, in press).

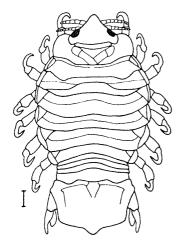


Figure 230. Braga cichlae.

Fig. 230. Braga cichlae Schioedte and Meinert V (17 mm)

Range: Northern South America to Brazil (Fresh and brackish waters)

The species is parasitic on fish that are found in rivers that empty into the Caribbean Sea.

Fig. 231. Braga occidentalis Boone (17 mm)

Range: Coast of California

The species has a rounded, not acuminate, posterior border on the pleotelson.

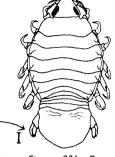


Figure 231. Braga occidentalis.

- 9a. Basal segments of antennae one dilated and touching; basal segments of antennae two compressed (Fig. 232a) 10

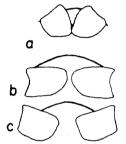


Figure 232. Antennal bases of Cymothoidae, ventral views. a. Contiguous. b. Slightly separate. c. Widely separate

10a. Male with large tuberous "shoulders" on posteriorlateral corners of peraeonal segments I; female with coxal extensions produced anteriorly; pleon deeply set into peraeon; first segment of pleon with lateral borders hid under peraeonal segment VII

Fig. 233. Ceratothoa impressa (Say) (33 mm)

Range: Atlantic coast and Gulf Stream

This species is a perasite in the mouth of flying fish and their near relatives. It has a large bilaterally symmetrical body, and the female is relatively narrower than the male. The dactylus on peraeopod III This is now Glossobius, by Bownan (1978).

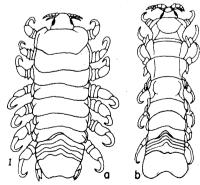


Figure 233. a. Ceratothoa impressa (male). b. Female.

is longer than the others and is a useful character for distinguishing species of the genus.

10b. Cephalon set only to moderate depth into peraeonal segment I; pleon constricted where attached to peraeon and edges visible in dorsal view on first pleonal segment Meinertia

GENUS MEINERTIA*

The species of this genus are large isopods which are parasites of large fish. They are found in the gill and mouth cavity and are frequently taken from the throat of their host. (Fig. 12g-h).

KEY TO SPECIES OF MEINERTIA

- 11a. Posterior margin of pleotelson rounded or pointed ______12
- 11b. Posterior margin of pleotelson truncate and very long13

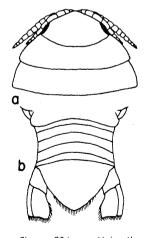


Figure 234. a. Meinertia transversa, anterior part. b. Posterior part.

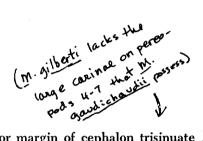
Fig. 234. Meinertia transversa Richardson 1900

Range: Key West to Delta of Mississippi River

The species is most probably based on a very young individual.

Fig. 235. Meinertia deplanata (Bovallius)
(18 mm)

Range: Coast of Haiti and West Indies



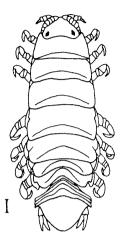


Figure 235. Meinertia deplanata.

13a. Posterior margin of cephalon trisinuateM. gaudichaudii

Fig. 236. Meinertia gaudichaudii (Milne-Edwards) (45 mm)

Range: Maxatlan, Mexico to west coast of South America (Chile, Galapagos, Peru).

The species is frequently taken from the throat of a *Thynnus* species and *Sarda chilensis*.

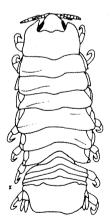


Figure 236. Meinertia gaudichaudii.

13b. Posterior margin of cephalon not trisinuate M. gilberti

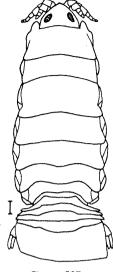


Fig. 237. Meinertia gilberti Richardson 1904 (20 mm)

Range: Mazatlan, Mexico to upper Gulf + w Baja.

The species was taken from a fish, Mugil hospes.

Figure 237.

- 14b. Antennae one widely separated at bases (Fig. 232c)16



Figure 238. Agarna carinata, lateral

1884

Fig. 239. Agarna carinata Schioedte and Meinert A (18 mm)

Range: Florida and West Indies

The species is the type-species of the genus.

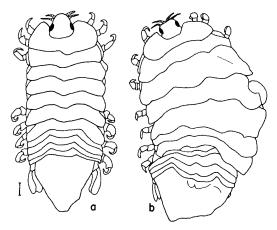


Figure 239. a. Agarna carinata (male). b. Female (Juvenile, Figure 217).

15b. Body margins evenly convex, not compressed; posterior border of pleotelson broadly rounded

Fig. 240. *Idusa carinata* Richardson v(13 mm)

Range: West coast of Panama

The species was found in the mouth of a species of fish of the genus *Mugil*.

Note: Only USUM type material of Idusa is I. carinata.

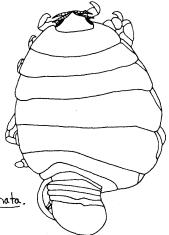


Figure 240. Idusa carinata.

- 16a. Pleon abruptly narrower than peraeon or narrowly attached to peraeon (body not forming continuous convex line between pleon and peraeon) 17
- 16b. Pleon and peraeon forming continuous convex body margin (or at least pleon and peraeon not abruptly constricted where they come together)

GENUS CYMOTHOA

The species of this genus are found on many of the economically important fish of the east coast of North America. They are parasites of the buccal region being found on the lip or within the mouth cavity itself.

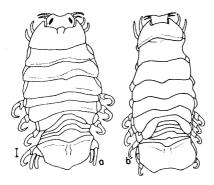


Figure 241. a. Cymothoa excisa (male). b. Female (Juvenile, Figure 216).

Fig. 241. Cymothoa excisa Perty v (23 mm)

Range: Massachusetts to Brazil including West Indies

The species has been found as a parasite on the lips of the chub, the gills of a sparid and on other fish. The female has rudimentary eyes and the male has small eyes. See Weinstein & Heek (1977).

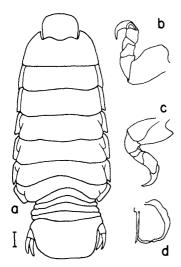


Figure 242. a. Cymothoa caraibica. b. Peraeopod VII. c. Peraeopod IV. d. Pleopod 2, male.

Fig. 242. Cymothoa caraibica Bovallius v (17 mm)

Range: West Indies, south of Haiti

The small eyes in this species are not visible in the dorsal view.

Cymothoo recta Dana 1853

Endemic to Hawaii. USNM has 7

lots (28977 through 28983)

Illa Al Mount land Latel A. C. Il in A

Fig. 243. Cymothoa oestrum (Linnaeus) (30 mm)

Range: Virginia to Venezuela

The species is blind. It is very abundant being found on at least five genera of near-shore fish. Usually it is found in the buccal cavity of the host, but it has been taken from the stomach of red fish after having been eaten.

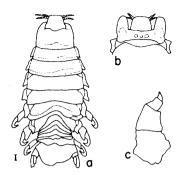


Figure 243. a. Cymothoa oestrum (male). b. Female, anterior part. c. Maxilliped.

Fig. 244. Cymothoa exigua Schioedte and Meinert V (20 mm)

Range: West coast of North America and Galapages Islands (Gulf of Colif to Ecvador)

The eyes of the parasite are moderately large. It was found in the mouth of a flat fish.

Louisiana by Comeaux, 1941 (probably in error)

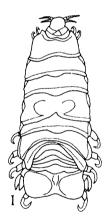


Figure 244, Cymothoa exigua,

17b. Pleon abruptly narrower than peraeon; frontal margin of cephalon broadly rounded; body nearly ovate or ovate

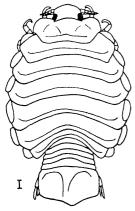


Figure 245. Telotha hanselii.

Fig. 245. Telotha henselii (von Martens)
(26 mm)

Range: Northern South America and West Indies

The species is found on cichlid fish and species of cat fish in brackish water.

18a. Pleon enveloped by peraeonal segment VII; only lateral borders of last two pleonal segments show in dorsal view

.....Mothocya (= Irona)

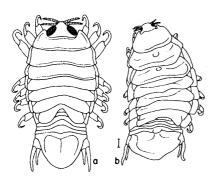


Figure 246. a. Mothocya nana, juvenile female. b. Male. (Juvenile, Figure 218).

Fig. 246. Mothocya nana Schioedte and Meinert 1864 (18 mm)

Range: Caribbean Sea, Bermuda and Brazil

The species is a non-obligate parasite found in the gill cavity of several species of fish.

GENUS LIRONECA

There are ten North American species in the genus. The animals are moderately large and are most frequently found in the gill cavities of their fish hosts. The species have moderately large eyes,

Bowman 1960

Livoneca was a typo-see Monod (1931) or Boman + Diaz-Ungria (1957) for explanation! and their body axes are symmetrical or slightly asymmetrical. The pleon is generally, but not always, broadly attached to the peraeon.

For key to E. Pacific KEY TO SPECIES OF LIRONECA Lironeca see Brusca (1978)

- 19a. Peraeon not enveloping pleon in any degree; body only slightly if at all asymmetrical; lateral edges of five pleonal segments visible on lateral margins 20
- 19b. Peraeon enveloping at least one pleonal segment; body symmetrical to greatly asymmetrical; lateral edges of four or five pleonal segments visible on lateral margins 22
- 20a. Peraeonal-pleonal separation straight 21
- 20b. Peraeonal-pleonal separation trisinuateL. redmannii

Fig. 247. Lironeca redmannii Leach (25 mm)
Range: West Indies to Brazil

The species was taken from the gill cavity of a kingfish and many other fish. (See also L. ovalis)

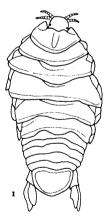


Figure 247. Lironeca redmannii.



Figure 248. Lironeca tenuistylis.

Fig. 248. Lironeca tenuistylis Richardson 1912 (13 mm)

Range: Fox Bay, Colon, Panama (E. gate of Canal)
The host fish was Anchovia browni.

= L. longistylis

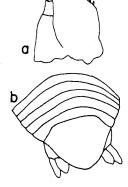


Figure 249. a. *Liron*eca ovalis, maxilliped. b. Posterior part.

Fig. 249. Lironeca ovalis (Say) (21 mm)

Range: Woods Hole, Massachusetts to Mississippi River and Texas

The species, although never well described, has been recorded as a parasite on many fish. It is most always taken from the gill chamber. It might be the same species as *L. redmannii* (p. 163)

Fig. 250. Lironeca vulgaris Stimpson 1857 (32 mm)

Range: Washington State to Baja California

The species is parasitic on the rock cod, flounder and fish of other genera. This is 5r.

syn of L. panameusis (see Brusco, 1978)

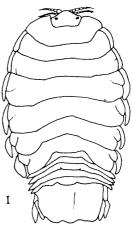


Figure 250. Lironeca vul-

23b. Body less than half as wide as long; proximal edge of pleotelson indented medially only L. symmetrica

Fig. 251. Lironeca symmetrica Van Name v (20.6 mm)

Range: Guyana and Caribbean Sea

The species is parasitic on river fish that are occasionally taken in the Caribbean Sea.

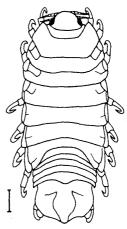


Figure 251. Lironeca

- 24a. Posterior border of pleotelson acutely pointed, rounded or broadly rounded 25
- 24b. Posterior border of pleotelson indented L. texana

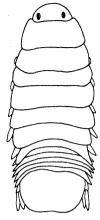


Figure 252. Lironeca texana,

Fig. 252. Lironeca texana Pearse $^{7932}_{\Lambda}$ (23 mm) Range: Off Padre Island, Texas (23 to 33 m)

Three fishes—the lizzard fish, the sea bass and the gaff-topsail catfish—serve as a host for this species.

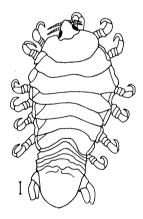


Figure 253. Lironeca californica.

Fig. 253. Lironeca californica Schioedte and Meinert v (16 mm)

1883 Alaska to M. Baja (Sahía Range: Mid-to southern California Son Quinta)

This species is parasitic on the "shiner."

26a. Frontal margin of cephalon rounded2726b. Frontal margin of cephalon truncateL. panamaensis

Fig. 254. Lironeca panamaensis Schioedte and Meinert v (27 mm)

Range: West coast of Mexico to Panama Host unknown. This is the jr. synonym of L. vulgaris (Brusco, 1978)

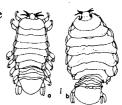


Figure 254. a. Lironeca pana-maensis (male). b. Female.

27a. Medial body axis straight L. convexa

Fig. 255. Lironeca convexa Richardson 1905 (21 mm)

Range: Southern California to Ecuador

The host is Chloroscombrus orgneta, a bait fish used for catching tuna.

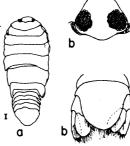


Figure 255. a. Lironeca b. Cephalon, bent convexa, b. upwards, c. Posterior part.

27b. Medial body axis curved markedly L. reniformis

Fig. 256. Lironeca reniformis Menzies and Frankenberg v (7 mm)

Range: Off Georgia Host unknown.

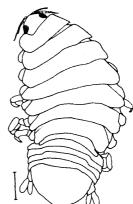


Figure 256. Lironeca reni-

CIROLANIDAE

Cirolanidae contains 12 genera divided into six with eyed species and six with blind species. One of the genera with blind species has two species and the other five have only one species each so there are only seven blind species. Of the genera that have eyed species one has only a single species and two have only two species, but the three other genera contain many species several of which are worldwide in distribution. The species in the genera in which most species are eyed live mainly in the near shore habitats. All the blind species, with the exception of a deep sea species, were found in caves on the approximate shore line of an ancient sea. The shore line of the ancient sea extended from northern Mexico to Yucatan, Mexico, then to Cuba.

The species are fairly common and are told from others by the presence of four or five free (one species has only one) visible pleonal segments. The number of prehensile and ambulatory pleopods vary. Some species have been found clinging on fish just as the fish parasites. The mouth parts are developed for biting and many species are scavengers. The maxillipedal palp has five articles of approximately equal width, and without large lobes. There are always three articles on the mandibular palp and the molar process is of a peculiar shape (Fig. 283a).

KEY TO GENERA AND SPECIES OF CIROLANIDAE

The following species, *Colopisthus parvus*, was keyed out in the family key (p. 111) because it is different from other flabelliferans in having no free pleonal segments.



Fig. 257. Colopisthus parvus (Richardson)

Range: Bermuda (Shallow water)

This unusually formed isopod is the typespecies of the genus. It was found among the corals. The pleon is apparently fused into one very short segment.

Figure 257. Colopisthus parvus.

Fig. 258. Bathynomus giganteus (Milne-Edwards) (280 mm)Range: Off the gulf and south Atlantic coast of United States

This species is truly a giant among the isopods growing to at least 280 mm or about 11 inches in length. Specimens have been taken in the nets of fishermen. and they are probably more commonly caught than reports from the literature indicate. They can probably avoid most deep sea bottom dredges. A species of the genus is found in the Indian Ocean and another in the Sea of Japan. The Japanese utilize the animals as food. The specimens of the genus are excellent examples to be used for the study of

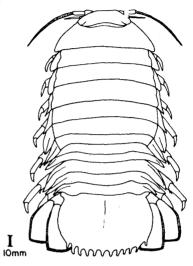


Figure 258. Bathynomus giganteus.

isopod morphology and anatomy since they represent a primitive type of isopod with five free pleonal segments and many other features of the primitive isopod. Use of microscope is not required in order to study them.

- 1b. Eyes visible in dorsal view (if present); posterior margin of uropodal rami and pleotelson various 2
- 2a. Basal segments of antennae one exit from cephalon at oblique or lateral angle; posterior border of pleotelson never with special pattern of setae or teeth on medial part of posterior margin

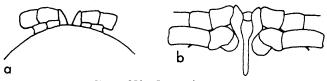
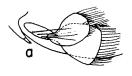


Figure 259. Bases of antennae, a. Coming off at right angles, b. Not coming off at right angles.

GENUS EURYDICE

Species of this genus are distinguished by the different patterns of fringing setae and spines along the posterior margin of the pleotelson (Fig. 260).



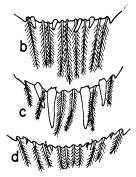


Figure 260. Some characters of Eurydice species, a, Uropod of E. littoralis. b. Posterior margin of pleotelson of E. branchuropus, c. of E. littoralis. d. of E. piperata.

KEY TO SPECIES OF EURYDICE

- 3b. Spines on posterior margin of pleotelson with two medial spines much longer than others E. littoralis

Fig. 261. Eurydice littoralis (Moore) (6 mm)

Range: Puerto Rico to Georgia (37 m)

On beaches in tropical regions the species can be very abundant in the surf and it will attack swimmers or skin divers. The bites can be annoying but they cause no damage to humans.

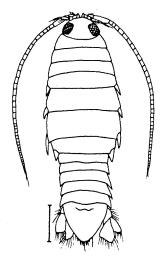


Figure 261, Eurydice littoralis.

4b. Posterior margin of pleotelson concave E. spinigera

Fig. 262. Eurydice spinigera (Hansen) (9 mm)

Range: Eastern Atlantic

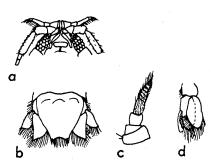


Figure 262. a. Eurydice spinigera, anterior part ventral aspect. b. Posterior part. c. Antenna one. d. Pleopod 2, male.

5a. Eight spines on posterior margin E. convexa

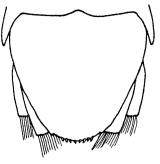


Fig. 263. Eurydice convexa (Richardson)

Range: Cape San Blas, Florida

Figure 263. Eurydice convexa.

5b. Four spines on posterior margin of pleotelson ______6

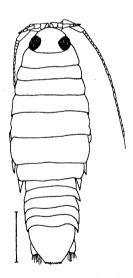


Fig. 264. Eurydice piperata (Menzies and Frankenberg) (5 mm)

Range: Off coast of Georgia (49 to 137 m)

Figure 264. Eurydice piperata.

6b. Spines moderately large, longer than posterolateral corners ... 7

7a. Uropodal rami distally truncate E. caudata

Fig. 265. Eurydice caudata (Richardson)

Range: Catalina Island, California to Sea Of Cortez; so. to Ecvador. (see Bowman, 1977)

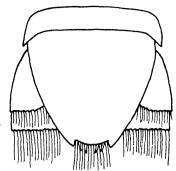


Figure 265. Eurydice caudata.

7b. Uropodal rami not distally truncate E. branchuropus

Fig. 266. Eurydice branchuropus (Menzies and Barnard) (3 mm)

Range: South of Pt. Loma, California (44 m)

this is a jr. syronym

of E. caudata (see Bowman, 1977)

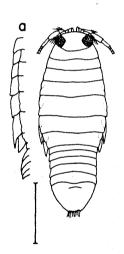


Figure 266. a. Eurydice branchuropus, lateral margin. b. Dorsal view.

- 8a. Eyes absent or present; peraeopods I, II, and III only prehensile; all others ambulatory or modified natatory (Fig. 31b).....9
- 8b. Blind; peraeopods all prehensile; only peraeopod I prehensile or no peraeopods prehensile 28
- 9a. First and second pairs of pleopods different from one another; first pair with both branches indurate and operculate, second pair normal or like third and fourth pairs Conibera

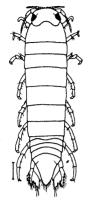


Figure 267. Conilera cylindracea.

Fig. 267. Conilera cylindracea (Montague) (10.5 mm)

Range: South Carolina to Mississippi (103 to 291 m)

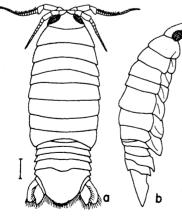
Two species of the genus have been described from North American waters. C. cylindracea is parasitic on the pilchard, the young of which are caught and sold as sardines. Species of the genus in Europe are abundant as scavengers, eating dead fish found on the sea coast. The species C. stygia Packard (25 mm) from well in Monterey, Mexico, is blind and has been recorded only once and perhaps is not a member of the genus.

10a. Cephalon only slightly produced medially if at all (Fig. 272)

10b. Cephalon with medial spatuliform process (Fig. 208)

Excirolana

- KEY TO SPECIES OF EXCIROLANA Rich. 1912.



18: 3 distinguishing affibutes characterize species of

Excirolana: (D

Figure 268. a. Excirolana mayana. b. Lateral view.

Four of the five species of Excirolana from North America are from the Pacific coast, and they are included in the key. E. mayana from the Caribbean Sea is not included in the key.

Fig. 268. Excirolana mayana (Ives) (10 mm)

Range: Caribbean Sea (Intertidal)

= <u>Cirolana mayana</u>

see menzies 9 Glynn (1968) d Lemos de Castro (1969)

Richardson says this species occurs in tropical E. Pacific.

MARINE ISOPOD CRUSTACEANS 175 11a. Pleotelson rounded and crenulate E. linguifrons antennae are setose Fig. 269. Excirolana linguifrons (Richardson, M(3.8 mm) a Range: Monterey Bay, California to so. Calif. this structure is on the pleatelson Figure 269. a. Excirolana linguifrons, anterior part. b. Posterior parts. c. Maxillij ed. Pleotelson obtusely rounded; spatulate frontal process almost as long as broad; distance between eyes about diameter of eves E. chiltoni Fig. 270. Excirolana chiltoni (Richardson, 1908) (11 mm)Range: San Francisco, California to So. Calif. includes: E. Kincaidi (Hatch, 1947) E. vancouverousis (Fee, 1926) E. japonica Richardson, 1912 Figure 270. a. Excirolana chiltoni, anterior part. b. Posterior part. All Bruse & Jones, 1981 for syronymy 12b. Pleotelson obtusely rounded or angulate; spatulate frontal process longer than broad; distance between eyes greater 13a. Posterior margin of pleotelson obtusely angulate

Excirolana hirsuticanda Menzies 1962 (chile) - frontal process Excirolana chilensis Rich. 1912 (chile) Exciralana braziliensis Rich. 1912 (Gulf of Calif. to N. Chile; also in Ponibles ...



Figure 271. Excirolana kincaidi.

Fig. 271. Excirolana kincaidi (Hatch 1947)
(7.6 mm) this is now a jr. syn. of E. chiltoni

Range: Washington and Oregon coasts to so. Calif.

The species was taken under dead crabs at the midtidal level on a beach so it apparently is a scavenger.

13b. Posterior margin of pleotelson obtusely rounded

E. vancouverensis

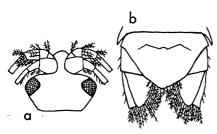


Figure 272. a. Excirolana vancouverensis, anterior part. b. Posterior part.

Fig. 272. Excirolana vancouverensis (Fee) (7 mm)

Range: Vancouver Island, British Columbia

note: Fee described this species as a subspecies of E. <u>Chiltoni</u>; Hatch (1947) appears to have exected it to species level. It is now a yr. syn. of E. <u>Chiltoni</u>.

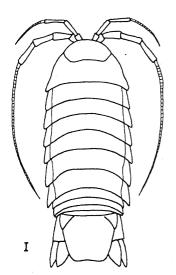


Figure 273. Specirolana bolivari.

Fig. 273. Specirolana bolivari (Rioja) (33 mm)

Range: Caves near Valles, San Luis Potosi, Mexico

This large blind species was found in the freshwater of the limestone caves found in northern Mexico. Fig. 274. Speocirolana pelaezi (Bolivar y Pieltain) (26 mm)

Range: Cave, San Luis Potosi, Mexico

This species was found in the same limestone cave region as the above species. The posterior margin of the pleotelson in this species is concave in contrast to the broadly rounded posterior margin in S. bolivari.

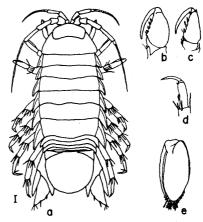


Figure 274. a. Specirolana pelaezi, b. Peraeopod II. c. Peraeopod III. d. Peraeopod IV. e. Pleopod 2, male.

14b. Eyes absent or present; endopods of pleopods 1 to 4 usually fringed with setae Cirolana

GENUS CIROLANA

For Key & California Cirolana see Brusca & Ninos 1978

The species of the genus are common, widespread and abundant in almost every near shore isopod habitat. They are found in all marine waters at many depths and are abundant in brackish waters as well. The species have been found in freshwater caves and have occasionally been recorded as being attached to fish. It was perhaps as fish parasites that the ancestors of the cave cirolanids reached the freshwater habitats. In North American waters, both marine and brackish water species are abundant. So far there have been 16 species recorded, the largest number of species for any genus with North American representatives.

One of the characters usually mentioned as being definitive of species of *Cirolana* is the presence of five free pleonal segments plus a pleotelson. In many instances the first pleonal segment is completely covered by peraeonal segment VII or in some instances only the very narrow posterior margin of pleonal segment 1 shows. Some illustrations of the same species will show five free segments, others only four.

Cirolanids are distinguished by reference to tooth and setal arrangement on the posterior margin of the pleotelson. Also the configuration of the pleotelson margin and the structure of the pleopodal rami are to be considered. Pleopods 1 and 2 are similar and in most species the medial angle of the uropodal basis is produced.

By examining the above characters and recording the nature of the second male pleopod, the species of the genus can be separated from one another. All characters mentioned should of course be mentioned in new species descriptions when they are made.

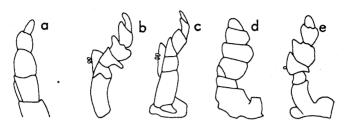


Figure 275. Maxillipeds of Cirolana species. a. C. concharum. b. C. obtruncata. c. Cirolana sp. d. C. polita. e. C. joanneae.

KEY TO SPECIES OF CIROLANA

15a. With eyes 15b. Blind (or eyes who pigmentation) ntides 11-21 flagellar articles Natatolana Fig. 276. Cirolana californiensis Schultz 1966 (8 mm)Tanner & Catalina Range: Coronado, \(\Lambda \) Canyon on shelf off southern California (4 San Diego Trough), S. to Peru-chile Trench. 40-2,000 m.
This blind, pigmentless species is found at a depth much greater than that of any other species of Cirolana in North American waters. This is the Sr. synonym of C. deminuta (see Brusca & Ninos 1978) ___ margin of pleotelson with both spines

16a. Lateral margins of cephalon pointed; anterolateral margins of cephalon straight; cephalon medially produced

and setap

Figure 276. Cirolana cali-

1.	Margin of pleotelson with fine setae but without spines; miped with 1 coupling hook; edges of pleonite 5 not hidden by pleonite 4; distal margin of pleotelson truncate, but strongly scalloped
	C. joanneae Margin of pleotelson with spines; maxilliped with 2 coupling hooks; edges of pleonite 5 hidden by pleonite 4; distal margin of pleotelson subacute 2
2.	Without eyes, or eyes unpigmented; antenna 2 with flagellum of 10–21 articles; frontal lamina narrow; pleotelson with 8–12 spines; uropods not notched apically; rarely encountered in depths less than
-	100 m C. californiensis Blindness extremely rare; antenna 2 with flagellum of 22–32 articles; frontal lamina broad; pleotelson with 8–32 spines; uropods notched apically, or not notched; rarely encountered in depths greater than
3.	100 m Uropodal rami with deep apical notch; pleotelson margin always
_ _	with 8 spines Uropodal rami without apical notch; pleotelson margin with at least 9-32 spines C. parva C. harfordi
C. ancuato	: plt. w/setae & spines; mxp w/1 coupling hook: S.F. Bay only
16b	. Lateral and anterolateral margins of cephalon rounded17
	. Edges of pleonal segment five covered by pleonal segment
17b	Edges of pleonal segment five not covered by pleonal segment four
	eyes often red 12 flagellar anticles
	278. Cirolana joanneae Schultz (3 mm)
Ran so	278. Cirolana joanneae Schultz (3 mm) ge: Santa Cruze Canyon on shelf off outhern California (218 m)
Ran so	278. Cirolana joanneae Schultz (3 mm) ge: Santa Cruze Canyon on shelf off outhern California (218 m)

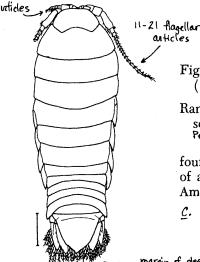
Key to the Species of Cirotana Known From

e examined (including Schultz's types) conform to the figure presented Menzies and Ged We have redrawn these structures from Schultz's type (Fig. 1). Schultz's figure of the first maxilla differs conisderably n that given by Menzies and George. Examination of this appendage a both holotypes has revealed Schultz's figure to be misleading, in that igured the exopod (outer ramus) only (see Fig. 2, this paper).

lenzies and George suggested that C. deminuta could represent a enic species, as evinced by the reduced size of the seventh percopods. erials we have studied, however, reveal that the reduction of the nth pereopods is, like reduction of the eyes, a variable character. The e of this reduction is not known; however, when reduction does occur right and left pereopods are always affected, suggesting a genetic s (rather than predation or some other exogenous factor).

here are now 12 species of Cirolana known from the eastern Pacific: californiensis Schultz 1966 (= C. deminuta), southern California to e; C. joanneae Schultz 1966, also known from the submarine canyons outhern California; C. harfordi (Lockington, 1877), a shallow water ies ranging from British Columbia to central west Baja California; varva Hansen 1890 (= C. diminuta), a eurythermal, circumtropical, ow water species known in western America from Point Conception fornia) south at least to central west Mexico; C. bathyalis Menzies

15b. Blind (or eyes who pigmentation) C. californiensis



Notatolana

Fig. 276. Cirolana californiensis (Schultz 1966 (8 mm)Tanner & Cataline

n the time locality /Down Chile Tree LI

Range: Coronado, \(Canyon on shelf off southern California (4 San Diego Trough), S. to Peru-chile Trench. 40-2,000 m.
This blind, pigmentless species is found at a depth much greater than that

of any other species of Cirolana in North American waters. This is the Sr. synonym of

C. deminuta (see Brusca & Ninos, 1978)

_ margin of pleotelson with both spines Figure 276: Cirolana caliand setae

16a. Lateral margins of cephalon pointed; anterolateral margins of cephalon straight; cephalon medially produced

metacirolana

Fig. 277. Girolana sphaeromiformis Hansen, v (4.3 mm)

Range: St. Thomas, West Indies, Bermuda, Puerto Rico, Howaii, Pacific Costa Rica.

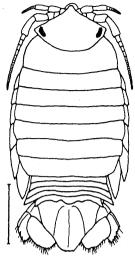


Figure 277. Cirolana sphaeromiformis.

- 16b. Lateral and anterolateral margins of cephalon rounded17

eyes often red 12 flagellar anticles 1966 Itz 1 (3 mm)

Metacirolana 1966 Fig. 278. Cirolana joanneae Schultz A (3 mm)

Range: Santa Cruze Canyon on shelf off southern California (218 m)

margins of unopods strongly servate-

pleatelson with setae, but.

Figure 278. Cirolana joanneae.

no spines

18a.	Epimeral extension of peraeonal segment VII completely visible in dorsal view 20
18b.	Epimeral extension of peraeonal segment VII not completely visible in dorsal view19
19a.	Posterior margin of pleotelson and endopod of uropod broadly rounded

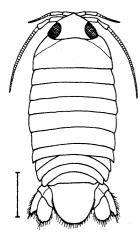


Fig. 279. Cirolana minuta (Hansen) (4.8 mm)

Range: St. Thomas, West Indies

Figure 279. Cirolana minuta.

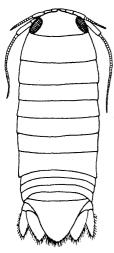


Figure 280. Cirolana albida.

Fig. 280. Cirolana albida (Richardson)

Range: Sugarloaf Key, Florida

The species is close to *C. parva* in appearance and it should be more closely examined.

20a.	Posterior margin of pleotelson broadly rounded or trunca	
	Posterior margin of pleotelson obtusely rounded or narrow rounded (more or less pointed)	ly
21a.	Posterior margin truncate	
	Posterior margin broadly rounded	
	Posterior margin broadly truncate; exopod and endopod	
	uropod apically rounded	

Fig. 281. Cirolana obtruncata (Richardson)

Range: Puerto Rico and Jamaica (Shallow water)

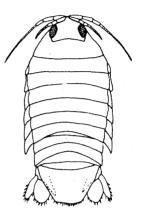


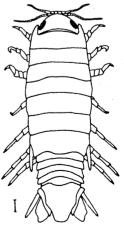
Figure 281, Cirolana obtruncata.

22b. Posterior margin narrowly truncate; exopod or uropod narrow, endopod with deep notch on exterior margin C. concharum

Figs. 282 and 283. Cirolana concharum (Stimpson) (23 mm)

Range: Nova Scotia to South Carolina (Surface to 33 m)

The species was taken on muddy and sandy bottoms.



Cirolana Figure 282. concharum.

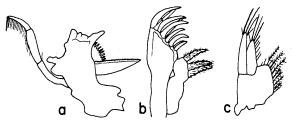


Figure 283. Mouth parts of C. concharum, a. Mandible, b. Maxilla one, c. Maxilla two.

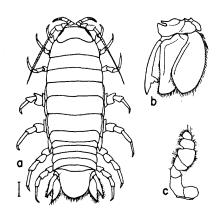


Figure 284. a. Cirolana borealis. b. Pleopod 2, male. c. Maxilliped.

Fig. 284. Girolana borealis (Lilljeborg) (12 mm)

Range: Atlantic coast of North America (55 to 1478 m)

 Fig. 285. Girolana polita (Stimpson) (16 mm)
Range: Bay of Fundy to Georgia (2 to 587 m)

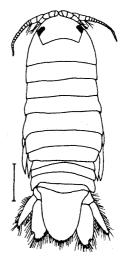
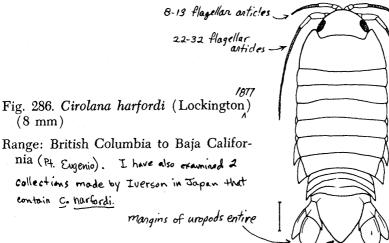


Figure 285. Cirolana polita.

- 24a. Lateral margins (i. e., epimeral extensions) of peraeonal segment VII much wider than pleon 25
- 24b. Lateral margins (i. e., epimeral extensions) of peraeonal segment VII not much wider than pleon 26
- 25a. Posterior margin of pleotelson narrowly obtusely pointed; notches not present on exterior margin of endopods



magin of pleotelson with 10-32 spines

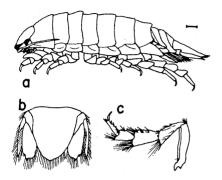
Figure 286. Cirolana harfordi. 

Figure 287. a. Cirolana impressa. b. Posterior part. c. Peraeopod VII.

Fig. 287. Cirolana impressa (Harger) (21 mm)

Range: New York to North Carolina (183 to 587 m)

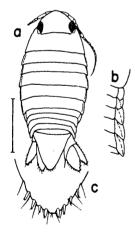


Figure 288. a. Cirolana diminuta, b. Lateral margin. c. Detail, posterior margin of pleotelson.

Fig. 288. Cirolana diminuta Menzies, 1962. (3.2 mm)

Range: San Quintin Bay, Baja California (Shallow water)

This was made the jr. synonym of C. Parva by Menzies & Glynn (1968:38). A circumtropical Species, on our coast Pt. Concaption South to at least (osta Rica. However, Bruce & Bowman (1982) removed it from synonymy!

26b. Peraeonal segment VII as long (wide) as other peraeonal segments; frontal margin of cephalon very slightly produced27

Fig. 289. Cirolana gracilis (Hansen) (8 mm)

Range: St. Thomas, West Indies

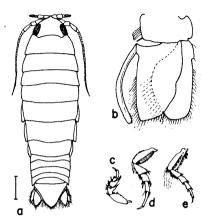
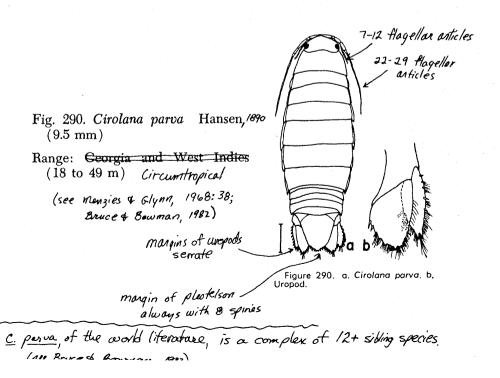


Figure 289. a. *Cirolana gracilis*. b. Pleopod 2, male. c. Peraeopod II. d. Peraeopod VII. e. Peraeopod V.



28a. All peraeopods prehensile Haptolana

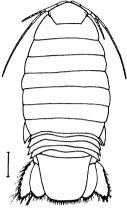


Figure 291, Haptolana trichostoma.

Fig. 291. Haptolana trichostoma (Bowman) (13.8 mm)

Range: Camaguey Province, Cuba (Cave)

This blind, unpigmented cave species has seven pairs of prehensile peraeopods. Whether it clings to a fish host or to the wall of the cave is unknown.

28b.	Peraeopod I only, or no peraeopods prehensile	29
29a.	Peraeopods I prehensile	30
29b.	No peraeopods clearly prehensile	31
	Exopod of pleopod 2 and endopod of pleopods 3	
	two segments	Cirolanides

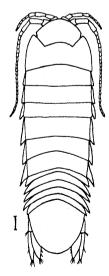


Figure 292. Cirolanides texensis.

Fig. 292. Cirolanides texensis (Benedict) (17 mm)

Range: South central Texas (Caves and underground water)

This species was first taken from a well at San Marcos, Texas, and was the first North American freshwater cirolanid described. Fig. 293. Antrolana lira (Bowman) (12 mm) Range: Madison Cave, Virginia

The species is the type and only one in the genus, and is morphologically much different from the circlanids of Texas, Mexico and Cuba. It is also the most geographically isolated of the species.

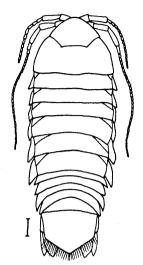


Figure 293. Antrolana lira.

31a. Able to roll into ball

Creaseriella

Fig. 294. Creaseriella anops (Creaser) (18 mm)

Range: Yucatan, Mexico (Cave)

This species was found in a cave in the same region where very primitive terrestrial isopods were obtained also.

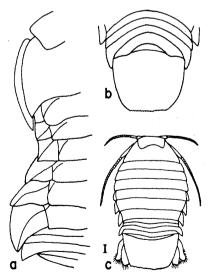


Figure 294. a. Creaseriella anops, detail lateral margin. b. Posterior part. c. Dorsal view.

31b. Not able to roll into ball Troglocirolana

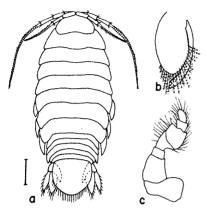


Figure 295. a. Troglocirolana cubensis. b. Pleopod 2, male. c. Maxilliped.

Fig. 295. Troglocirolana cubensis (Hay) (7 mm)

Range: San Isidro, Cuba (Cave)

The first pair of legs are somewhat prehensile; all others are ambulatory.

AEGIDAE Leach Dana, 1853

The species of this family are moderately large, common ectoparasites on body fishes, sharks and skates. They generally are found clinging to the outside skin of the fish host, but occasionally they have been found just inside the gill slit. Apparently the species are able to live when not attached to the hosts because many times they have been collected in bottom samples. Fish on which the isopods have been clinging are usually scarred or wounded where the claws of the specimens were attached. They are probably of ecological importance since many economically valuable fish—halibut, haddock and cod—are parasitized by the species of the family.

The species with very few exceptions have eyes which are generally quite distinct and sometimes extremely large. All peraeonal segments are well developed, and the pleon is usually composed of five segments plus a pleotelson; but in some species there are only four free segments plus the pleotelson. The pleotelson and the uropods usually form a broad caudal fan. The first three pairs of peraeopods are modified into distinct prehensile hooks and the next four are usually ambulatory. The distinction between the two types of peraeopods is usually quite marked. On the tip of the maxillipedal palp there are usually large teeth or spines, but they are sometimes absent in gravid females.

KEY TO GENERA AND SPECIES OF AEGIDAE

- 1a. Eyes present or absent; body depressed; maxillipedal palp composed of only two articles; front of cephalon covers peduncle of antenna one; flagellum of antenna one of four to six articles

GENUS AEGA

Fourteen species of this genus have been recorded from North America. The species are very common ectoparasites of fish, but are occasionally taken from the bottom not attached to fish.

KEY TO SPECIES OF AEGA

2a.	Eyes moderately large to large3
2b.	Eyes small A. microphthalma
	No Fig. Aega microphthalma (Dana)
	Range: Monterey Bay, California
	This species is perhaps the young of one of the west coast species he genus. It is difficult to tell since there are no figures available.
3a.	Pleotelson triangulate or with posterior margin smooth and produced medially; crenulations or teeth never present; lateral margins straight (or slightly concave) or convex
3 b .	Pleotelson various; posterior margin many times with crenulations or teeth
4a.	Lateral margins of pleotelson straight (or slightly concave); eyes touching

Aega deshaysiana (H. Milne-Edwards, 1840). Circumtropical. West Indies; Hawaii (Rich.).

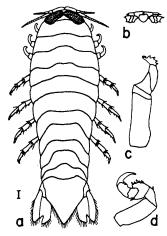


Figure 296. a. Aega antillensis. b. Frontal lamina. c. Maxilliped. d. Peraeopod 11.

Fig. 296. Aega antillensis (Schioedte and Meinert) (35 mm)

Range: West Indies (298 to 422 m)

Alis is the jet syranger of A. Led engine a Committee, Miss (committee)

4b. Lateral margin of pleotelson convex; eyes large but separate

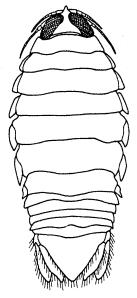


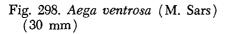
Figure 297. Aega psora.

Fig. 297. Aega psora (Linnaeus) (16 mm) Range: North Greenland to Florida Keys

(60 to 1280 m)

The fish that are frequently to

The fish that are frequently the host are bottom fishes—halibut and skates—so when they become detached from one host there is probably little difficulty in finding a new one. Cod fish are also parasitized by the isopod.



Range: Greenland (37 to 571 m)

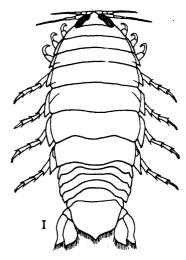


Figure 298. Aega ventrosa.

- 6a. Pleotelson broadly rounded, posterior margin various; i.e., smooth or with crenulations or medial notch or both; basal segments of antennae usually not broad and set in front of cephalon (dorsal view)
- 6b. Pleotelson truncate, sinuate, concave, never with crenulations; basal segment of antennae broad and set in front of cephalon (dorsal view)

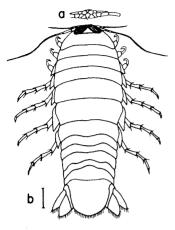


Figure 299. a. Aega tenuipes, frontal lamina. b. Dorsal view.

Fig. 299. Aega tenuipes (Schioedte and Meinert) (11.5 mm)

Range: Mid- to southern California

Richardson (1901, 1905) reports it

from Cuba - following 54 m's original

record. I have no idea how Schultz

Come up with California on this one.

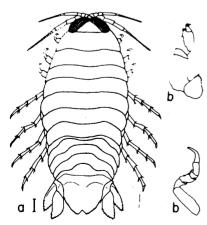


Figure 300. a. Aega incisa. b. Maxilliped. c. Peraeopod II.

Fig. 300. Aega incisa (Schioedte and Meinert) (18 mm)

Range: Georgia to Florida (481 to 805 m) 9b. Eyes separated medially; medial length of first pleonal segment about as long as others

Fig. 301. Aega arctica (Lutken) (34 mm)

Range: Greenland

The species is parasitic on Somniosus microcephalus.



Figure Aega arotica. Frontal lamina.

10a. Crenulated posterior margin of pleotelson with small medial projection; less than five pleonal segments apparent 11

10b. Simple broadly rounded, crenulated posterior margin; all five pleonal segments apparent and about equal in length A. symmetrica

Fig. 302. Aega symmetrica Richardson) (17 mm)

Range: Alaska to mid-California (75 to 878 m)

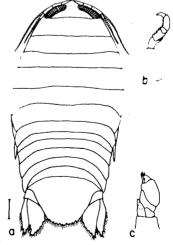


Figure 302. a. Aega symmetrica. b. Peraeopod III. c. Maxilliped.

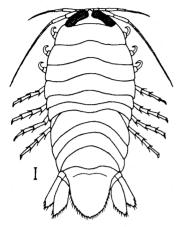


Fig. 303. Aega dentata (Schioedte and Meinert) (7.5 mm)

Range: Cuba

Figure 303. Aega dentata.

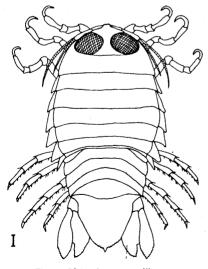


Figure 304. Aega gracilipes.

Fig. 304. Aega gracilipes (Hansen) (21 mm)

Range: North Atlantic to Gulf of Mexico (1335 to 2787 m)

12a. Posterior margin of pleotelson sinuate. E. ecarinata Fig. 305. Aega ecarinata (Richardson) (24 mm) Range: West Indies (298 to 422 m) Coventry (1944) reported this species from off Acapulco. Figure 305. a. Aega ecar-inata. b. Maxilliped. 12b. Posterior margin of pleotelson truncate, concave or convex 13a. Posterior margin of pleotelson truncate; eyes touching medial-A. crenulata Fig. 306. Aega crenulata (Lutken) (28 mm) Range: Greenland The species is parasitic on the Greenland shark. crenulata. Figure 306. a. Aega e Peraeopod II. c. Maxilliped. 13b. Posterior margin of pleotelson concave or convex; eyes large, but not touching medially 14

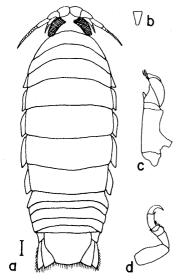


Fig. 307. Aega lecontii (Dana) (20 mm)

Range: Mid- and southern California

Figure 307. a. Aega lecontii. b. Frontal lamina. c. Maxilliped. d. Peraeopod II.

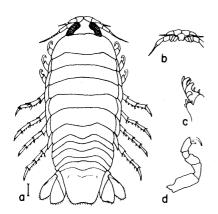


Figure 308. a. Aega webbii. b. Frontal lamina. c. Apex of maxilliped. c. Peraeopod II.

Fig. 308. Aega webbii (Guerin) (16 mm)

Range: Off Fernandina, Florida (609 m)

15a. Blind; peraeopods 1, II and III without expanded propodal segments (Fig. 307d); dactyli abruptly curved in middle and terminating in very sharp point (Fig. 298); pleon abruptly narrower than peraeon Syscenus

Fig. 309. Syscenus infelix (Harger) (28 mm)

Range: Atlantic coast south to Delaware Bay (146 to 1170 m)

This large isopod is blind. All peraeopods are prehensile, but there is a gradual change towards more ambulatory kind from front to rear. The species is apparently the only one in the genus.

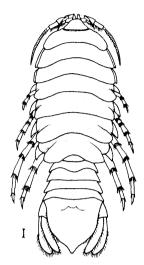


Figure 309. Syscenus in-

15b. Eyes present; peraeopods I, II and III with propodus more or less expanded and armed with spines (Fig. 312b); dactyls forming large evenly rounded hooks on peraeopods I to III; pleon slightly narrower than peraeon

Rocinela

GENUS ROCINELA

The species in this genus, like those in Aega, have been found both as parasites on the skin of fish and apparently living free on the bottom. Sometimes they have been taken from the gill cavity of fish. The species are common and there are about 15 of them in North American waters.

KEY TO SPECIES OF ROCINELA

16a. Frontal part of cephalon greatly produced more than length of eyes 17
16b. Frontal part of cephalon broadly rounded or produced, but never more than length of eyes 19
17a. Frontal projection of cephalon with single process 18
17b. Frontal projection of cephalon trifid R. tridens

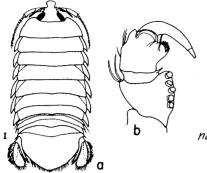


Figure 310. a. Rocinela tridens. b. Per-

Fig. 310. Rocinela tridens (Hatch) (25 mm)

Range: San Juan County, Washington State

note: heavily selose uropoods unusual "rostrum"

18a. Frontal margin broadly rounded wider along most of length than distance between eyes R. cubensis

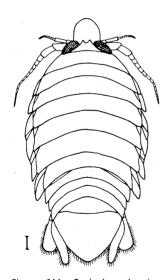


Figure 311. Rocinela cubensis.

Fig. 311. Rocinela cubensis (Richardson) (16 mm)

Range: Cuba (261 m)

18b. Frontal margin gradually tapers to truncate end, end much Fig. 312. Rocinela cornuta (Richardson) (30 mm)

Range: Off Shumagin Bank, Alaska (1143 m) ~ Gulf of Alasko

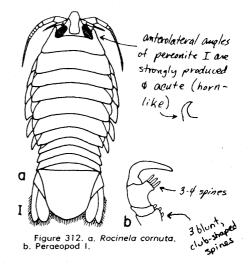


Fig. 313. Rocinela laticauda (Hansen) (40 mm)

Range: West coast of Mexico

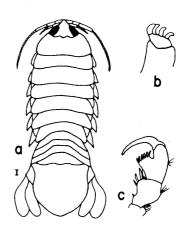


Figure 313. a. Rocinela laticauda. b. Apex of maxilliped. c. Peraeopod II.

19b. Body less than two and one-half times as long as broad; pleotelson narrower than peraeon 20

20a. Eyes touching medially R. oculata

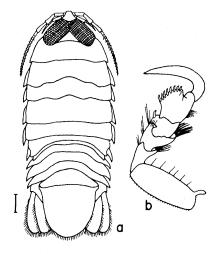


Fig. 314. Rocinela oculata (Harger) (13.5 mm) Range: Near Bermuda (461 m)

Figure 314. a. Rocinela oculata. b. Peraeopod I.

20b.	Eyes not touching medially
21a.	Five pleonal segments distinct, but not necessarily along entire width 22
21b.	Only four pleonal segments distinct (they are distinct along entire width)—16 pleonite 1 more-or-less hidden by personite. 24
22a.	First pleonal segment not distinct along entire width23
22b.	First pleonal segment distinct along entire width R. americana

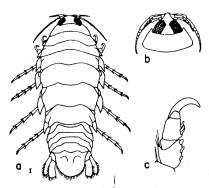


Figure 315. a. Rocinela americana (female). b. Male. c. Peraeopod II.

Fig. 315. Rocinela americana (Schioedte and Meinert) (20 mm)

Range: North Atlantic and Maine (155 to 287 m)

23a. First pleonal segment distinct only on lateral marginsR. signata

Fig. 316. Rocinela signata (Schioedte and Meinert) (13 mm)

Range: West Indies and Central America (4 to 48 m)

The species was found on groupers. It also has been taken on bottoms of coarse sand and in coral formations.

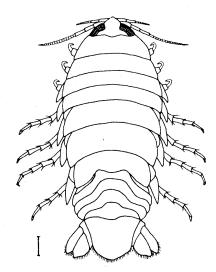


Figure 316. Rocinela signata.

23b. First pleonal segment distinct only medially R. aries

Fig. 317. Rocinela aries (Schioedte and Meinert)

(20 mm)

Range: Gulf of California and west coast of Mexico

= R. <u>signata</u> (syn. by Honges 4 Glym, 168) (see Moreira, 1971, 1972, 1977)

(see Bowman, 1977)

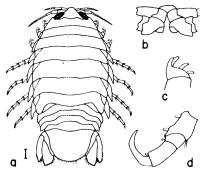


Figure 317. a. Rocinela aries, b. Frontal lamina. c. Apex of maxilliped. d. Peraeopod II.

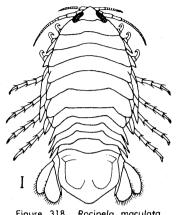


Fig. 318. Rocinela maculata (Schioedte and Meinert) (23 mm)

Range: Greenland

24b. Posterior margin of cephalon not trisinuate	24b.	Posterior	margin	of	cephalon	not	trisinuate		25
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25a. Eyes almost touching medially; edges of pleonal segments not acutely pointed.

R. insularis

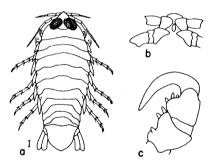


Figure 319. a. Rocinela insularis. b Frontal lamina. c. Peraeopod II.

Fig. 319. Rocinela insularis (Schioedte and Meinert) (25 mm)

Range: Florida to Mississippi and West Indies (415 to 499 m)

- 25b. Eyes large but not almost touching; edges of pleonal segments mostly all acutely pointed 26
- 26a. Frontal margin of cephalon only slightly produced and only acutely pointed; peraeonal segment VII much shorter than others
- 26b. Frontal margin of cephalon moderately produced or truncate (if broadly round then peraeonal segment VII much shorter or equal to other peraeony in length)

27a. Peraeonal segment VII laterally quite like pleonal segments 1, 2 and 3, and also about same length along entire width

R. bellicens

Fig. 320. Rocinela bellicens (Stimpson) (22 mm)

I are rounded

Range: Alaska to southern California (9 to 1258 m)

This species is a non-obligate parasite on cod and sculpin and other fish. At times it is found under shells and stones on intertidal beaches.

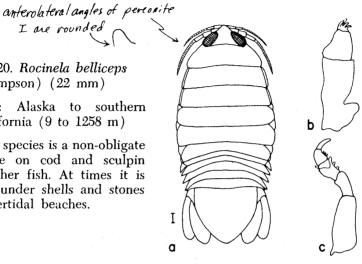


Figure 320. a. Rocinela belliceps. b. Maxilliped. c. Peraeopod III.

27b. Peraeonal segment VII with epimeral extensions much larger than lateral ends of pleonal segments 1 and 2 and also shorter (or thinner) medially R. tuberculosa

Fig. 321. Rocinela tuberculosa (Richardson) (13 mm)

Range: Southern part of Gulf of California (15 to 18 m)

The species has small tubercles on the dorsal surface.

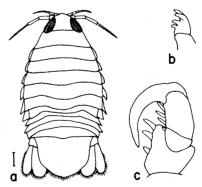


Figure 321. a. Rocinela tuberculosa. b. Apex of maxilliped. c. Peraeopod II.

28a. Frontal margin of cephalon broadly rounded or with truncate frontal margin

28b. Frontal margin of cephalon produced into knob (Fig. 322a, b) R. dumerilii

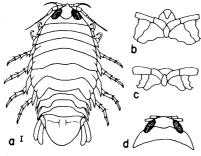


Figure 322. a. Rocinela dumerilii (male). b. Frontal lamina (male). c. Frontal lamina (female). c. Cephalon (female).

Fig. 322. Rocinela dumerilii (Lucas) (30 mm)

Range: Cuba (421 m)

29a. Frontal margin of cephalon not greatly produced; peraeopods with comparatively elongate segments (Fig. 323d) R. propodialis

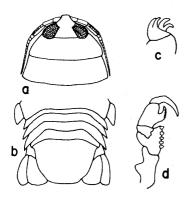


Figure 323. a. Rocinela pro-podialis, anterior part. b. Posterior part. c. Apex of maxilliped. d. Peraeopod III.

Fig. 323. Rocinela propodialis (Richardson) (22 mm)

Range: Port Townsend, Puget Sound, Washington (27 to 48 m)

29b. Frontal margin of cephalon produced, ending truncate; peraeopods with comparatively compressed segments (Fig. 324b) R. angustata Fig. 324. Rocinela angustata (Richardson) (28 mm)

Range: Alaska to mid-California (123 to 461 m)

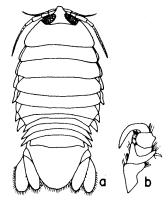


Figure 324. a. Rocinela angustata. b. Pergeopod I.

EXCORALLANIDAE

This small family is represented in North American waters by only one genus, Excorallana, of 13 species. The species are moderately large and the posterior part of the body in many species is covered with long setae which sometimes are thick and look like encrusting protozoans. This latter character is only found in some members of the corallanid genus Alcirona. The species of the excorallanids, however, differ from those of Alcirona in the structure of the mouth parts. In species of excorallanids the mandibles are very long and the molar process is rudimentary (Fig. 3415). There is a characteristic uncinate process on the first maxilla (Fig. 325b). The maxillipedal palp is narrow and the second to last article is elongate (Fig. 156a). Although species of the single genus are free living, they occasionally are found as external parasites on fish. The eyes are extremely large in most species, and in some they cover the whole length of the frontal margin, in effect forming one large eve.

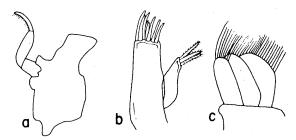


Figure 325. Mouth parts of Excorallanidae species. a. Mandible, b. Maxilla one, c. Maxilla two.

1549

KEY TO SPECIES OF EXCORALLANA

- 1a. Eyes large taking up about 1/3 or more of frontal margin; pleotelson various ______3
- 1b. Eyes moderately large, not taking up 1/3 or more of frontal margin of cephalon; pleotelson truncate or broadly rounded 2
- 2a. Pleotelson truncate; pleotelson much narrower than peraeon

 E. subtilis

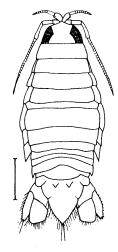


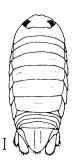
Fig. 326. Excorallana subtilis (Hansen) (4.7 mm)

Range: St. Thomas and West Indies (Shallow water)

Figure 326. Excorallana subtilis.

2b. Pleotelson broadly rounded; pleotelson continuous with peraeon E. berbicensis

Fig. 327. Excorallana berbicensis (Boone) (3 mm)



Range: Rio Berbice, Guyana

Figure 327. Excorallana berbicensis.

- 3a. Eyes large, but extending with only small medial across gap across frontal margin of cephalon
 3b. Eyes large, but not extending across frontal margin of cepha-
- 3b. Eyes large, but not extending across frontal margin of cephalon 5
- 4a. Pleotelson with posterior margin broadly rounded; no notches on lateral margins E. warmingii

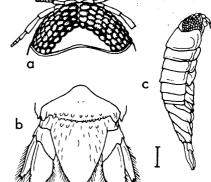


Fig. 328. Excorallana warmingii (Hansen) (9 mm)

Range: West Indies

Figure 328. a. Excorallana warmingii, cephalon. b. Posterior part, c. Lateral view.

4b. Pleotelson with posterior margin pointed; notches present on lateral margins E. oculata



Fig. 329. Excorallana oculata (Hansen) (11.5 mm)

Range: West Indies

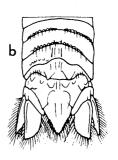
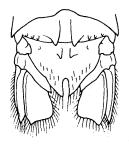


Figure 329. a. Excorallana o cullata, cephalon. b. Posterior part.



Range: West Indies

(11 mm)

Figure 330. Excorallana fissicauda, posterior part.

Range: Southern California and northern Mexico (Shallow water)

Fig. 330. Excorallana fissicauda (Hansen)

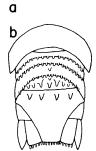


Figure 331. a. Excorallana truncata, anterior part. b. Posterior part.

Fig. 332. Excorallana kathae (Menzies)

Range: San Quintin Bay, Baja California (Shallow water) to So. California.

now a jr. syn. of E. truncata (see Delaney, 1982)

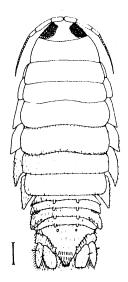


Figure 332, Excorallana kathae.

8b. Pleon continuous with peraeon; first and part of second pleonal segment enclosed within margins of peraeonal segment VII

E. mexicana

Fig. 333. Excorallana mexicana (Richardson) (8 mm)

Range: Gulf of Mexico (Shallow water to 48 m)

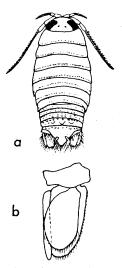


Figure 333. a. Excorallana mexicana. b. Pleopod 2, male.

9a.	Cephalon with dorsal and frontal tuber	cles	10
	Cephalon smooth with no tubercles		
	Cephalon with six large tubercles		serticornis

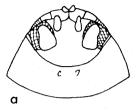


Fig. 334a. Excorallana sexticornis (Richardson)

Range: Key West, Florida



Figure 334, a. Excorallana sexticornis. b. E. quadricornis.

10b.	Cephalon with less than six tubercles 11
	Cephalon with four tubercles E. quadricornis
	Fig. 334b. Excorallana quadricornis (Hansen) (12 mm)
	Range: West Indies and Bermuda (Shallow water)
	The species was found in grass patches and in sponges.
11b.	Cephalon with two dorsal and one medial rostral tubercles
	E. tricornis

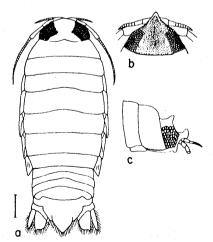
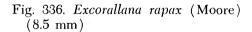


Figure 335. a. Excolallana tricornis (female). b. Cephalon (male). c. Cephalon lateral view (male).

Fig. 335. Excorallana tricornis (Hansen) (12 mm)

Range: Southern Florida, West Indies and Central America (44 to 503 m) 

Range: Southeastern United States and Puerto Rico (46 to 137 m)

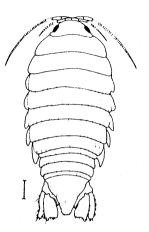


Figure 336. Excorallana rapax.

12b. Body with lateral margins ovate or convex

13

13a. Antenna one not extending beyond posterior margin of cephalon E. antillensis

Fig. 337. Excorallana antillensis (Hansen) (16 mm)

Range: Florida and West Indies (Intertidal)

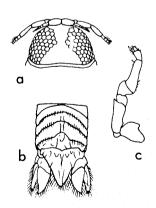


Figure. 337. a. Excorallana antillensis, cephalon. b. Posterior part. c. Maxilliped.

The female of the species lacks the three tubercles on the cephalon. Other females of other species can perhaps be keyed out at this point so it is very necessary to consider just where the specimens were caught and to use a male for identification if possible to be correct in the determination of *Excorallana* species.

CORALLANIDAE

This small family of Flabelliferans are set apart from other species on particulars in the structure of the mouth parts. The distal half of the mandibles are narrow and directed inwardly. There is no molar process on the mandible in most species. The second to the last article of the maxillipedal palp is not elongate as in species of Excorallanidae. The mouth parts are illustrated in figures 333 and 341. There are five or three free pleonal segments plus a pleotelson present. Much work must be done with actual specimens of members of the family so that a more rigid definition of it can be made.

KEY TO THE GENERA OF CORALLANIDAE

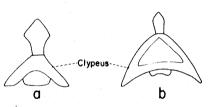


Figure 338. Clypeus of two corallanid species compared. a. Tridentella virginiana. b. Alcirona krebsii.

Range: Chesapeake Bay (148 m) and (southern California)

This species was taken in Chesapeake Bay and also in southern California. It is not a well defined genus and species and more specimens should be collected. It is rare among isopods to have the same species present on both coasts of North America, except for those species easily transported by man.

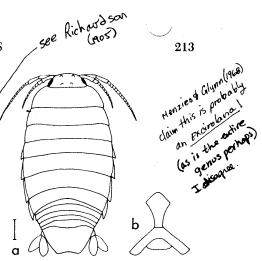


Figure 339. a. *Tridentella virginiana*. b. Clypeus and frontal lamina.

GENUS ALCIRONA - Ant 1 with on 2 peduncular articles.

This genus is not well defined as are some other isopod genera. There are five free pleonal segments in A. krebsii and three in A. hirsuta.

Figs. 340 and 341 Alcirona krebsii (Hansen) (11 mm)

Range: Yucatan, Mexico; St. Thomas, W. I.; Bermuda (50 to 56 m)

This species is found in sponges and dead coral and as an ectoparasite on the fin of a grouper. The species is not really modified as a parasite so that its presence on a fish was probably only casual.

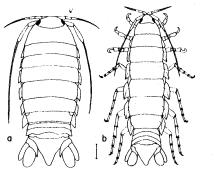


Figure 340. a. Alcirona krebsii (male). b. Female.

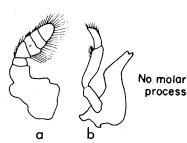


Figure 341. Mouth parts of Alcirona krebsii, a. Maxilliped, b. Mandible,

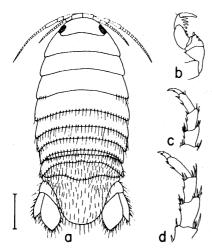


Figure 342. a. *Alcirona hirsuta*. b. Peraeopod I. c. Peraeopod IV. d. Peraeopod VII.

Fig. 342. Alcirona hirsuta (Moore) (6 mm)

Range: St. Thomas, W. I. (37 to 42 m)

SUBORDER MICROCERBERIDEA MICROCERBERIDAE

There are only about 22 described species of this suborder and family of isopods and only five of them have been recorded from North America. All 22 species are placed in one genus, *Microcerberus*, which apparently is represented on all continents of the world except Antarctica. The blind, pigmentless species are very tiny, and are the smallest known isopods. They most nearly resemble the anthurids to which they are closely related. The North American members of the genus sometimes get to be slightly over one millimeter long. They live among the fine grains of sand on intertidal beaches, especially where there is calcareous sand as on many tropical beaches. They are at times found in great abundance.

Freshwater species have been taken from the sand on the banks of streams in Hungary and from subterranean water in caves in Yugoslavia, but most species were collected from the beaches along the Mediterranean Sea. Like many tiny animals, they have been found only in places where particular people who have knowledge of their habitats have gone, and consequently there are many beaches of the world where no one has ever looked for them.

The species of *Microcerberus* live in sand like their nearest relatives the anthurids, but the *Microcerberus* species do not burrow into the sand as do the anthurids, but are found between the grains of sand. Species of animals that live between grains of sand are called psammophils or sand loving animals. All of them are very

tiny. Another species of isopod has been found exclusively in the sand of beaches (*Caecianiropsis psammophila*, p. 257), but it like the anthurids is large and lives among, not between, the sand grains. *Microcerberus abbotti* was found near to where *C. psammophila* was found in California. Three of the five known North American species were found on the small Bahaman island of Bimini so there are probably many more unrecorded species in subtropical and tropical North America.

The species are thought to be relict species; i. e., they have a morphological structure that is characteristic of the majority of animals of an earlier geological era or time. They are thought to approximate what the early anthurids were like. There are many characters of the species that can be called primitive, but there are also many modifications which are adaptations for living in the psammitic environment. No one has ever made a definitive, comparative study of the animals.

Since they are extremely small, a very good compound microscope is necessary for observing them. Very fine instruments are also necessary in order to properly dissect them. For easy observation, the animals must be mounted on slides. They can be collected from the sand by gathering sand in which they are thought to live and putting it into a pail and washing it. The animals will come off the sand and get into the wash water that is then strained through a very fine mesh screen. A screen made from a woman's silk or nylon stocking will do nicely. The animals can then be placed in 70% alcohol as is done for other isopods.

Fig. 343. Microcerberus mexicanus (Pennak) (1 mm)

Range: West coast of Mexico (Intertidal in sand)

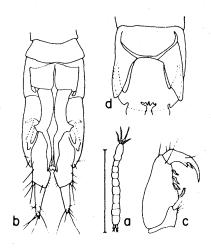


Figure 343. a. *Microcerberus mexicanus*, b. Ventral aspect posterior part with uropods, c. Peraeopod I. d. Ventral aspect posterior part without uropods.

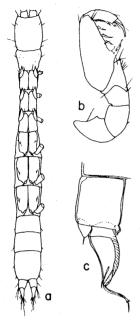


Figure 344. a. Microcerberus abbotti. b. Peraeopod I. c. Pleopod 2, male.

Fig. 344. Microcerberus abbottii (Lang) (1 mm)

Range: Monterey Bay, California (Intertidal in sand)

The species was taken from the wet sand of an intertidal beach. It was found in the same general habitat as the asellote, *Caecianiropsis psammophila* which is larger although it too has only been found in beach sand.

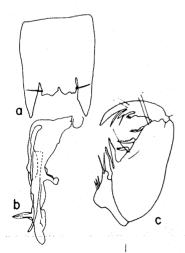


Figure 345. a. Microcerberus mirabilis, edge of a peraeonal segment. b. Pleopod 2, male. c. Peraeopod I.

Fig. 345. *Microcerberus mirabilis* (Chappuis and Delmare Deboutteville) (1 mm)

Range: Bimini, Bahamas (Near surface to 80 cm in sand)

Fig. 346. *Microcerberus renaudi* (Chappuis and Delmare Deboutteville) (1 mm)

Range: Bimini, Bahamas (To 40 cm in sand)



Figure 346. a. Microcerberus renaudi, uropod. b. Pleopod 2, male.

Fig. 347. Microcerberus littoralis (Chappuis and Delmare Deboutteville) (1 mm)

Range: Bimini, Bahamas (Near surface of sand)

This species was found in the reefs near the location where the other two species from Bimini were found.

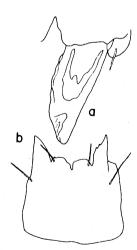


Figure 347. a. Microcerberus littoralis, pleopod 2, male. b. Edge of a peraeonal segment.

SUBORDER GNATHIIDEA GNATHIIDAE

Species of the family Gnathiidae are found in the deep sea as well as in the shallow waters off most of the continents of the

world. Specimens range from small to moderately large and there are well over 65 known species in the group. The family is divided into six genera but many of them are based on only one or a few species. The largest number of species is in the nominal genus *Gnathia*. Because females look so much different than males, early taxonomists placed the males in the genus *Gnathia* and the females

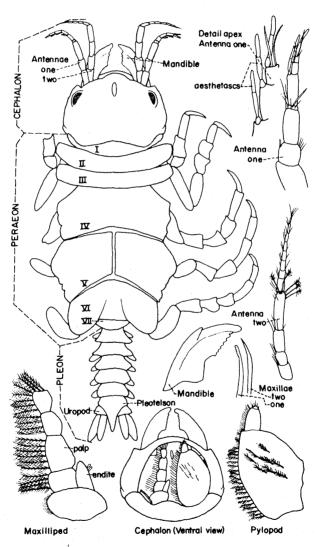


Figure 348. Gnathiidea (General nomenclature).

in a separate genus. The juveniles were placed in *Pranzia*. There are still many males, females and juveniles that have not been associated as a species unit.

Male gnathiids with their large mandibles are like termite or ant workers in appearance. However, when the ventral aspect of the male cephalon is examined, the mandibles are discovered to be apparently ineffectual as gnathal structures and are not even able to transport anything to the buccal cavity. The mandibles are apparently secondary sexual characters such as are found in many lucanid beetles that have mandibles 1/3 the length of their body but have a rudimentary or no mouth (see Fig. 349a). Female

gnathiids have no mandibles. The males are free living on the bottom, and they are many times taken in sponges or from the burrows of tubiferous worms. Apparently they obtain their food by straining the mud on the bottom through their maxillipeds.

The body of the gnathiid, both male and female, has only five pairs of peraeopods. It is the presence of the five pairs of peraeopods and the presence of the very large mandi-

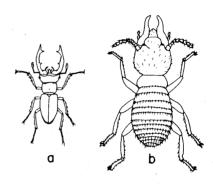


Figure 349. Two insects that resemble a gnathiid, a. Lucanid beetle. b. Termite.

bles that set the group apart from other isopods. Their bodies are highly calcified and elaborately ornamented. Some species are quite rococo in appearance (Fig. 359).

The females are also easily distinguished from other species of isopods, but there is no immeditae reason to believe that they are in any way related to the males. They have a very large, rotund peraeon with a translucent wall that is large even if not filled with eggs. The cephalon is narrow and generally with a pointed or rounded produced rostrum, with proportionately large eyes and without mandibles or without a conspicuous buccal cavity as in the males. The pleon is narrow and without any apparent long spines or processes. The pleotelson is "T" shaped or triangulate. The pleon in males and females is very much the same and is the most conspicuous reason why male and female, although placed in different genera, were not placed in even more widely divergent groups by early zoologists. The juveniles (Fig. 350) resemble the female in size and

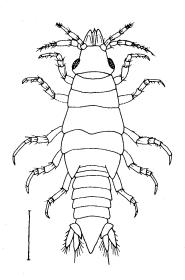


Figure 350. Gnathia cerina, a juvenile gnathiid.

general body configuration. Juvenile males have small mandibles protruding from the frontal margin of the cephalon. For most species where the juvenile is known it is a parasite on the body and sometimes in the gill chamber of fish.

The cephalon of the male is large and broad and with a very large distinct buccal cavity. The first pair of peracopods or what really should be peracopods, the pylopods, forms a covering over the large buccal cavity (Fig. 348), and the only mouth appendage that is well developed except for the large mandibles is the maxilliped which is found within the buccal cavity. The first and second maxillae are rudimentary structures. On the top of the buccal cavity in some species there is a hole

that opens to the dorsal surface (perhaps an artifact of preservation?). Its function is not known. The pylopods are fringed with large plumose setae as are the articles of the maxillipeds, and they most probably serve as filter feeder mechanisms.

Eyes usually are present, even in many deep water forms, and they are at times on short stalks. The female has a very small cephalon compared to that of the male. The eyes are about the same size of a comparable size male and they consequently appear large on the small cephalon of the female. Apparently the mouth parts of the female are modified for sucking. Only the pylopods and maxillipeds are present. The cephalons of the females of different species are very similar, and it is difficult, if not impossible, to determine the species of a female gnathiid if only female specimens are present. There are some differences in the pylopods, pleons and pleotelsons of females of different genera, however.

The species of the genus are occasionally found in large numbers in relatively shallow water. Benthic samples contain males, females and juveniles so the juveniles are apparently non-obligate parasites on the fish. Without a doubt, they play an important role in the ecology of the benthic ecosystem. Their general abundance and the presence of particular species on particular bottom types are facts that demonstrate that gnathiids deserve much more attention of investigators.

Only two genera have representatives in North American waters—Gnathia and Bathygnathia. Bathygnathia contains only one species. In the key female gnathiids are keyed out as if they were an independent group. If males and females are taken at one collection site, then it is safe to assume that they are of the same species. However, if more than one species of male are taken then it is difficult to state which male belongs with which female without careful study.

See Winn key"

KEY TO THE GENERA AND SPECIES OF GNATHIIDAE

- 1a. Peraeon with segments distinct; mandibles large and visible in dorsal view; pleon abruptly narrower than peraeonal segment VI
- 1b. Peraeon round and large, segments not distinct; cephalon small and narrow; pleon also small and narrow

 Female gnathiid (Figs. 21 and 360c)
- 2a. Blind; front of cephalon with frontal line expanded into very long pointed process Bathygnathia

not keved

Fig. 351. Bathygnathia curvirostris (Richardson) (11 mm)

Range: Georges Bank and south of Martha's Vineyard (709 to 1232 m)

It would be interesting to know if the species are parasites on deep water fish, but so far no juveniles have been recorded. The species is the type and only one in the genus.

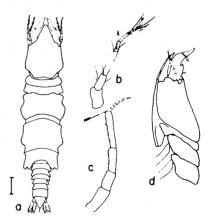


Figure 351. a. Bathygnathia curvirostris. b. Antenna one. c. Antenna two. d. Anterior part, lateral view.

GENUS GNATHIA

For the discussion of the genus see the general discussion under the family.

3a. Blind (Careful! One species has the eyes visible only in the ventral view.)

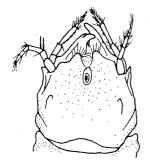


Fig. 352. Gnathia coronadoensis (Schultz) (3.5 mm)

Range: Coronado Canyon on shelf off southern California (812 m)

Figure 352. Gnathia coronadoensis.

3b.	With eyes	4
4a.	Eyes visible in dorsal view	5
4b.	Eyes visible in ventral view only	ata

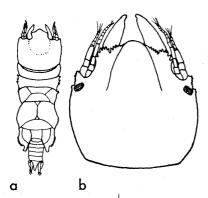


Fig. 353. Gnathia serrata (Richardson) (3.4 mm)

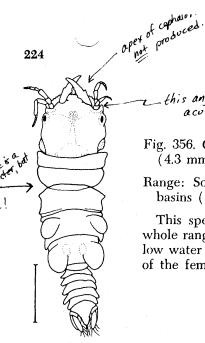
Range: South of Martha's Vineyard (713 m)

Figure 353. a. Gnathia serrata. b. Cephalon, ventral aspect.

5a. Cephalon longer than wide G. triospathiona Fig. 354. Gnathia triospathiona Boone, 1918 (2.9 mm)Range: Gulf Stream off Key West, Florida (199 m) Figure 354. Gnathia triospathiona. 5b. Cephalon shorter than wide 6 6a. Eyes not stalked 7 6b. Eyes on short stalks G. clementensis Fig. 355. Gnathia clementensis (Schultz) (8.5 mm) Range: San Clemente Canyon on shelf off southern California (162 m)

Figure 355. Gnathia clementensis.

7a. Frontal margin of cephalon crenulate; cephalon about as long as wide G. crenulatifrons



this angle more rounded than acute!

Fig. 356. Gnathia crenulatifrons Monod 1926 (4.3 mm)(oceanside)

Range: Southern California coastal shelf / and basins (9 to 1258 m) to Monterey Bay.

This species is extremely abundant over its whole range. It is also found in relatively shallow water as well as deep water. The host fish of the female is still unknown.

356. Gna-Figure thia latifrons.

7b.	Not as above8
8a.	Frontal margin of cephalon with more than one frontal process, extension or crenulation 9
8b.	Frontal margin of cephalon with only single or without large frontal process
9a.	Frontal margin without large medial process; eyes small and surrounded by long tubercles

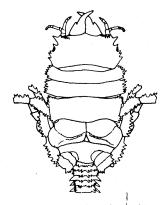


Figure 357. Gnathia multi-

Fig. 357. Gnathia multispinis (Richardson')

Range: Off Delaware (1765 m)

The specimen from which the posterior part of the pleon is missing is 5.5 mm long.

- 9b. Frontal margin of cephalon expanded into large rounded process; eyes surrounded by short tubercles 10

Sanctaecrucis

Fig. 358. Gnathia hirsuta, Schultz (4 mm)

Range: Santa Cruz Canyon on shelf off southern California (218 m)
(see 5chultz 1972: 1/12)

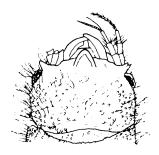
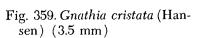


Figure 358. Gnathia hirsuta.



Range: North Atlantic, 72° N X 58° W (212 m)

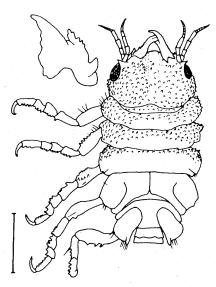


Figure 359. Gnathia cristata.

Fig. 360. Gnathia cerina (Harger) (4.8 mm)

Range: Gulf of St. Lawrence to south of Martha's Vineyard (18 to 891 m)

This species is found in the northern waters off the Atlantic coast of North America. The female is a parasite on sculpin, cod and other fish.

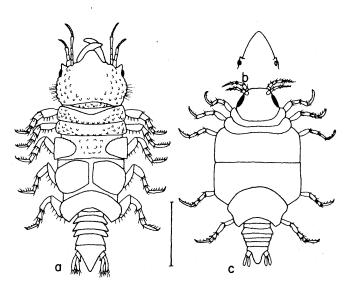


Figure 360. a. Gnathia cerina (male). b. Frontal view cephalon female. c. Female.

Fig. 361. Gnathia steveni (Menzies) (2.3 mm)

Range: Northern Baja California (Shallow water) Collected in King Horbor by D. Stroughn Euroeys.

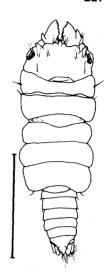


Figure 361. Gnathia steveni

- 12b. Eyes large or prominent; frontal margin with three small lobes, teeth or crenulations of about equal length; mandibles various
- 13a. Eyes prominent; hole or conspicuous deep spot medial on cephalon G. trilobata

Fig. 362. Gnathia trilobata (Schultz) 1966 (5 mm)

Range: Coronado Canyon on shelf off southern California (812 m)

This species was caught in the same canyon as *G. coronadoensis*, but it is not blind as is that species.

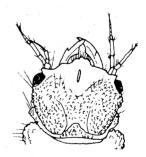


Figure 362. Gnathia triliobita.

- 13b. Eyes of moderate size; cephalon depressed, but without hole or conspicuously deepened spot on dorsal surface 14
- 14a. Cephalon rectangulate; anteriolateral corners prominent

 G. elongata

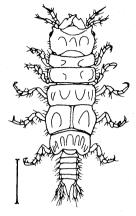


Fig. 363. Gnathia elongata (Kroyer) (4 mm) Range: Greenland (90 to 101 m)

Figure 363. Gnathia elongata.

14b. Cephalon not especially rectangulate and without prominent anterolateral corners 15

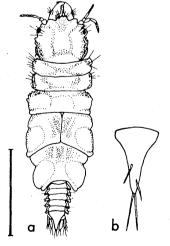


Figure 364. a. Gnathia productatridens. b. Pleotelson.

Fig. 364. Gnathia productatridens (Menzies and Barnard) (3.2 mm)

Range: Pt. Conception to Santa Barbara, California (73 to 91 m)

Fig. 365. Gnathia tridens (Menzies and Barnard) (3 mm)

Range: Pt. Conception, California (11 to 27 m)

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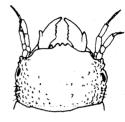


Figure 365, Gnathia tridens.

SUBORDER ASELLOTA

The Asellota are small isopods that are very abundant in certain intertidal and near-shore habitats and in the deep-sea as well. There are many species and they are divided into about 17 families, the largest number for any isopod suborder. The major portion of asellote species are small, blind, pigmentless animals found in the deep-sea, but the larger, pigmented, eyed relatives in the nearshore habitats are the most frequently encountered. One family with the largest number of recorded species, the Janiridae, has many species that are found in both the near-shore habitats and in habitats of the deep-sea as well. Several other families also have deep-sea and near-shore species in them, but species of most families are found exclusively in the deep-sea. The Asellidae are mainly freshwater species, and only Asellus sp. (Fig. 379) is included in this book for comparison. The suborder Asellota contains a large number of species, and has a great economic importance in the near-shore habitats. However, species of Asellota are abundant in the deep-sea and are perhaps the dominant crustacean found there so they are of undoubtedly of greater importance than at first is apparent. They probably account for much of the organic transformation of bottom organic material to soil or its deep-sea equivalent.

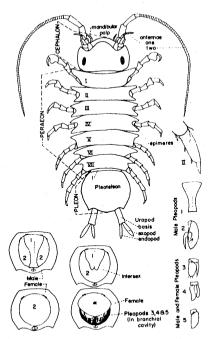


Figure 366. Asellota (General nomenclature). (Pleopods of Paraselloidea). See Fig. 373 for stenetrioidea and Fig. 378 for a selloidea.

Asellota species are characterized by the structure of their pleon. All, or all but one or two, of the pleonal segments are fused, and there is a branchial cavity which contains the respiratory pleopods, usually pleopods 3, 4 and 5. One pleopod or a combination of pleopods 1 and 2 (in males) forms as operculum to the branchial cavity. Male sexual pleopods are sometimes enclosed within the operculate covering, or they are a part of the operculum or they are sometimes outside of it. The suborder Asellota is divided into three superfamilies on the basis of the pleopodal structures. The three superfamilies are the Aselloidea, the Stenetrioidea and the Paraselloidea. Aselloidea consists of the single family, the Asellidae, which are freshwater forms

and which have the male pleopods one and female pleopods two separate; i. e., they are never fused on the midline (Fig. 378d, e). The third pleopods are operculate in both sexes.

The Stenetrioidea also consists of a single family (in North American waters), the Stenetriidae. They are perhaps the most primitive species of the suborder Asellota and have two free pleonal segments and their pleopods are arranged in various ways. Male pleopods 1 are operculate to sexual pleopods 2 and they are fused at least at the basal part of the appendages. The female operculate pleopods are large, but not fused. The forms of the pleopods found on the North American members of the family are shown in figure 373.

The Paraselloidea which contains the species of all other families found in North American waters has the first male pleopods always fused into a narrow structure that fits between the separated second pleopods which together with the first fused pair form the operculum to the branchial cavity. The second pleopods of the female are always fused and operculate to the other pleopods (Fig. 373b).

The peracon of the species many times has fused segments, and it is frequently fused to the cephalon and/or the pleon. The degree of fusion varies somewhat with age, sex (intersexes are frequently encountered) and amount of calcification of the integument. A suture is sometimes present to indicate fusion. Sometimes fusion is indicated laterally by the presence of slits, or at times it is absent. Other structures such as spines and rostral processes are also altered according to age, sex or degree of calcification so that Asellota

species must always be examined with care when encountered, since the differences found among the various forms of one species are frequently larger than those found between species in other suborders of isopods. On the other hand, the differences between isopod species in the Asellota are sometimes quite subtle and

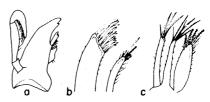


Figure 367. Mouth parts of Asellota. a. Mandible, b. Maxilla one, c. Maxilla two.

great care must be used to make sure that two very similar looking animals are not of different species. An Asellota species at first appearance appears to be quite different and indeed at times unique, but the differences soon disappear when they are compared to other asellote species and they can be at times confusing. Much work is necessary to identify species of Asellota.

Species of Asellota generally have well developed antennae, antenae two almost always being the longest. Eyes are not present in most species, but the species that will be encountered by most

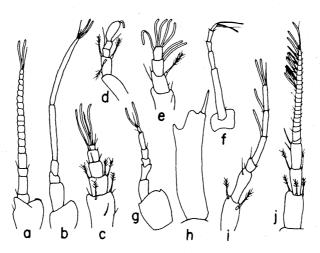


Figure 368. Antenna one types of Asellota. a. Eurycopidae (Eurycope). b. Munnidae (Munna). c. Jaeropsis. d. Nannoniscidae (Nannoniscus). e. Macrostylidae (Macrostylis). f. Haplomesus. g. lanirella. h. Echinothambema. i. Haploniscidae (Haploniscus). j. Stenetrium.

collectors do have eyes. They are usually dorsally or dorsolaterally placed on the cephalon. The mouth parts are generally of the biting and chewing types with maxillae that are fringed with long setae for straining food particles.

The maxillipeds are well developed and when more is known about them in some genera, their forms will probably become very good criteria for separation of the Paraselloidea into further subgroups and revealing the evolution of the group. The peraeopods

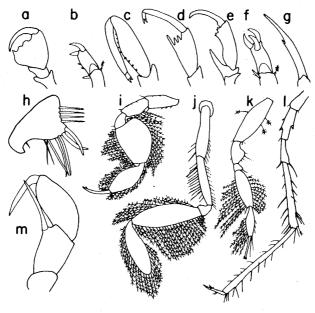


Figure 369. Peraeopods of Asellota. a. Stenetrium (I). b. Jaeropsis (I). c. Janiralata. d. Janira. e. Stenetrium (I). f. Jaeropsis (II-VII). g. Haploniscidae. h. Macrostylis. i. Ilyarachna j. Munnopsis. k. Desmosoma (II-VII). I. Munna. m. Desmosoma (I).

for the most part are ambulatory, but natatory peraeopods with broaded segments are found in some deep-sea and some partially planktonic species. Peraeopod I is generally gnathal and sometimes the male will have a large sexually modified grasping peraeopod. The number of large dactyl claws varies from one to three in the different species and on different peraeopods. Frequently if there are three dactyl claws on most peraeopods, peraeopod I will have only two. Many times the third claw is not observed, even in a compound microscope, because it is in a different plane of focus than the other two and hence out of focus when the others are observed. Knowledge of peraeopod type and dactyl claw number

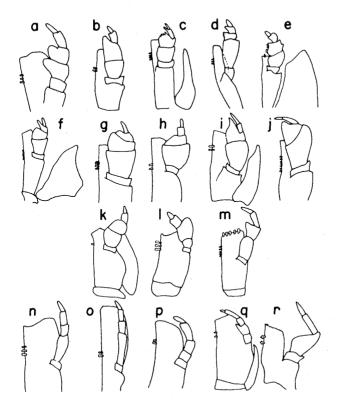


Figure 370. Maxillipeds of Asellota (for species of Janiridae, see Fig. 392).
a. Munna. b. Desmosomatidae. c. Nannoniscidae. d. Ilyarachna. e. Syneurycope.
f. Eurycope. g. Storthyngura. h. Macrostylis. i. Munnopsis. j. Acanthocope. k. Ischnomesus. l. Haplomesus. m. Stenetrium. n. Haploniscidae. o. Mesosignum.
p. Dendrotion. q. Jaeropsis. r. Abyssijaera.

also will be useful criteria for identification of Asellota, but unfortunately in some genera it has never been described.

KEY TO FAMILIES OF ASELLOTA

1a. Male pleopods 1 and 2 and female pleopod 2 much smaller than pleopods 3 which are fused and operculate to pleopods 4 and 5 (Fig. 373) Stenetriidae (p. 238)
1b. Male and female pleopods not as above 2
2a. Male pleopods 1 and 2 and female pleopods 2 forming covering operculate to pleopods 3 to 5 (Fig. 366) (Paraselloidea) 3

2b.	Male pleopods 1 and 2 and female pleopods 2 distinctly separated and branchial only (never operculate) Asellidae (p. 241)—freshule
3a.	Only five peraeonal segments visible in dorsal view; lateral borders of peraeonal segments II to V with small spines; spines also present on dorsum of cephalon, peraeon and pleotelson; cephalon and pleotelson narrower than widest part of peraeon; antenna two more than twice length of body
3b.	Not as above4
4a.	Cephalon fused with peraeonal segment I; blind; peraeonal segments IV and V elongate, twice as long as wide Ischnomesidae (p. 243)
4b.	Not as above
5a.	Blind; cephalon not fused to peraeon, but peraeonal segment VI and VII fused and also fused to pleotelson (which has indications of one pleonal segment); antenna one with very long, flattened basal segment (Fig. 368h)
	Echinothambematidae (p. 247)
5b.	Not as above 6
6a.	Body with any or all of these structures—dorsal spines, elongate lappets on peraeonal segments, cephalon or pleotelson; long coxal processes or other produced structures; elongate peraeopods (spider-like) or extremely long antennae; peraeopods I subchelate or ambulatory, peraeopods II to IV always ambulatory; peraeopods V to VII sometimes natatory; anterolateral borders of peraeonal segments sometimes acutely produced (15)
6b.	Body without any long or strongly produced spines or processes (see above couplet part 6a for types), except perhaps posterolateral projections or rostral projection; never with greatly elongate peraeopods; never with more than first three peraeonal segments acutely produced or with strongly produced coxal processes; peraeopods I prehensile or ambulatory, sometimes sexually modified or gnathal (i. e., subchelate); natatory peraeopods if present on segments II to VII, never fewer segments (7) With eyes
7a.	
	Blind (or occassionally with eyes) 8
8a.	Frontal margin rounded and serrated; antennae deeply set into cephalon Antiasidae (p. 248)
8b.	Not as above 9

9a.	Antennae one and two large and well developed; body oblong
	and dorsoventrally compressed; uropods always distinct and
	most always biramus; peraeonal segments always shorter than
	broad and of approximately uniform length; co. al plates al-
	ways visible in dorsal view Janiridae (p. 250)

9b. Not as above

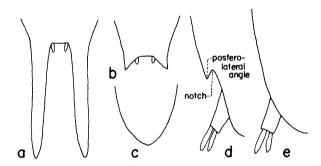


Figure 371. Posterolateral angles and notches, a-c. Haploniscidae, a. Elongate, b. Moderately long, c. Absent (Hydroniscus), d. A large notch, e. Absent

10b.	Not as above11
lla.	Uropods uniramus and very elongate, emerging from posterior border of pleotelson; posterolateral border of pleon never with posterolateral notches (Fig. 371e); mandibles without palp; dactylus of peraeopods triangulate with single unguis (claw) (Fig. 369h)
11b.	Not as above 12
12a.	Coxal extensions present on at least anterior three peraeonal segments; peraeonal segment IV almost quadrangular with subparallel lateral margins; all peraeopods except first natatory Desmosomatidae (p. 278)
12b.	Coxal extensions absent or very inconspicuous on peraeonal

segments I to III; peraeonal segment IV always constricted where attached to peraeonal segment III; all peraeopods ambulatory

Nannoniscidae (p. 281)

13a. Cephalon with rounded frontal margin; antenna one and two small and closely adherent to frontal margin (especially antenae two); lateral margins of body subparallel including cephalon and pleotelson; coxal extensions never present, but coxal bulges present on margins between peraeonal segments; posterolateral notch present on border of pleotelson; notched border of pleotelson where uropods emerge; mandibular molar process spiniform (Fig. 37e) ... Jaeropsidae (p. 284) 13b. Not as above 14a. Eyes somewhat stalked; frontal margin of cephalon concave with antennae inserted in deep grooves Antiasidae (p. 248) 14b. Eyes not stalked; frontal margin of cephalon and antennal insertions various Janiridae (p. 250) Body without spines or processes; peraeonal segments of markedly different widths: uropods small and inconspicuous or moderately long and uniramus: pleotelson always narrower than peraeon; peraeopod I gnathal and short, others long or extremely long and ambulatory or natatory; if eyes present they are stalked or laterally placed on bulges of cephalon ... 16 15b. Body usually with spines or processes or scales or if none of these present, first and sometimes other anterolateral corners of peraeonal segments produced anteriorwardly and usually those of peraeons III and IV acutely pointed; uropods conspicuous or inconspicuous and uni- or biramus; pleotelson with broadly rounded posterior border or shield-shaped, sometimes with medial posterior spine or posterior border; peraeopods various

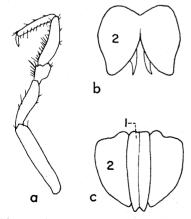


Figure 372. Peraeopod I and pleopods 2 (male) of Murinopsidae. a. Gnathal peraeopod I. b. Pleopods 2, male (fused) of eurycopids. c. Pleopod I and 2 Murinopsis.

16a. Blind; peraeon constricted after peraeonal segment IV; peraeopod I gnathal (Fig. 372a), II long and ambulatory, III and IV very long and ambulatory, V to VII natatory (and without dactyls); antennae one and two long; male pleopods two not fused at base (Fig. 372c)

.... Munnopsidae (p. 288)

100.	to peraeon, and always longer than wide; if eyes present they are laterally placed on stalks or bulges; peraeopod one short and gnathal, others elongate and ambulatory Munnidae (p. 289)
17a.	Eyes, if present, dorsally located; maxillipedal palp always wide with articles two and three always much wider than four and five; peraconal segment I usually with lateral lappets and dorsal spines (i. e., it is not markedly different from other segments); natatory peracopods never present; pleotelson generally shield-shaped
17b.	Blind; maxillipedal palp various; peraeonal segments various; natatory peraeopods sometimes present on segments V and VI and VII
18a.	Maxillipedal palp narrow for complete length (much narrower than endite) (Fig. 370n-p); peraeon I and VII simple and abruptly narrower than others; without dorsal spines or long lappets; all peraeopods ambulatory 19
18b.	Maxillipedal palp with articles two and three wider than four and five, and generally as wide as or nearly as wide as endite; peraeonal segments never abruptly narrower than body; peraeopods I to IV ambulatory, V and VI or V, VI and VII natatory 20
19a.	One lateral spine or lappet (sometimes with secondary spines on tip) on peraeonal segments II, III and IV; dorsal spines usually present Dendrotionidae (p. 299)
19b.	Two lappets on peraeonal segments II, III and IV; dorsal spines rarely present Mesosignidae (p. 298)
20a.	Antenna two very close together on medial part of frontal line; frontal line never greatly produced; body rarely with any fused segments; peraeonal segment V produced anteriorwardly; peraeonal segments VI and VII virtually never produced anteriorwardly; peraeopods I to IV ambulatory with I and II shorter than III and IV, V and VI natatory and VII ambulatory with natatory setal configuration Ilyarachnidae (p. 300)
	(p. 900)

STENETRIIDAE

The species of this family represent primitive though modified members of the Asellota. They are much like species of the freshwater family Asellidae in their general body structure, and they have two free pleonal segments. There are also differences in the structure of the pleopods, the first three pleopods being very much modified (Fig. 373). The genus Stenetrium, the only representative of the family in North American waters, is composed of many species from both shallow and deep sea habitats. Species are fre-

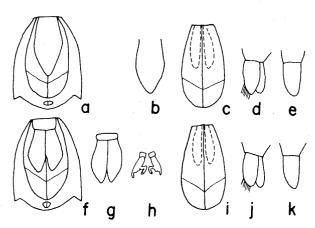


Figure 373. Ventral aspect of pleon and pleonal appendages of Stenetrium. a-e. Female. a. Ventral aspect of pleon. b. Pleopod 2. c. Operculate pleopods 3. d. Pleopod 4. e. Pleopod 5 (uniramus) f-k. Male. f. Ventral aspect of pleon. g. Pleopod 1. h. Pleopods 2. i. Pleopods 3. j. Pleopod 4. k. Pleopod 5 (uniramus).

quently taken in coral reefs in tropical parts of the world. The North American species were taken from coral reefs in Bermuda and the West Indies. Males have the first peraeopod highly modified as a grasping appendage. The North American members of the genus are among the shorter species found in the genus.

Fig. 374. Stenetrium serratum (Hansen) (6 mm)

Range: St. Thomas, West Indies (Coral reef)

This species has very large teeth on the lateral edges of the pleotelson.

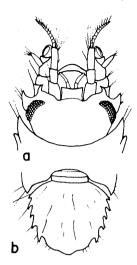


Figure 374. a. Stenetrium serratum, anterior part. b. Posterior.

Fig. 375. Stenetrium occidentale (Hansen) (3.3 mm)

Range: St. Thomas, West Indies (Coral reef)

This species has very large posterolateral angles on the pleotelson and the posterior margin is produced into a broad lobe.

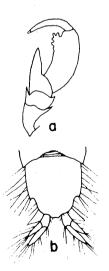


Figure 375. a. Stenetrium occidentale, peraeopod I (male). b. Posterior part.

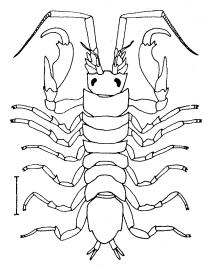


Figure 376. Stenetrium antillense.

Fig. 376. Stenetrium antillense (Hansen) (4.5 mm)

Range: West Indies (Shallow water)

This species has only two teeth on the produced inner distal angle of the propodus of male peraeopod I.



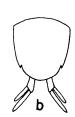


Figure 377. a. Stenetrium stebbingi, peraeopod l (male). b. Posterior part.

Fig. 377. Stenetrium stebbingi (Richardson)

Range: Bermuda (1/2 to 4 m)

This species, much like *S. occidentale* in appearance, has small posterolateral angles and a broadly rounded posterior margin on the pleotelson.

ASELLIDAE

Species of the family are found in most still and permanent bodies of freshwater in southern Canada and the northeastern part of the United States. There are many additional species and subspecies from caves and other underground water sources. Some species have been living in the underground dark habitats long enough to have become blind. The loss of eyes in total darkness is similar to that of species in the deep-sea.

Several species of asellids are commonly found widespread in North America. Specimens never have been critically examined and compared and they have not been consistently named. The genus with no named species is included here because there is no agreement as to what is the name of the most common North American species, or even if there is one "most common" North American species. Most freshwater isopods are in the family Asellidae, but species of the other families of other suborders get into freshwater. The freshwater species have different ancestors than terrestrial species. The asellid species are most closely related to the species of Stenetridae.

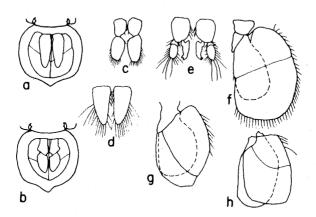


Figure 378. Pleopods of Aselloidea..a. Ventral aspect of pleon of female. b. Ventral aspect of pleon of male. c. Pleopod 1, male. d. Pleopod 2, female (There is no pleopod 1 in the female). e. Pleopod 2, male. f-h. Male and female pleopods. f. 3. g. 4. h. 5 (biramus).

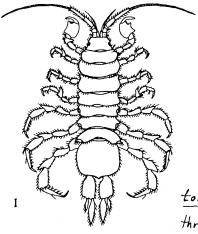


Fig. 379. Asellus sp. (10 to 30 mm)

Range: Much of southern Canada and the United States (Freshwater)

Probably several species. Asellus tomalensis Harford 1877 is common throughout No. Calif.

Figure 379. Asellus sp.

HAPLOMUNNA CAECA INCERTAE SEDIS

This asellote has never convincingly been placed in a category higher than genus. It appears to be a munnid, but there are some differences which make it impossible to assign it to that family. The species was not well described.

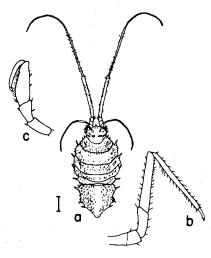


Figure 380. a. *Haplomunna* caeca. b. Peraeopod II. c. Peraeopod I.

Fig. 380. Haplomunna caeca (Richardson) (8 mm)

Range: Off Santa Catalina Island, California (3990 m)

This species is only indirectly related to the species of *Munna* that are described in this book. It is not clear whether it would be included in the family Munnidae or not. The species is blind and the type and only one in the genus.

ISCHNOMESIDAE

The species of this family are interesting to see. When viewed alone or drawn without appendages they look like a turned piece of wood-like the rung of the back of an old rocking chair. When grouped together in a bowl awaiting study they look like a bunch of twigs. They are the walking sticks among the isopods. The segments of the elongate body of the ischnomesids are fused in various ways and the appendages are thought to be long, but for most species they have never been seen. The body of the species is generally highly calcareous and they can become very brittle if not preserved properly. The general body configuration, the presence or absence of free or fused segments and the type of uropods are used as criteria to distinguish the different genera. There are eight known genera three of which have been recorded in North American waters. The genus Ischnomesus, the largest of the family, contains about 20 species. Heteromesus and Haplomesus contain 8 and 11 species respectively, but only one species of Haplomesus and three species of *Heteromesus* have been recorded from North American waters.

KEY TO THE GENERA OF ISCHNOMESIDAE

- 1a. Uropod with basis only; pleon with segments completely fused and various numbers of peraeonal segments also fused and many times fused with pleon 2
- 1b. Uropods with one ramus (uniramus); pleon with one free segment and with little or no indication of other segments

Ischnomesus

Fig. 381. Ischnomesus profundus (Hansen) (4 mm)

Range: North Atlantic, southern part of

Davis Strait (3521 m)

No long anterolateral projections are present on the peraeonal segments of this species.

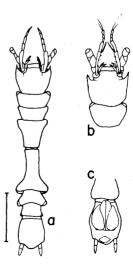


Figure 381. a. Ischnomesus profundus. b. Anterior part. c. Posterior part.

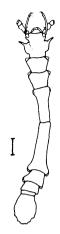


Figure 382. Ischnomesus caribbicus.

Fig. 382. Ischnomesus caribbicus (Menzies) 7.8 mm)

Range: Caribbean Sea (1714 m)

Only peraeonal segment I which is fused to the cephalon has the anterolateral corners produced.



Figure 383 (a) Ischnomesus armatus.

Fig. 383(a). Ischnomesus aramtus (Hansen) (4.8 mm)

Range: Davis Strait to north of Puerto Rico (2702 to 5494 m)

Peraeonal segments I to VI are armed with large spines.

Fig. 383(b). Ischnomeus multispinis (Menzies) (7.4 mm)

Range: Caribbean Sea (975 m)

Many large spines are present on this species.

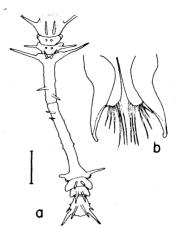


Figure 383(b) a. 'schnomesus multispinis. b. Apex pleopod 1, male.

Fig. 384. Heteromesus spinescens (Richardson)

Range: Georges Bank to Virginia (2155 to 3337 m)

The anterior part of the lateral margins of the pleon are bulged in this species.



Figure 384. Heteromesus spinescens.

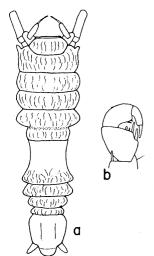


Figure 385. a. Heteromesus grandulatus. b. Peraeopod I.

Fig. 385. Heteromesus granulatus (Richardson)

Range: South of Martha's Vineyard to Block Island, New York (713 to 3235 m)

The lateral borders of the pleon are regularly convex.



Figure 386. Heteromesus bifurcatus.

Fig. 386. Heteromesus bifurcatus (Menzies) (3.5 mm)

Range: Caribbean Sea, north of Columbia (4071 m)

This species, known only from the posterior part of the animal, has furrows representing pleonal segments.

2b. Peraeonal segments V to VII and pleon fused; antenna one with flagellum not rudimentary; i. e., normal Haplomesus

Fig. 387. Haplomesus tropicalis (Menzies) (4.1 mm)

Range: Caribbean Sea, north of Columbia (4071 m)

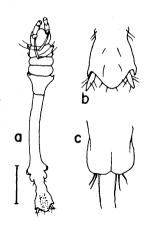


Figure 387. a. Haplomesus tropicalis. b. Detail, posterior part. c. Detail, apex male pleopod 1.

Fig. 388. Haplomesus quadrispinosus (Sars) (4.5 mm)

Range: Davis Strait (3420 m)

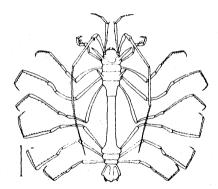


Figure 388. Haplomesus quadrispinosus.

ECHINOTHAMBEMATIDAE

This is a good example of a single specimen that is so different from other species that it forms the basis of, not just a new species or genus, but a new family as well. The differences that distinguish the specimen from others are logical differences and are of such a nature as to be considered biologically normal; hence, the specimen is not considered a freak or injured specimen or a specimen that has grown up distorted. There are two spines on peraeonal segment II which probably indicates two lateral lappets. Peraeonal segment VII and peonal segment I each have a single spine and are fused into one large body section. Antenna one has a large flattened basal segment and antenna two is normal.

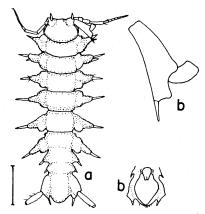


Figure 389. a. Echinothambema ophiuroides. b. Antenna one. c. Ventral aspect of pleon, intersex.

Fig. 389. Echinothambema ophiuroides (Menzies) (5 mm)

Range: North of Puerto Rico (5104 to 5122 m)

The specimen on which the species is based in an intersex; i. e., it is not a male or female, but rather has some of the external structures of both.

ANTIASIDAE

There are about 12 species in three genera in the family. They are small species and in some ways resemble species of Munnidae. Occasionally species of the family have eyes, but most are blind. There are never more than three articles in the flagellum of antenna one. Peraeopod I is prehensile; the remaining ones are ambulatory with two dactyl claws. Biramus uropods are found and they are inserted dorsally or laterally on the pleotelson. Two genera each with one species are known from North American waters.

KEY TO GENERA OF ANTIASIDAE

 Fig. 390. Abyssianira dentifrons (Menzies) (2.8 mm)

Range: North of Puerto Rico (5104 to 5122 m)

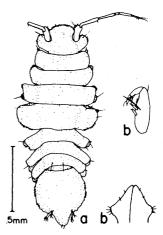


Figure 390. a. Abyssianira dentifrons. b. Peraeopod 1. c. Detail, apex male pleopod 1.

Fig. 391. Antias hirsutus (Menzies) (1.5 mm)

Range: Tomales Bluff, California (Intertidal)

All five articles on the maxillipedal palp are narrow on this species.

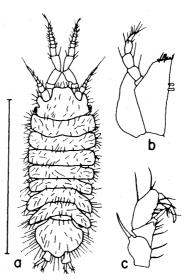


Figure 391. a. Antias hirsutus. b. Maxilliped. c. Peraeopod I.

JANIRIDAE

This family is one of the most diverse asellote families and contains common well defined species as well as poorly defined ones. The species have very little in common and yet there are not really well defined differences that separate them; hence, much difficulty is encountered when it comes to putting the species into taxonomic categories within the family. There have been many published attempts to organize the family, either by removing the troublesome species to other facilities or by making a very broad definition of the family to include all of its diverse elements. Neither method has been too successful. Here the genera and species are put together in one broadly defined group which is called the family Janiridae.

Both blind and species with eyes are included and they are from very diverse ecological and geographical regions. There are species from near-shore and species from the deep-sea. The body of the species is usually dorsoventrally compressed. The cephalon, peraeonal segments and anterior one or two pleonal segments are usually free and not fused. Generally there are two free pleonal segments, but they are generally too small and indistinct to be useful as key characters. The pleon, except for the anterior segments, is generally shield shaped. The mouth parts of the animals are normal; i.e., there are no criteria that can be used to set them apart from those of other species. The first peraeopod is for most species gnathal or prehensile, and the next six are ambulatory. There are two or sometimes three claws on the dactyls (Figs. 29d and 369f). The uropods are usually biramus, but there are some species on which they are uniramus. There are about 35 genera in the family and 13 of them have representatives in North American waters.

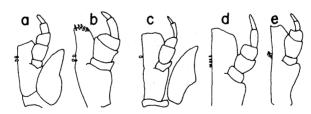


Figure 392. Maxillipeds of some janirid genera, a. lanirella, b. laniropsis, c. Janira, d. Acanthomunna, e. Rhacura.

KEY TO GENERA AND SPECIES OF JANIRIDAE

la.	With large spines (lateral lappets and/or dorsal projection	ıs)3
1b.	Body smooth (lateral processes if present short)	2
29	Fues present	11

Fig. 393. Ioella speciosa (Bovallius) (21.5 mm)

Range: Baffin Bay, Canada

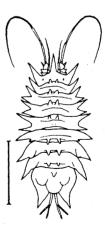


Figure 393. loella speciosa.

5b. Pleotelson relatively smooth I. spinosa

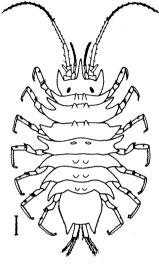


Fig. 394. Ioella spinosa (Harger) (8 mm)

Range: Davis Strait (146 to 183 m)

Figure 394. Ioella spinosa.



Figure 395. Ioella libbeyi.

Fig. 395. *Ioella libbeyi* (Ortmann) (8 mm) Range: Cape Alexander, North Greenland

Fig. 396. Ioella glabra (Richardson)

Range: Off Cape Hatteras, North Carolina (1624 m)



Figure 396, loella alabra.

Fig. 397. Rhacura pulchra (Richardson)

Range: Georges Bank (3219 m)

Minute eyes are present on this type and only species of the genus. The first pair of peraeopods are prehensile and the other pairs are ambulatory.

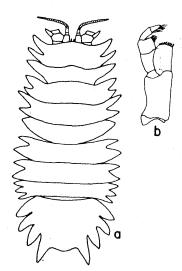


Figure 397. a. Rhacura pulchra. b. Maxilliped.

7b. Lateral border of cephalon with only one prominent spine; lappets on peraeonal segments arranged according to formula -1, 2, 2, 2, 1, 1, 1

8a. Dorsal spines and strongly produced lateral spines on peraeon and pleon; pleotelson secondarily produced posteriorward with two spines on posterior margin; uropods biramus

Microproctus

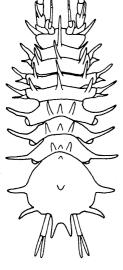


Fig. 398. *Microproctus caecus* (Richardson) (12 mm)

Range: Bering Sea, near Agattu, Aleutian Islands (549 m)

This blind, pigmented species is the type and only species in the genus. The species description is based on one male specimen.

Figure 398. Microproctus caecus.

8b. No dorsal spines, only lateral lappets; pleotelson much narrower than peraeonal segment II; uropods uniramus

Ianirella

GENUS IANIRELLA

There are eleven species in the genus. All are from the Atlantic Ocean, but only three are from North American waters.

Fig. 399. Ianirella lobata (Richardson)

Range: Southeast of Georges Bank (2480 to 3225 m)

This species has parallel rows of mediodorsal spines and lateral lappets that are rounded apically and apparently without spines.

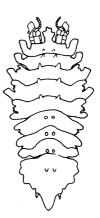


Figure 399. Ianirella lobata.

Fig. 400. Ianirella vemae (Menzies) (3.3 mm)

Range: North of Puerto Rico (5104 to 5122 m)

No dorsal spines are present on this species. It might perhaps be a juvenile form of the species *I. caribbica*.

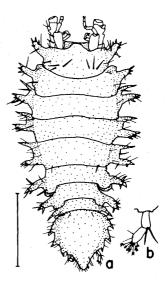


Figure 400. a. lanirella vemae. b. Uropod.

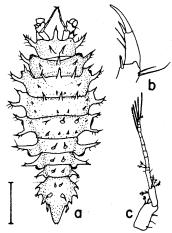


Figure 401. a. Ianirella caribbica. Peraeopod I. c. Antenna one.

Fig. 401. Ianirella caribbica (Menzies) (5.5 mm)

Range: Caribbean Sea, south of Jamaica (1244 m)

The juvenile of this species perhaps is the species I. vemae which has no spines. More must be learned to discover if dorsal spines are an adult character.

9a. Body with subparallel lateral borders 10

9b. Body with peraeonal segment IV widest Abyssijaera

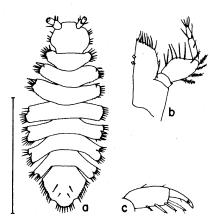


Figure 402. a. Abyssijaera acarina, b. Maxilliped, c. Peraeopod I.

Fig. 402. Abyssijaera acarina (Menzies) (1.6 mm)

Range: South of Bermuda (5779 m)

This species forms the type of a new genus of which there are now several species known.

10a. Body elongate; antenna one with first basal segment elongate, but not wide; uropod elongate with endopod much longer than exopod Caecianiropsis Fig. 403. Caecianiropsis psammophila (Menzies and Pettit) (2.2 mm)

Range: Mid-California (Intertidal)

This blind species lives between the sand grains on the intertidal beaches. They live several centimeters below the surface and can be quite abundant. It shares the sand beach habitat with species of *Microcerberus* (p. 216).

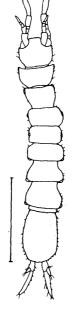


Figure 403, Caecianiropsis psammophila.

Fig. 404. Caecijaera horvathi (Menzies) (1.7 mm)

Range: Los Angeles Harbor, California (Intertidal)

This very small, blind species is commensal with the boring isopod Limnoria. Limnoria specimens, which are about twice as large as C. horvathi specimens, burrow into wood and the burrows also serve as the home of C. horvathi. Limnoria species have been found without the C. horvathi species, but C. horvathi have never been found without their Limnoria host to make the holes.

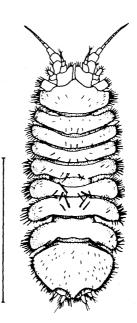


Figure 404. Caecijaera horvathi.

lla.	Lateral margins of peraeon subparallel 12
11b.	Lateral margins of peraeon convex, or at least some of peraeonal segments noticably larger than others 24
12a.	Frontal margin of cephalon without medial projection; pleon variously attached to peraeon; anterolateral angles of peraeonal segments II, III and IV produced only slightly or not at all
12b.	Frontal margin of cephalon with rostral projection; pleon narrowly attached to peraeon; anterolateral angles of peraeons II, III and IV produced

GENUS JANIRA

The species in this genus, although common, are not known as well as they should be because most of the descriptions of the species are old and too general for critical taxonomic distinctions. The species listed here in the genus most probably represent a heterogeneous assemblage of genera.

KEY TO SPECIES OF JANIRA

- 13a. Pleotelson with posterolateral angles or crenulations on posterior margin

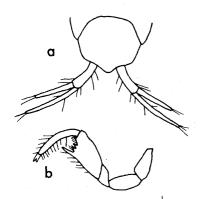


Figure 405. a. Janira minuta, poserior part. b. Peraeopod 1, male.

Fig. 405. Janira minuta (Richardson)

Range: Castle Harbor, Bermuda (In dead coral)

14a. Posterolateral angles present and widely separate

Fig. 406. Janira tricornis (Kroyer) Range: Greenland (9 to 91 m)

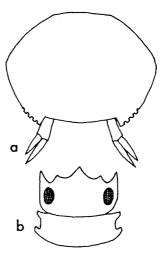


Figure 406. a. Janira tricornis, posterior part. b. Anterior part.

14b. Crenulations only present in posterolateral margins _____15

Fig. 407. Janira maculosa (Leach) (10 mm)

Range: Davis Strait (55 to 212 m)

This species has been recorded from the west coast of North America, but the report was probably in error or the specimen was really *J. occidentalis* which has now been placed in *Jani*ralata (p. 265)

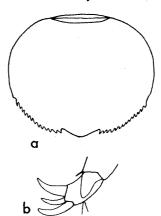


Figure 407, a. Janira maculosa, posterior part. b. Peraeopod V.

15b. Length of one posterolateral crenulation equal to or shorter than medial non-crenulated portion of posterior margin of pleotelson

1. alta

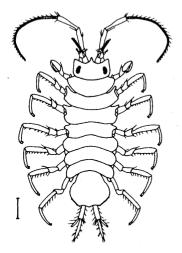


Fig. 408. Janira alta (Stimpson) 7 mm)

Range: Nova Scotia to Chesapeake Bay (64 to 891 m)

Figure 408. Janira alta.

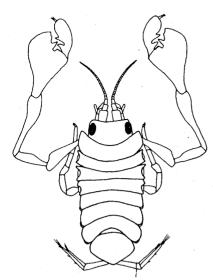


Figure 409. Carpias bermudensis.

Fig. 409. Carpias bermudensis (Richardson) (2.5 mm)

Range: Bermuda (Shallow water)

In this species the male has a very large subchelate peraeopod I. The female peraeopod I resembles that of the other peraeopods.

GENUS IANIROPSIS

The species of the genus are very common on the sea coasts of the world. There are three claws on the dactyls of peraeopods II to VII; only two are found on peraeopod I. The frontal margin of the cephalon is nearly straight, or only slightly produced, with rounded anterolateral corners in most species.

17a.	Lateral borders of pleotelson with small teeth	18
17b.	Lateral borders of pleotelson smooth	20
18a.	Lateral borders of pleotelson with more than four of frontal margin of cephalon rounded with small anterology projections I. an	ateral

Fig. 410. Ianiropsis analoga (Menzies) (3.7 mm)

Range: Northern Washington to Mid-California (Intertidal)

The specimens were taken from under rocks and from under the holdfasts of algae.

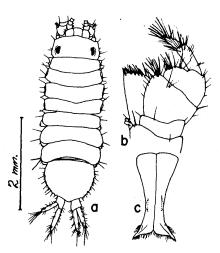


Figure 410. a. laniropsis analoga. b. Maxilliped. c. Pleopod 1, male.

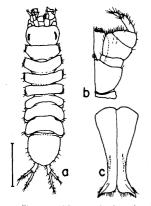


Figure 411. a. laniropsis epilittoralis. b. Maxilliped. c. Pleopod 1, male.

Fig. 411. *Ianiropsis epilittoralis* (Menzies) (3.8 mm)

Range: Mid-California (Intertidal)
This species was found in tidal pools.
Dillon Beach to San Luis Chispo.

19b. Peraeonal segment V with nearly square anterolateral corners; pleotelson shorter than wide; uropods shorter than pleotelson

I. tridens

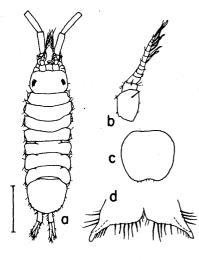


Fig. 412-413. Ianiropsis tridens (Menzies) (2.5 mm)

Range: Puget Sound, Washington to northern Chile (Intertidal)

The species is common on algae and is occasionally found in sponges.

Figure 412. a. laniropsis tridens. b. Antenna one. c. Pleopod 2, female. c. Detail, pleopod 1 male.



Figure 413. laniropsis tridens, lateral view.

straight or

(Menzies) (2.2 mm)

to 57 m)

20a. Frontal margin of cephalon with anterior margin not produced, but trisinuate; no anterolateral corners present 21

20b. Frontal margin of cephalon acutely produced; small antero-

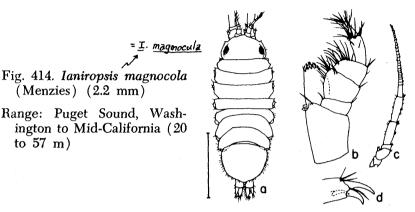


Figure 414. a. laniropsis magnocula. b. Maxilliped. c. Antenna two. d. Peraeopods II to VII.

21a. Uropods shorter than pleotelson 21b. Uropods elongate, much longer than pleotelson

I. montereyensis

Fig. 415. Ianiropsis montereyensis (Menzies) (3.6 mm)

Range: Mid-California (Intertidal)

The species was found under rocks on a rocky beach below high tide level.

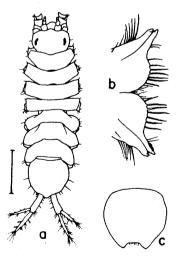


Figure 415. a. laniropsis monter-eyensis. b. Detail, pleopod 1, male. c. Pleopod 2, female.

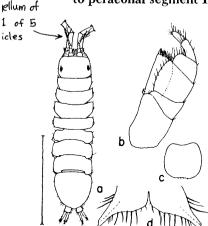


Figure 416. a. *laniropsis minuta*. b. Maxilliped. c. Pleopod 2, female.

d. Detail pleopod 1 male.

Fig. 416. *Ianiropsis minuta* (Menzies) (1.3 mm)

Range: Mid-California (Intertidal)

The species was collected among rocks and in sand.

22b. Body with lateral margins with cephalon and pleon narrower than peraeon and lateral parts of peraeonal segments separated at ends; cephalon much wider than long and narrowly attached to peraeonal segment I. kincaidi

Fig. 417. Ianiropsis kincaidi kincaidi (Richardson) (3.8 mm)

Range: Bering Sea to Mid-California (Surface to 69 m)

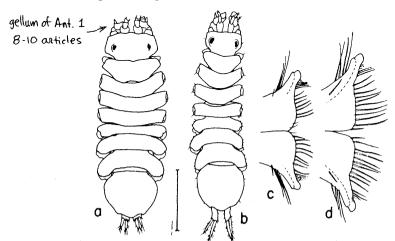


Figure 417. a. laniropsis kincaidi kincaidi. b. l. k. derjugini. c. Detail pleopod 1, male (l. k. derjugini). d. Detail pleopod 1, male (l. k. kincaidi).

The subspecies *I. kincaidi derjugini* Gurjanova (2.5 m) has been described within the range of the parent species. It (Fig. 417b) is narrow with elongate uropods when compared to the parent species (Fig. 417a).

- 23a. Cephalon with no medial rostral spine or process, but with acutely produced anterolateral projections; uropods either short or long

GENUS JANIRALATA

The species of this genus ought to be compared more closely with those of the genus *Janira*. They are very similar in many features. Most of the species of the genus are found in shallow, near shore habitats.

KEY TO SPECIES OF JANIRALATA

- 24a. Three lobes (posterolateral corners and medial lobe) or five lobes present and prominent on frontal margin of cephalon

Fig. 418. Janiralata rajata (Menzies) (4 mm)

Range: Monterey Bay, California (37 m)

The specimens were taken from the egg cases of ray fish.

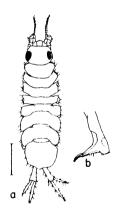


Figure 418. a. Janiralata rajata. b. 1/2 of pleopod 1, male.

on frontal margin of cedhalon

25b. Three lobes only present; medial and anterolateral margins of cephalon only slightly produced _______26

26a. Cephalon greatly wider than widest part of peraeon

1. occidentalis

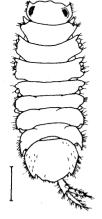


Fig. 419. Janiralata occidentalis (Walker) (5.5 mm)

Range: Puget Sound, Washington to Laguna Beach, California (18 to 69 m)

Figure 419, Janiralata occidentalis,

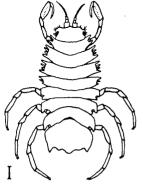


Fig. 420. Janiralata sarsi (Richardson) (10.5 mm)

Range: Amchitka Islands, Alaska (15 m)

Figure 420. Janiralata sarsi.

27a. Posterolateral corners of pleotelson pointed 28

27b. Posterolateral corners of pleotelson rounded (the three lobes on the posterior margin are about the same size.)
28a. Anterolateral corner and one lateral lappet or one bifurcate lateral lappet on cephalon
30
28b. Single anterolateral corner, no lateral lappets on cephalon

Fig. 421. Janiralata solasteri (Hatch) (5.7 mm)
Range: Alaska to southern California (Shal-

low water)

I'm seen speamens from Gulf of Alusica soft-bottom subtidal.

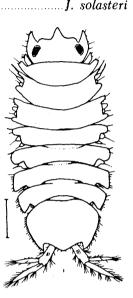


Figure 421. Janiralata solasteri.

29a. Medial margin of cephalon greatly produced J. triangulata

Fig. 422. Janiralate triangulata (Richardson) (3.8 mm)

Range: Monterey Bay, California (Shallow water)

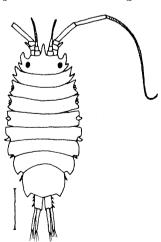


Figure 422, Janiralata triangulata.

29b. Medial margin of cephalon not greatly produced ... J. holmesi

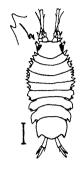


Fig. 423. Janiralata holmesi (Richardson) (5.6 mm)

Range: Stephens Passage and southeastern Alaska (80 to 344 m)

Figure 423. Janiralata holmsei.

30a. Medial margin of cephalon greatly produced J. alascensis



Figure 424. Janiralata alascensis.

Fig. 424. Janiralata alascensis (Richardson) Range: Just south of Alaskan Peninsula (35 m)

30b. Medial margin of cephalon not greatly produced . J. erostrata

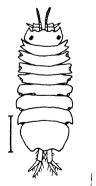


Figure 425. Janiralata erostrata.

Fig. 425. Janiralata erostrata (Richardson) (3.5 mm)

Range: Attu, Aleutian Islands (Subtidal)

Fig. 426. Jaera marina (Fabricus) (3 mm)

Range: Laborador to New York (Intertidal)

This is one of the most frequently encountered species in the intertidal and slightly below the low tide waters of northeastern North America. It is frequently found in tide pools and other shallow water retreats. Occasionally the species is found in brackish water.

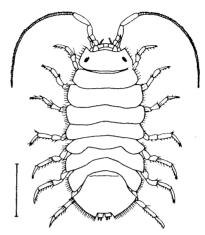


Figure 426. Jaera marina.

No. Fig. Jaera wakishiana (Spence Bate) Range: Esquimault Harbor, British Columbia (Intertidal)

The species is named after the tribe of Indians on whose land it was collected. No figure or really good description of the species is to be found.

Fig. 427. Iais californica (Richardson) (1.9 mm)

Hompoidt Bay, to Nowport Bay

Range: Sausalito, A California (Shallow water)

The species is found almost exclusively among the pleopods of the species *Sphaeroma pentodon* (p. 129), an isopod that burrows into soft rock.

Also reported from Australia, New Zealand & Singapore.

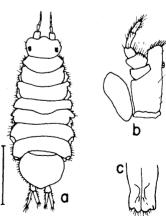


Figure 427. a. lais californica. b. Maxilliped. c. Detail, pleopod 1, male

HAPLONISCIDAE

"Haplo-" in Greek means plain or without ornamentation and when one first encounters a member of the family Haploniscidae the name seems appropriate. After much study of many specimens, however, certain consistent differences are discovered, and it is soon realized that the animals are not plain at all, but rather are part of a group with a very complex, but very subtle morphological diversity.

The species are almost all from the deep-sea where they are apparently somewhat like terrestrial isopods in their general biology. They are very small and blind and crawl about on short legs on the bottom where they are most probably omnivorous detritus feeders. Their exoskeletons are highly calcified in most species. In addition to male-female differences there are also intersexes with their particular differences. They have a morphology that is intermediate between that of males and females in some ways and it is also like that of juveniles. Confusion then in placing specimens in the correct genus and species can result.

The species of the genus are one of the most frequently encountered groups found in the deep-sea and four genera and eight species are included here. The species of the family are so numerous and so geographically widespread that when more deep-sea regions are explored many more genera and species will be discovered.

A KEY TO THE GENERA OF HAPLONISCIDAE

1a. Uropods shaped like ten-pins (Fig. 428b) Apsidoniscus

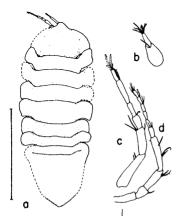


Figure 428. a. Apsidoniscus perplexus. b. Uropod. c. Antenna one. d. Antenna two.

Fig. 428. Apsidoniscus perplexus (Menzies and Schultz) (2.1 mm)

Range: Caribbean Sea, north of Columbia (3071 m)

This is the type and only species in the genus.

1b. Uropods, if present, with more or less parallel edges 2

2a. Antenna two with fifth and sixth peduncular segments fused (Fig. 35e); subrostral plate quite large

Antennuloniscus

Fig. 429. Antennuloniscus dimeroceras (Barnard) (2.2 mm)

Range: North of Puerto Rico (1816 m)

The species is the type-species of the genus.

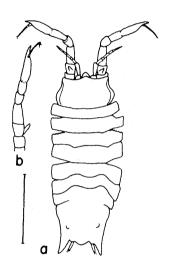


Figure 429. a. Antennuloniscus dimeroceras. b. Antenna two

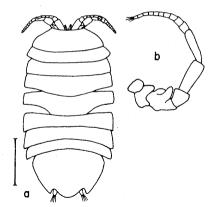


Figure 430. a. Haploniscus excisus, b. Antenna two.

Fig. 430. Haploniscus excisus (Richardson) (3.5 mm)

Range: Georges Banks and North Atlantic (3235 m) This species is the type-species of the genus.

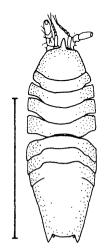


Figure 431. Hap-Ioniscus unicornis.

Fig. 431. Haploniscus unicornis (Menzies) (1.5 mm)

Range: North of Puerto Rico (5104 to 5122 m)

Fig. 432. Haploniscus tropicalis (Menzies) (1.5 mm)

Range: Caribbean Sea (1714 m)

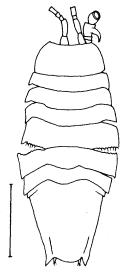


Figure 432. Haploniscus tropicalis.

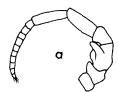


Fig. 433. Haploniscus retrospinis (Richardson)

Range: South of Martha's Vineyard (713 m)

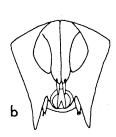


Figure 433. a. Haploniscus retrospinis, antenna two. b. Posterior part, ventral view.

3b. Antenna two and antenna one with basal segments of peduncle not easily distinguishable from those of flagellum (a spine is never present on any segment of peduncle of antenna two); last three segments of peraeon fused and abruptly wider than

first four peraeonal segments (frequently it is flared forward into two acute anterolateral angles); posterolateral angles of pleotelson weakly developed if at all, and never visible in dorsal view; frontal margin of cephalon always greatly produced

Hydroniscus

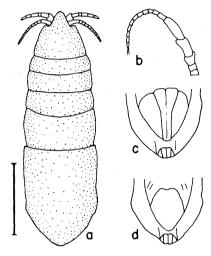


Figure 434. a. *Hydroniscus abyssi*. b. Antenna two. c. Posterior part, male (ventral view). c. Posterior part, female (ventral view).

Fig. 434. Hydroniscus abyssi (Hansen) (2.8 mm)

Range: North Atlantic, Davis Strait (3521 m)

This species is the typespecies of the genus.

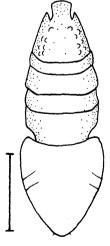


Figure 435. Hydroniscus quadrifrons.

Fig. 435. Hydroniscus quadrifrons (Menzies) (3.2 mm)

Range: North of Puerto Rico (5163 to 5684 m)

MACROSTYLIDAE

The family contains species in a single genus *Macrostylis* that are very common in the deep-sea. They are small and have long uniramus uropods. All the peraeopods are short and ambulatory. The species are blind and pigmentless and have short antennae. In the more northern part of their range the species are collected in moderately deep water. A species of the genus, *M. galatheae*, was collected from the Pacific Ocean near the Philippine Islands from a depth of six miles. The depth exceeded that of any other isopod and is the depth record for all crustacean species.

KEY TO THE SPECIES OF MACROSTYLIS

1a. Posterior margin of pleotelson broadly rounded, produced and rounded or obtusely pointed 2

Fig. 436. Macrostylis truncatex (Menzies) (3.5 mm)

Range: Off northern Florida (3950 to 3963 m)

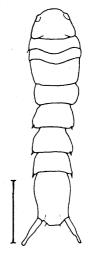


Figure 436. Macrostylis truncatex,

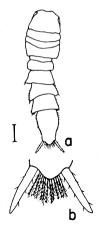


Fig. 437. Macrostylis setifer (Menzies) (6.1 mm) Range: North of Puerto Rico (5477 to 5484 m)

Figure 437. a. Macrostylis setifera. b. Detail, posterior border pleotelson.

- 2b. Peraeonal segment IV slightly narrower and as long or longer than some of other segments 3

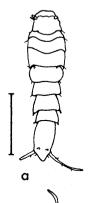


Fig. 438. Macrostylis minutus (Menzies) (2.5 mm)

Range: North of Puerto Rico (5163 to 5944 m)

Figure 438. a. Macrostylis minuta. b. Antenna one.

3b. Frontal margin of cephalon straight or broadly rounded; posterior margin of pleotelson broadly rounded or produced and broadly rounded

4a. Lateral borders of pleotelson parallel for entire length

Fig. 439. Macrostylis abyssicola (Hansen) (3.1 mm)

Range: Davis Strait (3229 m)

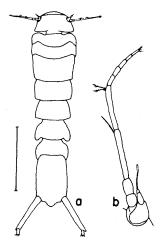


Figure 439. a. Macrostylis abysicola, b. Antenna two and one

- 4b. Lateral borders of pleotelson indented in posterior 1/3 of length 5

Fig. 440. Macrostylis vemae (Menzies) (3.9 mm)

Range: North of Puerto Rico (5440 to 5410 m)

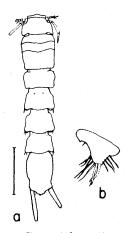


Figure 440. a. Macrostylis vemae. b. Peraeopod III.

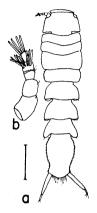


Figure 441. a. Macrostylis caribbicus. b. Antenna one.

Fig. 441. Macrostylis caribbicus (Menzies) (4.8 mm)

Range: Caribbean Sea, north of Columbia (2875 to 2941 m)

DESMOSOMATIDAE

The species of this family have six natatory peraeopods only peraeopod I being ambulatory. They are occasionally caught in deep plankton tows. Only the genus *Desmosoma* has been collected from North American waters. There are about 27 species widely distributed in the deep seas of the world.

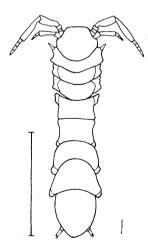
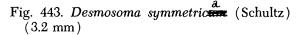


Figure 442. Desmosma plebeium.

Fig. 442. Desmosoma plebejum (Hansen) (2 mm)

Range: North Atlantic and Arctic Ocean (1040 to 1613 m)

This deep water species was first taken in the North Atlantic off Iceland and then was taken from the water below an ice island station (Station Bravo) in the Arctic Sea northeast of Alaska. The species differs from many members of the genus in that peraeonal somites IV and V are narrower than the other peraeonal segments.



Range: Tanner Canyon on shelf off southern California (469 m)

The body of this species gradually narrows caudaly.

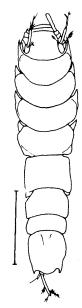


Figure mosoma cum.

ure 443. Desna symmetri-

Fig. 444(a). Desmosoma birsteini (Menzies) (2.3 mm)

Range: Near Bermuda (5166 m)

The posterior half of the body of this species is abruptly narrower than the anterior half.

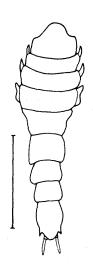


Figure 444(a). Desmosoma birsteini.

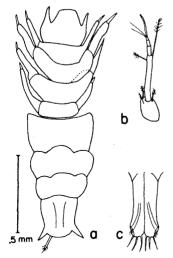


Figure 444(b). a. Desmosoma magnispinium. b. Antenna one. c. Apex pleopod 1, male.

Fig. 444(b). Desmosoma magnispinum (Menzies) (2.1 mm)

Range: Caribbean Sea near Panama (1906 m)

The lateral lappets on the anterior peraeonal segments are very long.

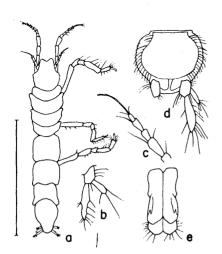


Figure 444(c). a. Desmosoma lobiceps.

Fig. 444(c). Desmosoma lobiceps (Blake) (2 mm)

Range: Mount Desert Island, Maine (Intertidal)

Fig. 445. Desmosoma tenuimanum (Sars) (4 mm)

Range: East Greenland (22 m)

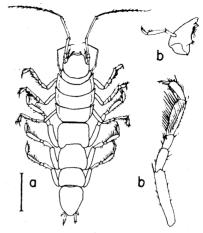


Figure 445. a. Desmosoma tenuimanum. b. Mandible. c. Peraeopod 11.

Fig. 446. Desmosoma armatum (Sars) (2 mm)

Range: East Greenland (350 m)

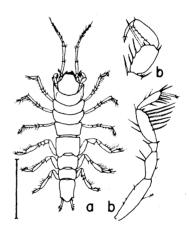


Figure 446. a. Desmosoma armatum. b. Peraeopod I. c. Peraeopod V

NANNONISCIDAE

The species of the family have no peraeopods modified for swimming, so they probably are weak swimmers that live on the bottom. The molar process of the mandible in the animals is reduced to a short setiferous lobe (Fig. 37g). Members of two genera are found in North American waters. They have a bulbous organ on the flagellum of antenna one (Fig. 448b). It is apparently a modified aesthetase which is a special chemical receptor, but its true func-

tion remains unknown. About 17 species are in *Nannoniscus* the nominal genus, the largest one in the family. Four species have been collected in North American waters.

KEY TO THE GENERA OF NANNONISCIDAE

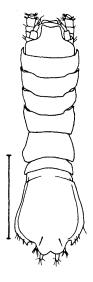


Fig. 447. Nannonisconus latipleonus (Schultz) (2.8 mm)

Range: Redondo Canyon on shelf off southern California (465 m)

Figure 447, Nannonisconus latipleonus,

1b. Pleotelson not wider than peraeon; anterolateral lobes present on cephalon Nannoniscus

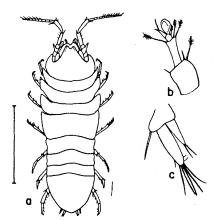


Fig. 448. Nannoniscus oblongus (Sars) (2.2 mm)

Range: Davis Strait and west of Iceland (225 to 248 m)

Compared to other species of the genus *N. oblongus* has a very squat body and it is without posterolateral angles on the pleotelson.

Figure 448. a. Nannoniscus oblongus. b. Antenna one. c. Uropod. Fig. 449(a,b). Nannoniscus camayae (Menzies) (4.1 mm)

Range: Caribbean Sea, near Cayman Islands (1714 m)

This species is fairly elongate with the anterior part of the body wider than the posterior part. Simple posterolateral angles are present on the pleotelson.

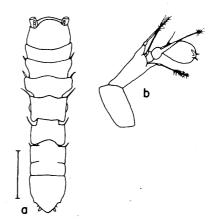


Figure 499(a, b). a. Nannoniscus camayae, b. Antenna one.

Fig. 449(c,d). Nannoniscus primitivus (Menzies (1.8 mm)

Range: Off Cartagena, Columbia (2868 to 2875 m)

This species has a peculiar bulbous organ. There are two posterolateral corners on the pleotelson.

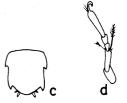


Figure 449(c, d). c. N. primitivus, posterior part. d. Antenna one.

Fig. 450. Nannoniscus arcticus (Hansen) (2.8 mm)

Range: Just south of Greenland (75 m)

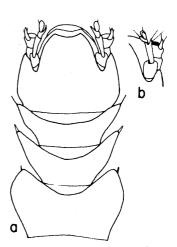


Figure 450. a. Nannoniscus arcticus, anterior part. b. Antenna one.

JAEROPSIDAE

The family contains only one genus Jaeropsis in which there are about 20 species. They are small, have eyes and are rarely over 5 mm in length. They differ from other asellotes in the presence of very short antennae one and two and in the spiniform molar process on the mandible (Fig. 37e). The species themselves, although they at first seem to be quite similar to one another in morphology, have many distinguishing features. The configuration of the frontal margin of the frontal plate is one of them (Fig. 451). Others include the absence or presence and the number of lateral spines on the cephalon border or the pleotelson border. The configuration of the uropods is also an important feature, but it is not used here because it must be viewed at high magnification. As further more intensive collecting is made in North American tropical waters more species will most probably be found.

KEY TO THE SPECIES OF JAEROPSIDAE

1a. Frontal plate (Fig. 451) of cephalon rounded or obtusely pointed; spines on lateral margins of pleotelson absent or present

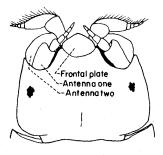


Figure 451. Cephalon of Jaeropsis species.

Fig. 452. Jaeropsis coralicola (Schultz and McCloskey) (2.8 mm)

Range: Cape Lookout, North Carolina (10 m)

In the deep water off North Carolina large coral heads which are the home of many animals including isopods are found on rock jetties.

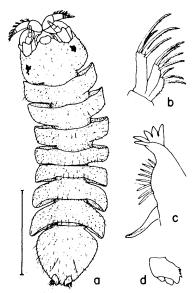


Figure 452. a. Jaeropsis coralicola. b. Maxilla two. c. Mardible, left. d. Uropod.

Fig. 453. Jaeropsis lobata (Richardson) (3.5 mm)

Range: Monterey Bay, California (Shallow water)

The species is not at all well defined and more specimens must be collected for a more proper definition.

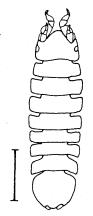


Figure 453, Jaerop-

- 2b. Lateral margin of pleotelson with spines; eyes dorsally placed on cephalon 3

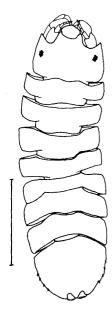


Fig. 454. Jaeropsis rathbunae (Richardson) (3.2 mm)

Range: Bermuda (Shallow water)

The original description of this species stated that there were no spines on the lateral borders of the pleotelson. The four shown here were present when the type specimen (the specimen on which the description of Richardson was based) was examined under high magnification.

Figure 454. Jaeropsis rathbunae.

- 3b. Frontal plate rounded; more than four spines on lateral margin of pleotelson 4
- 4a. Body somewhat elongate; lateral margins of peraeon concave

 J. concava



Fig. 455. Jaeropsis concava (Schultz) (3.1 mm)

Range: Santa Cruz Canyon on shelf off southern California (221 m)

figure 455, Jaeropsis concava,

Fig. 456(a). Jaeropsis dubia (Menzies) (1.8 mm)

Range: Mid-California to northern Baja California (18 to 91 m)

This species is usually collected with algae. A subspecies, J. dubia paucispinis, has been recorded which differs from the parent species in that there are less spines on the lateral margins of the pleotelson and there is different dorsal color pattern.

Menzies (1951) considered this a "low intertidal" species. Brazil (Pires, 1981)

Gulf of Calif. (Carvacho, 1983)

Figure 456(a). Jae-

5b. Lateral margins of cephalon coverage anteriorly with no lateral margin indentations; body covered with long setae

J. setosa

Fig. 456(b). Jaeropsis setosa (George and Stromberg) (2.3 mm)

Range: Puget Sound, Washington (50 m)

The species was collected with red and brown algae.

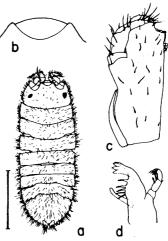


Figure 456(b). a. Jaeropsis setosa. b. Frontal plate of cephalon. c. Maxilliped. d. Mandible, right.

MUNNOPSIDAE

Munnopsis is the only genus of the family found in North American waters. Peraeopods I and II are gnathal, and II and IV are extremely long and ambulatory. Peraeopods V, VI and VII are natatory and the dactylus is absent. Antenna two is as long as peraeopods II and III. Antenna one is much shorter and with a broad basis. Most species of the family are truly planktonic or at least able to swim well and found above the bottom of the sea. Two species, one found principally in near-shore and the other in open sea habitats are described here.

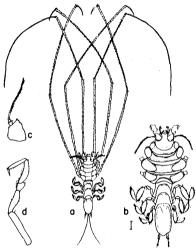


Figure 457(a), a. Munnopsis typica (male), b. Female, c. Antenna one, c. Peraeopod I (see also Fig. 372).

Fig. 457(a). Munnopsis typica (M. Sars) (13 mm)

Range: East Greenland, Cape Cod to Arctic Sea (9 to 732 m)

This moderately large asellote species with its very long legs swims backward as well as forward. It is able to extend its legs to the side so that it can get its body close to the bottom. The species is found in algae in the near-shore and in some deep-sea locations.

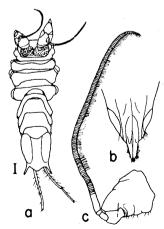


Fig. 457(b). Munnopsis longiremis (Richardson) (15 mm)

Range: Pacific Ocean off Costa Rica (Open ocean)

The species is a true planktonic species. It was taken at 1485 m in water that was 3570 m deep.

Figure 457(b). a. Munnopisis longiremis. b. Apex pleopod 1, male. c. Antenna one.

MUNNIDAE

The species of the Munnidae, especially those of the genus *Munna*, are the daddy long leg spiders of the isopod world. All have very long spider-like legs and have small bodies. Some species have a compact body and the ends of all peraeonal segments are closely adherent along their entire lengths. Other species are more dorsoventrally flattened and have peraeonal segments on which the ends are separated widely from each other.

Species of the genus *Munna* are common in the near-shore habitat, but species are also present in the deep-sea. Because of their small size, many specimens were poorly observed and described, hence, many apparently widespread species will undoubtedly be divided into several species when specimens are reobserved and redescribed. Species of the other genera mentioned here have been taken in the near-shore and deep-sea habitats and they are abundant at times. Members of the family although small in body are abundant and have a very important role to play in the habitats of which they are a part.

The peraeopods and antenna two of the specimens of the family are usually long. Antenna one and the uropods on the other hand are generally short. The two apical articles of the maxillipedal palp are in most instances much narrower than those of the first three articles.

KEY TO THE GENERA AND SPECIES OF MUNNIDAE

1a. Eyes, if present, visible in dorsal view 2

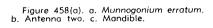
1b. Eyes visible only in ventral view Munnogonium

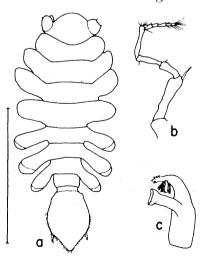
Fig. 458(a). Munnogonium erratum (Schultz) (1.8 mm)

Range: Santa Barbara Channel, near Gaviota, California 135 m)

This species has eyes that are on stalks which are bent downward so that the eyes are visible only in the ventral or ventrolateral view.

see Bowman & Schultz (1974)





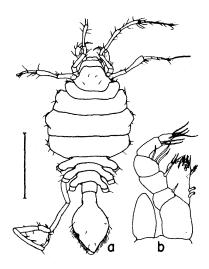


Fig. 458(b). Munnogonium waldronensis (George and Stromberg) (1.8 mm)

Range: Washington State (50 m)

Figure 458(b), a. Munnogonium waldronense, b. Maxilliped.

GENUS PLEUROGONIUM

This small group of species is blind and lives on muddy bottoms. Four species, three of which have been recorded from both northeastern United States and Norway are found in North American waters.

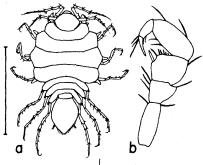


Figure 459. a. Pleurogonium rubicundum. b. Peraeopod I.

Fig. 459. Pleurogonium rubicundum (Sars) (1.5 mm)

Range: Puget Sound, Washington and St. Andrews, New Brunswick, Canada (4 to 9 m) Fig. 460. Pleurogonium inerme (Sars) (2 mm)

Range: Bay of Funday, Canada (4 to 27 m)

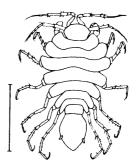


Figure 460. Pleurogonium inerme,

Fig. 461. Pleurogonium spinosissmum (Sars) (2.3 mm)

Range: Bay of Fundy, Canada (4 to 137 m)

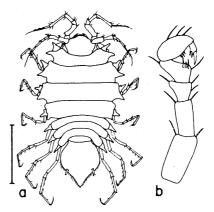


Figure 461. a. Pleurogonium spinossimum. b. Peraeopod I.

Fig. 462. Pleurogonium californiense Menzies (1.4 mm)

Range: Sonoma to Pt. Loma, California (93 to 99 m)

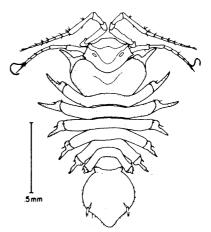
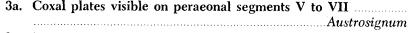


Figure 462. Pleurogonium californiensis.



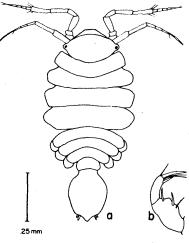


Figure 463. a. Austrosignum tillerae. b. Peraeopod I.

Munnogonium Fig. 463. Austrosignum tillerae (Menzies) and Barnard)

(Menzies) and Barnard) (1 mm)

Range: Pt. Loma, California to

Mexican border (18 to 183 m)

The other species of the genus have been taken in the Antarctic Sea.

See Bowman & Schultz, 1974: 266

3b. Coxal plates visible on peraeonal segments II to VIIMunna

GENUS MUNNA

The genus *Munna* contains a great many species that are common in the near shore habitats along all continents of the world. Species are also found in moderately deep water and in the deepsea. The animals are small and difficult to see even with a microscope so that the ones that have been described were described in a very cursory fashion, and it is difficult to make comparisons of them and newly discovered specimens. However, some attempts have been made to assemble all species of the genus and put them into subgroups or subgeneric categories.

KEY TO SPECIES OF MUNNA

4a.	Eyes	present		 	 	 	 	 5
44	Blind		}					

Fig. 464. Munna truncata (Richardson) (2 mm)

Range: Nova Scotia to south of Martha's Vineyard (146 to 713 m)



Figure 464. Munna truncata

- 5a. With few, if any, spines (not setae) or projections on lateral parts of body 6
- 5b. With many small spines (not setae) or lateral projections on anterior margin of cephalon, on lateral edges of peraeonal segments and on anterior parts of lateral borders of pleotelson

 M. spinifrons

Fig. 465. Munna spinifrons (Menzies and Barnard) (1.5 mm)

Range: Off Pt. Conception, California (12 m)

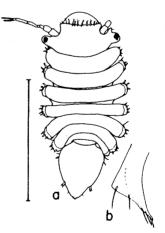


Figure 465. a. Munna spinifrons. b. 1/2 pleopod 1, male.

6a. No ventral posteriorly projecting extension of pleotelson visible in dorsal view (not to be confused with projecting pleopods in preserved specimens); frontal margin of cephalon concave or rounded.

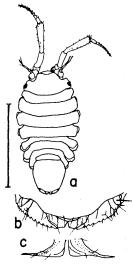


Figure 466(a). a. Munna halei. b. Detail, posterior margin pleotelson. c. Apex pleopod 1, male.

Fig. 466(a). Munna (Munna) halei (Menzies) (1.5 mm)

Range: Mid-California (Intertidal) (Tona/es Bay to San Lois obis po

The species is found under stones and the holdfasts of algae, 4 in sea urchin spines (Iverson, 1975).

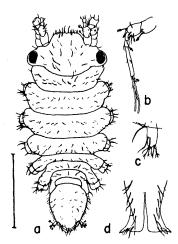


Figure 466(b). a. Munna fernaldi. b. Antenna one. c. Uropod. d. Apex pleopod 1, male.

Fig. 466(b). Munna (Munna) fernaldi (George and Stromberg) (1.5 mm)

Range: Friday Harbor, Washington (Intertidal)

The species is from algae among stones.

8a.	Frontal margin of cephalon concave 10
8b.	Frontal margin of cephalon broadly rounded or straight 9
9a.	Frontal margin broadly rounded; peraeonal segment VII as wide as or slightly narrower than pleotelson 10
9b.	Frontal margin straight; peraeonal segment VII wider than pleotelson

Fig. 467. Munna (Uromunna) reynoldsi (Frankenberg and Menzies) (1.5 mm)

Range: Georgia (Coastal swamps)

This species was taken in oak leaves on the bottom of shallow water in the swamps and marshes along the Georgia seacoast. It, like *M. ubiquita* and *M. magnifica*, is in the subgenus *Uromunna*.

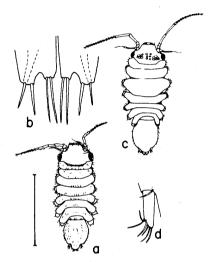


Figure 467. a. Munna reynoldsi (male). b. Apex pleopod 1, male. c. Female. d. Uropod.

Fig. 468. Munna (Neomunna) kroyeri (Goodsir) (3 mm)

Range: Greenland (18 to 110 m)

West coast records of this species most always refer to *M. stephenseni* mentioned below. It is found in Norway as well as Greenland.

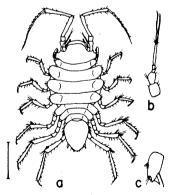


Figure 468. a. Munna kroyer, b. Antenna one, c. Uropod.

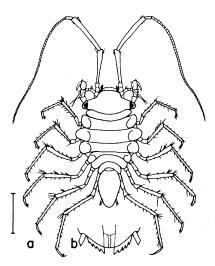


Figure 469. a. Munna fabricii. b. Detail, ventral aspect posterior margin of pleotelson.

Fig. 469. Munna (Neomunna) fabricii (Kroyer) (3 mm)

Range: Greenland to Maine (7 to 366 m)

Both this species and *M. halei* have a posteriorly directed projection from the ventral part of the pleotelson. In *M. halei* it is generally apparent in the dorsal view; in *M. fabricii* it is generally hidden in the dorsal view. The frontal margin of the cephalon in *M. halei* is concave; whereas, it is broadly rounded in *M. fabricii*.

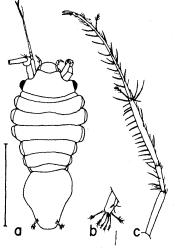


Figure 470. a. Munna magnifica. b. Uropod. c. Peraeopod V.

Fig. 470. Munna (Uromunna) magnifica (Schultz) (2 mm)

Range: Near Santa Barbara Island off southern California (500 m)

The species was taken from a bottom with black mud, nodules and flat shaley rocks.

12a. Body with about 1/2 length1312b. Body width about 1/3 lengthM. chromatocephala

Fig. 471. Munna (Neomunna) chromatocephala (Menzies) (2.2 mm)

Range: Puget Sound, Washington to mid-California (Intertidal)

This species is found among encrusting organisms on rocks and piles.

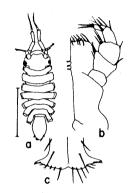


Figure 471. a. Munna chromatocephala. b. Maxilliped. c. Apex pleopod 1, male.

Fig. 472. Munna (Neomunna) stephenseni (Gurjanova) (0.7 mm)

Range: Bering Sea to mid-California (Intertidal to 18 m)

The records of *M. kroyeri* from western North America refer to this species.

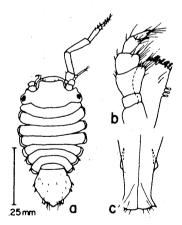


Figure 472. a. Munna stephenseni, b. Maxilliped, c. Apex pleopod 1, male.

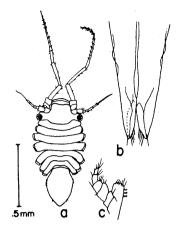


Figure 473. a. Munna ubiquita. b. Apex pleopod 1, male. c. Maxilliped.

Fig. 473. Munna (Uromunna) ubiquita (Menzies) (1.2 mm)

Range: Puget Sound, Washington to northern Baja California (Subtidal to 37 m)

San Quintin Bay (Meuzies, 1962) N.W. Gulf of Calif. (Carvacho, 1983)

MESOSIGNIDAE

The family Mesosignidae is composed of one genus, Mesosignum. The species are very small, pigmentless and blind and are found in the deep sea. Peraeonal segment I is short being only slightly broader than the cephalon. Neither the cephalon or peraeonal segment I bear spines or lateral lappets. The palp of the mandible has three articles and the palp of the maxilliped is narrow with four articles. The uropods are uniramus. Two species from the Caribbean Sea are mentioned here.

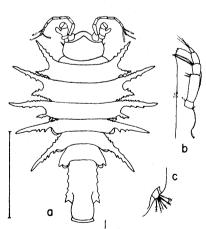


Figure 474. a. Mésosignum kohleri. b. Maxilliped. c. Uropod.

Fig. 474. Mesosignum kohleri (Menzies) (2.5 mm)

Range: Caribbean Sea, north of Columbia (4076 m)

The species has a pleotelson with two short posterolateral spines. The lateral margins of the pleotelson are parallel.

Fig. 475. Mesosignum usheri (Menzies) (2 mm)

Range: Caribbean Sea off Cartagena, Columbia (2875 to 2941 m)

The species was found near to *M. kohleri*, but it differs from that species in that it has long posterolateral spines on the pleotelson. The lateral margins of the pleotelson are convex.

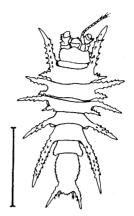


Figure 475. Mesosignum usheri.

DENDROTIONIDAE

The species in the family are similar to those in the family Munnidae. They, however, have spines on their bodies and many species have in addition long lateral lappets. Both species with eyes and blind ones are included in the family. The apical 2 articles on the maxillipedal palp of 5 articles are much narrower than the endite of the maxilliped itself (Fig. 370p). The peraeopods, except for the first one which is gnathal, are ambulatory and with a single dactyl claw. Only two species, each in a different genus, are found in North American waters.

KEY TO THE GENERA OF DENDROTIONIDAE

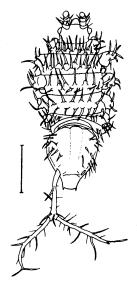


Figure 476. Acanthomunna tannerensis,

Fig. 476. Acanthomunna tannerensis (Schultz) (3.5 mm)

Range: Tanner Canyon on shelf off southern California (813 m)

The uropods on this specimen are the only ones to have ever survived the rough process of sorting to which isopods and other benthic animals taken from the deep sea are usually subjected. Whether they are representative of the uropods found on other species is not known.

1b. Eyes or lateral bulges absent; cephalon quadrangular or longer than broad; lateral spines on all peraeonal segments simple or serrated, but never multibranchedDendrotion

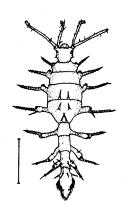


Figure 477. Dendrotion hanseni.

Fig. 477. Dendrotion hanseni (Menzies) (3.5 mm)

Range: Caribbean Sea, south of Jamaica (1244 m)

This species is easily distinguished from other asellotes by the very long pointed lateral lappets on the edges of the peraeonal segments. It is interesting to note that those coming from peraeonal segments I to IV come from the edge of the peraeonal segment itself, but those arising from segments V to VII are really from the coxal segments of the peraeopods, not from the segment edges.

ILYARACHNIDAE

There are about 36 species in the widely distributed genus Ilyarachna, but there are only 3 recorded from North American

waters. The species are characterized by long peraeopods, a wide cephalon and narrow pleotelson. Peraeopod I is gnathal, peraeopods II to IV are ambulatory and the last three are natatory. Occasionally the species are taken in plankton tows.

Fig. 478. *Ilyarachna acarina* (Men zies and Barnard) (4 mm)

Range: Southern California coastal slopes (73 to 507 m)

The species is common and abundant in the moderately deep water off southern California and northern Mexico.

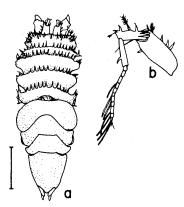


Figure 478. a. Ilyarachna acarina. b. Antenna one.

Fig. 479. Ilyarachna profunda (Schultz) (3 mm)

Range: La Jolla Canyon on shelf off southern California (461 to 1298 m)

The species lacks the conspicuous spines on the anterior borders of the peraeonal segments that are found in *I. acarina*. It also was taken in much deeper water than that species.

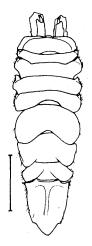


Figure 479. Ilarachna profunda.

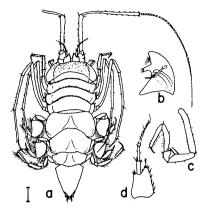


Figure 480. a. Ilyarachna hirticeps. b. Mandible. c. Peraeopod I. d. Antenna one.

Fig. 480. Ilyarachna hirticeps (Sars) (7.5 mm)

Range: Greenland (20 to 435 m)

The species has very small spines on the anterior margins of the anterior peraeonal segments.

EURYCOPIDAE

The many species of this family are divided into 12 genera which are placed in four subfamilies. Species of three genera, in three of the subfamilies, have been recorded from North American waters. The peraeopods of the specimens are long and of two types. Peraeopods I to IV are ambulatory (peraeopod I can be gnathal) and peraeopods V to VII are natatory. A dactylus is present on the natatory peraeopods (it is absent on natatory peraeopods in members of Munnopsidae, a closely related family). The small body and very large peraeopods of the isopods enable the species to swim well and they are caught, sometimes in very large numbers, in plankton tows. Species of the family are blind and frequently have peraeonal segments fused with each other or with the pleon. The cephalon is usually separated from the peraeon. Both uniramus and biramus uropods are present among the species. The maxillipedal palp of five articles has the two distal narrower than the other three. The wider of the three basal articles has a particular pattern of lobes and/or setae on the medial border.

KEY TO THE GENERA AND SPECIES OF EURYCOPIDAE

1a. Uropods uniramus; pleotelson suboval with two pairs of lateral and one terminal spine; peraeopods I to IV of subequal length; peraeopods V to VII with narrow carpus and propodus (natatory setae present)

Acanthocope

Fig. 481. Acanthocope spinosissima (Menzies) (8 mm)

Range: Caribbean Sea, south of Jamaica (1224 m)

The food of this species, as stomach content analysis has revealed, consists of what are probabably bacteria, protozoa and forminifera (a particular kind of abundant calcareous protozoan). Also polychaeta (a sea bottom worm related to the earthworm) and sponge spicules (the calcareous or silicate frame work of sponges) have been found. The isopod then apparently eats either live or dead animal and bacterial material that is present in its habitat in the deep sea.



Figure 481. Acanthocope spinosissima.

- 1b. Uropods biramus; pleotelson rarely with two pairs of lateral spines; peraeopod I much shorter than III and IV; peraeopods V and VII generally with carpus and propodus moderately or greatly expanded 2
- 2a. Maxilliped with articles four and five or palp abruptly narrower than article three and with medial margins never produced; articles three with fringe of setae or crenulations or both or medial border (Fig. 370e) Syneurycope

Fig. 482. Syneurycope parallela (Hansen) (3.7 mm)

Range: North Atlantic to past Cape Farewell (3474 m)

This species has coupling hooks on the medial edge of the maxilliped; S. hanseni, mentioned below, has none.

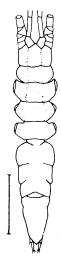


Figure 482. Syneurycope parallela

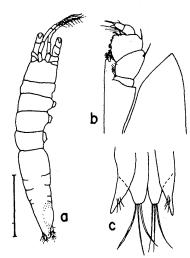


Figure 483. a. Syneurycope hanseni. b. Maxilliped. c. Apex pleopod 1, male.

Fig. 483. Syneurycope hanseni (Menzies) (3.8 mm)

Range: North of Puerto Rico (5104 to 5122 m)

The lateral margins of the body in this species are convex when compared to the parallel body margin in S. parallela.

GENUS EURYCOPE

This large genus of about 36 species contains most non-spined members of the family. The body is generally oval and there are long peraeopods. Only the four following species have been recorded from North American waters until now.

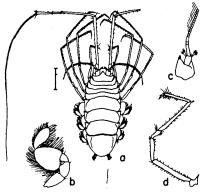


Figure 484. a. Eurycope cornuta. b. Peraeopod V. c. Antenna one. d. Peraeopod I.

Fig. 484. Eurycope cornuta (Sars) (4 mm)

Range: Greenland, Gulf of St. Lawrence and Atlantic coast of North America (91 to 732 m)

This species has a pointed rostrum or medial cephalic projection.

Fig. 485. Eurycope complanata (Bonnier) (5 mm)

Range: Davis Strait and southeast of Bermuda (2258 to 5779 m)

This species has a bifid medial cephalic projection.

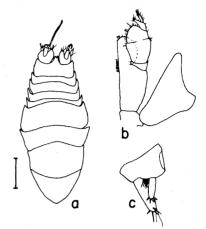


Figure 485. a. Eurycope complanata. b. Maxilliped. c. Uropod.

Fig. 486. Eurycope mutica (Sars) (1.5 mm)

Range: Bay of Fundy to Norway (9 to 27m)

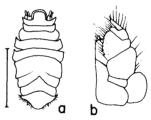


Figure 486, a. Eurycope mutica. b. Maxilliped.

Fig. 487. Eurycope californiensis (Schultz) (3.5 mm)

Range: Newport Canyon on shelf off southern California (478 m)

The species has a produced, rounded medial cephalic projection.

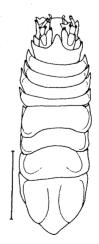


Figure 487. Eury-

GENUS STORTHYNGURA

The members of this genus were considered to be members of the genus *Eurycope* by the people who first describe them. Now there are about 32 species in the genus. Several species listed here are probably the same species only one is the adult and the other the juvenile.

KEY TO SPECIES OF STORTHYNGURA



Figure 488. Storthyngura truncata.

Fig. 488. Storthyngura truncata (Richardson) (3.8 mm)

Range: Georges Bank and near Martha's Vineyard (2788 to 3225 m)

This species based on female specimens might prove to be the young or female of S. magnispinis mentioned below (p. 308), but more specimens must be collected before it can be definitely established.

Fig. 489. Storthyngura snanoi (Menzies) (3.2 mm)

Range: Caribbean Sea, north of Columbia (4071 m)

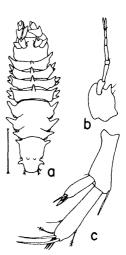


Figure 489, a. Storthyngura snanoi, b. Antenna one. c. Uropod.

Fig. 490. Storthyngura caribbea (Benedict) (12 mm)

Range: Windward Islands, West Indies (1256 m)

The species has been considered as a subspecies; i.e., S. pulchra caribbea.

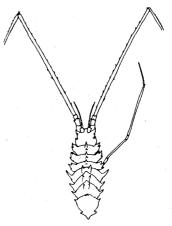


Figure 490. Storthyngura caribbea.

- 7a. Lateral lappets on peraeonal segments I, II and III at least as long as segment is long; coxal plates not apparent
 - S. magnispinis

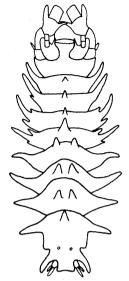


Figure 491. Storthyngura magnispinis.

Fig. 491. Storthyngura magnispinis (Richardson) (4 mm)

Range: Off Nantucket Shoals (2258 to 2704 m)

The species description is based on a single male specimen. It should be compared with specimens of *S. truncata* mentioned above (p. 306).

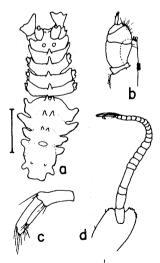


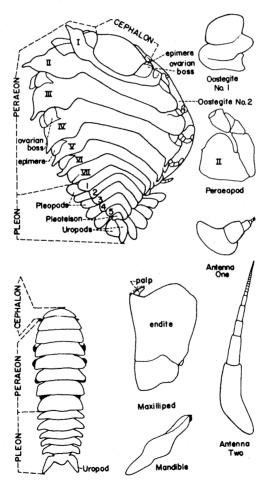
Figure 492. a. Storthyngura vemae. b. Maxilliped. c. Uropod. d. Antenna one.

Fig. 492. Storthyngura vemae (Menzies) (3.2 mm)

Range: Caribbean Sea, north of Columbia (4071 m)

SUBORDER EPICARIDEA

In the past the suborder Epicaridea has been called Bopyroidea. The species of the suborder range from very primitive isopods that are much like the archetype in body segmentation to very highly modified species that only superficially resemble other isopods. Epicaridean isopods undergo greater changes during development than do species of other groups. The egg develops into a larva with six peraeonal segments, the "epicaridean" stage. Animals of this stage free themselves from the marsupium and attach to a planktonic host (mostly copepods). There they molt to a stage with



Figure, 493. Epicaridea (General nomenclature),

seven segments, the "microniscus" stage. The animals then molt again to a "cryptoniscus" stage and begin sexual differentiation (both males and females). The maturing juveniles detach from the first host and become planktonic again, then attach to a second host (almost any crustacean including other isopods) and grow. Through metamorphosis and maturity the female becomes somewhat of a peculiar egg case sometimes becoming quite indistinguishable as an isopod (Fig. 7h, i).

The male attaches to the pleopods or gets into the egg case of a female which is attached to a host and which is much larger than the male isopod. There it changes in form slightly, matures sexually, but does not usually grow. It is then able to fertilize the eggs of the female and the life cycle is repeated. The relations between the hosts and the life stages of the parasitic isopods are only beginning to become known, and the above stages have never been clearly recorded for any one species. Four families are recognized here as being present in North American waters although some workers recognize more.

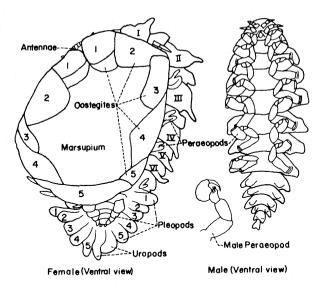


Figure 494. Epicaridea (Ventral view, general nomenclature).

KEY TO FAMILIES OF EPICARIDEA

1b.	Not as above
2a.	Body without indications of rigid exoskeleton; body segmentation present; peraeons laterally expanded into thin, pleural lamellae
2b.	Not as above 3
3a.	Body of female symmetrical or irregularly symmetrical in outline; segmentation apparent, but reduced; peraeopods from two to five; mouth in form of sucker
3b.	Body of female distinctly segmented and more or less asymmetrical; seven pairs of peraeopods, six of which can be absent on one side (exception, peraeopods I which are always paired) Bopyridae (p. 312)

BOPYRIDAE

There are more described species of Bopyridae than in the other epicaridean families in North American waters. The species are parasitic on decapods (crabs and shrimp). The body of both the male and female is segmented, and peraeopods are present and differ little in size and shape. Females have large asymmetrical bodies and males are small and symmetrical, frequently being found attached to or among the pleopods, or in the broad pouch of the female. The cephalon is generally distinct and eyes, if present, are tiny. The antenna and mouth parts are rudimentary. All seven segments of the peraeon are always laterally distinct, but in some instances some of the anterior segments are fused with the cephalon. Five pairs of oostegites are present, and knowledge of their form is useful since the most commonly encountered bopyrid is the gravid female. Coxal separations are present on the edges of the peraeonal segments, and large bosses or swellings are frequently present just medially to the coxal sutures. Seven pairs of prehensile peraeopods are almost always present, but in one group peraeopods are absent on one side on peraeons II to VII.

The pleon is always distinct and generally segmented or with indications of segmentation on the lateral margins. The lateral margins are frequently elongate. The pleopods are sometimes absent and when present they can be uniramus, biramus or triramus. They can be mistaken for extensions of the peraeonal segments if one is not careful. The uropods, if present, are generally simple and composed of lobes which look like the ends of the pleonal segments. A distinct telson is sometimes present, but many times it is fused to a pleonal segment to form a pleotelson.

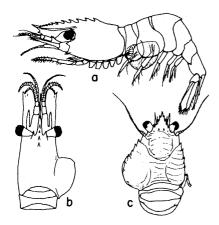


Figure 495. Bopyrid parasites on shrimp, a. On abdomen, b-c. Within branchial cavity of host (dorsal views). (After Sars),

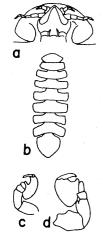


Figure Bathygyge ventral view grandis, cephalon. b. Dorsal view. c. Peraeopod I. d. Peraeopod

A small male is frequently found clinging to the pleopods of the female in some species or living within the brood pouch with the eggs in other species. The male is small, symmetrical and distinctly segmented. It has very small antennae, mouth parts and short prehensile peraeopods mostly have pointed dactyls. Sometimes the peraeopods are vestigal; i.e., consist only of stubs. The pleon is always distinct, but the segmentation of the pleon is not always distinct. The pleon usually lacks appendages, but a well defined

pleotelson with long uropods are present in some species. The uropods are generally no longer than the ends of the pleonal segments in most species. The bopyrids are generally found in the branchial cavity of their decapod hosts. The males that are pictured here with the females are many times drawn at a scale much greater than that of the female. Most of the males are under 1.5 mm long.

Only a large male specimen is known for one species, Bathygyge grandis Hansen (7 mm), from Acapulco, Mexico. It was taken from the branchial cavity of the shrimp Glyphocragon spinulosa, and is sufficiently different from other males to be described as a new species.

It is illustrated in figure 496.

KEY TO GENERA AND SPECIES OF BOPYRIDAE

1a. Body asymmetrical with one side greatly enlarged and longer than other side; only peraeopod I present on enlarged side; pleon composed of 4 segments and usually with pleotelson ... 2

1b. Body slightly to greatly asymmetrical, but with both sides more or less developed; i. e., with all peraeopods present although they might be small and difficult to find; pleon usually of 5 segments plus pleotelson (some segments are at least laterally indicated)

Fig. 497. Hemiarthrus subcaudalis (Hay) (6 mm)

Range: Onslow Bay, North Carolina (about 20 miles off Beaufort Inlet) (18 m)

The species is from the gill chamber of the shrimp Synal-pheus longicarpus which is found within the cavity of a sponge. The species of a different genus Synsynella deformans (p. 333) is found as a parasite on the same shrimp in the same locality. This is an example of two species of isopods exploiting the same host animal.

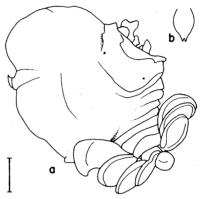


Figure 497. a. Hemiarthrus subcaudalis. b. Pleon male.

Fig. 498. Hemiarthrus abdominalis (Kroyer) (9 mm)

Range: Circumpolar south to Massachusetts and Washington State (9 to 642 m)

The species is found as a parasite on the decapods of the genera *Spirontocaris* and *Pandalus* inhabitants of northern waters.

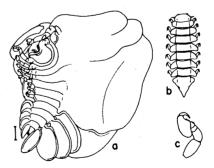


Figure 498. a. Hemiarthrus abdominalis, b. Male. c. Peraeopod of male.

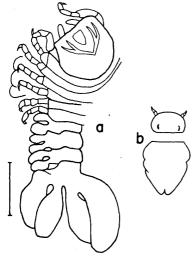


Figure 499. a. Hemiarthrus schmitti. b. Anterior and posterior parts of male.

Fig. 499. Hemiarthrus schmitti (Pearse) (4.6 mm)

Range: East coast and Bimini, Bahamas

The species is parasitic on the shrimp Synalpheus brooksi. The lateral extensions of the pleonal segments are bifid with those of segment 4 the longest in this species.

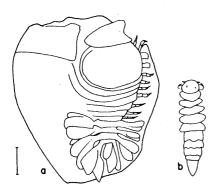


Figure 500(a). a. Diplophryxus synalphei, b. Male.

Fig. 500(a). Diplophryxus synalphei (Pearse) (6.2 mm)

Range: South Carolina near mouth of New River

The species is found on the snapping shrimp Synalpheus fritsmulleri elongatus.

3a. Cephalon more or less symmetrical with lateral lobes on anterior third and set slightly in front of body margin; pleon with five segments indicated; pleotelson (no telson indicated) with elongate, lobate uropods

Aporobopyrus

Fig. 500(b). Aporobopyrus muguensis Shiino (4 mm)

Range: Off Pt. Mugu, California (11 m)

The species is from the branchial cavity of *Pachycheles rudis*. A second species *A. oviformis* Shiino (3.5 mm) is also found in the same locality.

A. oviformis Shijno 1934

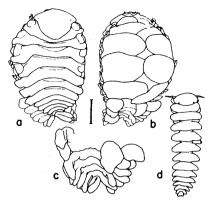


Figure 500(b). a. Aporobopyrus muguensis. b. Ventral view c. Ventral view, pleon. d. Male

- 3b. Cephalon various; pleon and pleotelson various 4
- 4b. Pleon with lateral extensions long at times, but never digitate or crenulate; pleotelson or telson various
- 5a. All pleonal extensions elongate and digitate 6
- 5b. All pleonal extensions with simple crenulate margins Ergyne

Fig. 501. Erygne rissoi (Nierstrasz and Brender (2 mm)

Range: Curacao, Dutch West Indies

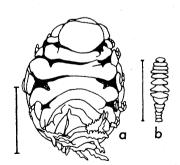


Figure 501. a. Ergyne rissoi. b. Male.

6a. Pleon with 6 segments visible; telson elongate, but never with digitate borders

Fig. 502. Ione thompsoni (Richardson) (16 mm)

Range: Massachusetts

The species is found on the mud shrimp Callianassa stimpsoni.

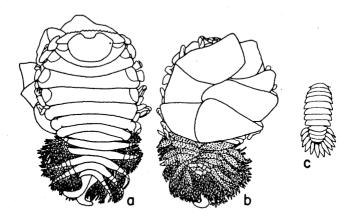


Figure 502. a. lone tompsoni. b. Ventral view. c. Male.

= I. cornuta

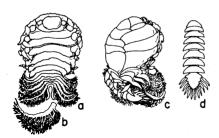


Figure 503. a. *Ione brevicaudata*. b. Detail, pleural lamella. c. Ventral view.

Fig. 503. Ione brevicauda (Bonnier) (6.7 mm) Range: San Francisco Bay, California

A third species has been found at Puget Sound, Washington, and is called *Ione cornuta* Spence-Bate (15.5 mm). It was found in the branchial cavity of *Callianassa longimana*, but it was never illustrated.

- 6b. Pleon with 6 or less segments; telson or pleotelson always with digitate boarders
- 7b. Cephalon with very little, if any, extension beyond lateral margin of oval cephalon (frontal extension present, however)

 Dactulokepon

Fig. 504. Dactylokepon hunterae (Wells and Wells) (6 mm)

Range: Off Core Bank, North Carolina (22 m)

The species is a branchial cavity parasite of the crab *Pinnotheres maculatus* which is associated with the calico scallop. Two specimens were found on the host.

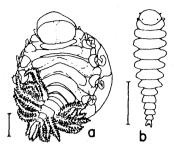


Figure 504. a. Dactylokepon hunterae. b. Male.

8a. Cephalon narrow, without frontal or lateral extensions Leidua

Fig. 505. Leidya distorta (Leidy) (8 mm)

Range: New Jersey, North Carolina and Bermuda

The species is found in the branchial cavity of the common fiddler crab, *Uca pugilator*, an abundant species in the salt marshes of the east coast of the United States. In Bermuda the species is parasitic on a different species of crab from a different family, *Pachygropsus transversus*. It thus is an example of a parasitic isopod which has several host species.

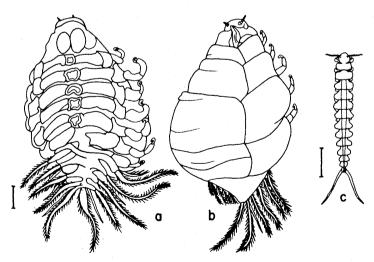


Figure 505. a. Leidya distorta. b. Ventral view. c. Male.

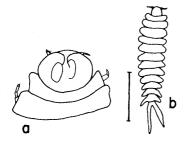


Figure 506. a. Leidya bimini, anterior part. b. Male.

Fig. 506. Leidya bimini (Pearse) (6.7 mm)

Range: Bimini Island, Bahamas

The species is found in the branchial chamber of *Pachygrapsus transversus*. It perhaps is really only a different form of *L. distorta*.

8b. Cephalon with large lateral and frontal extensions on ovate cephalon Grapsicepon

Fig. 507. Grapsicepon edwardii (Giard and Bonnier) (19 mm)

Range: Gulf Stream and Sargasso Sea

The species is a branchial parasite of crabs. Several males sometimes are found attached to one female. Large mediodorsal projections are found on peraeonal segments VI and VII.

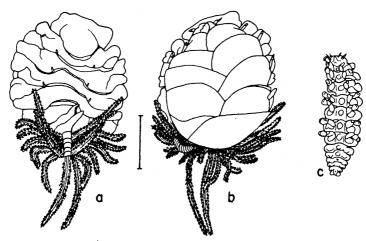


Figure 507. d. Grapsicepon edwardii. b. Ventral view. c. Male.

(6) M. irritans Boone Florida & Belize (host: M. irrasa)

(7) M. laterale Richardson. Philippines & Java Sea.

(host: ?)

Fig. 508. Grapsicepon choprae (Nierstrasz and Brender) (4mm)

Range: Curacao, Dutch West Indies

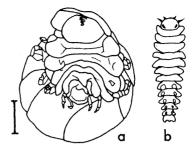


Figure 508. a. Grapsicepon choprae. b. Male.

- 9a. Pleonal terga produced into long pleural lamellae (Fig. 509a)
- 9b. Pleonal targa short, not produced into pleural lamellae (Fig. 509b)14

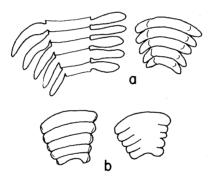


Figure 509. Pleural lamellae. a. long. b. Short.

note: There are 7 Species of N. American Munidon (see Markham,

- (1) Fig. 510. Munidion parvum (Richardson) (9 mm)

Range: Puget Sound, Washington and Departure Bay, British Columbia

The species is a parasite on the galatheid crab Munida quadrispina.

- (2) M. princeps Hansen. Pacific coast of Colombia (host: Munida refulgens Faxon).
- (5) M. pleuroncodis Markham. British Columbia to Baja (host: Ple roncodes planipes).
- (4) M. <u>cubense</u> Bourdon (fornerly M. parvum <u>cubensis</u>) Caribbean (hosts: <u>Munida</u> <u>stimpsoni & M. flinti</u>)





Fig. 525. Pseudione panopei (Pearse) (13.5 mm) Range: Beaufort, North Carolina

The species is parasitic on the crab Panopeus herbstei.



Figure 525. a. Pseudione panopei, posterior part. b. Oostegite 1. c. Posterior part, ventral view male.

- 23a. Pleon with five complete, free segments showing; mostly blind species ______24
- 24a. Pleon much narrower than peraeon, at least on short body side; posterior margin of pleotelson slit medially ... Palaegyge

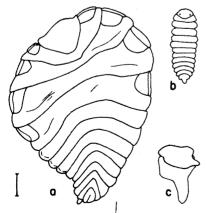


Figure 526, a. Palaegyge meeki. b. Male. c. Oostegite 1.

Fig. 526. Palaegyge meeki (Richardson) (9 mm)

Range: Freshwater streams on Atlantic coast of Panama

The species is a branchial parasite on the young of shrimps. The uropods of this species are in the form of two oval sac-like structures.

Fig. 527. Bopyroides hippolytes (Kroyer) (8 mm)

Range: Circumpolar, south to Cape Cod and to Puget Sound, Washington

There have been many different forms described for this widespread species that is parasitic on species of the genera *Pandalus*, *Pandalopsis* and *Spirontocaris*.

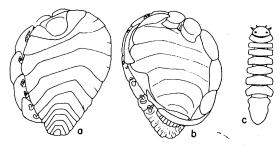


Figure 527. a. Bopyroides hippolytes. b. Ventral view. c. Male.

GENUS PROBOPYRUS

It is very difficult to tell the six species of the genus apart without knowledge of the host and of the structure of the male. The species are parasites of shrimp.

Fig. 528. Probopyrus alphei (Richardson)

Range: North Carolina to Brazil

The host species is the snapping shrimp Alpheus heterochaelis



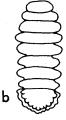


Figure 528. a. Probopyrus alphei. b. Male.

Figs. 529 and 530. Probopyrus pandalicola (Packard) (10 mm)

Range: New Hampshire to Florida and Mississippi

The species is parasitic on the shrimp of the genus *Palaemonetes* on the east and gulf coasts of the United States.

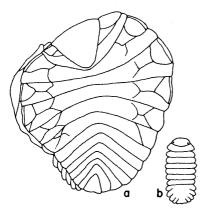


Figure 529. a. Probopyrus pandicola. b. Male.

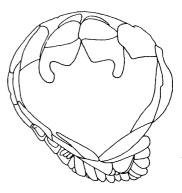


Figure 530. Probopyrus pandicola (ventral view).

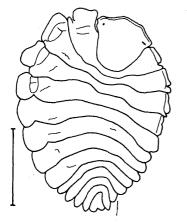


Figure 531. Probopyrus oviformis.

Fig. 531. Probopyrus oviformis (Nierstrasz and Brender) (2.5 mm)

Range: Jolly Hill, St. Croix, West Indies

The species is parasitic on the shrimp of small brooks of the genus *Macrobrachium*.

Fig. 532. Probopyrus panamensis (Richardson) (12 mm)

Range: Canal Zone, Panama

The species is a branchial parasite of the shrimp *Macrobrachium acanthurus*.

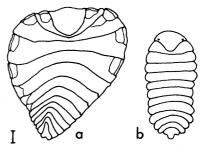


Figure 532. a. Probopyrus panamensis. b. Male.

Fig. 533. Probopyrus floridensis (Richardson) (4 mm)

Range: Northern Florida

The shrimp Palaemonctes exilipes is the host of the species.

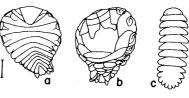


Figure 533. a. Probopyrus floridensis. D. Ventral view. c. Male.

Fig. 534(a). Probopyrus bithynis (Richardson) (3 mm)

Range: Mississippi River to Nicaragua

Shrimp of the genus *Macro-brachium* are the hosts.

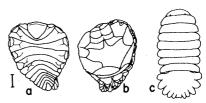


Figure 534(a). a. Probopyrus bithynis. b. Ventral view. c. Male.

26b. Cephalon never fused with peraeonal segment I and II ... 29

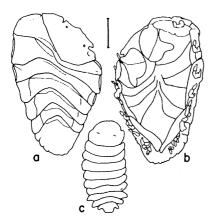


Figure 534(b). a. Bopyrella harmopleon. b. Ventral view. c. Male.

Fig. 534(b). Bopyrella harmopleon (Bowman) (4.8 mm)

Range: Los Roques Islands, Venezuela

The species is a parasite of a shrimp of the genus Synal-pheus.

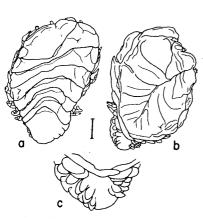


Figure 534(c). a. Bopyrella maginitiei. b. Ventral view. c. ventral view, pleon.

this of B. so-so-aban

Fig. 534(c). Bopyrella maginitiei Shiino v (5.7 mm)

Range: Santa Cruz Island, California (13 m)

The species is a branchial cavity parasite of the shrimp Crangon equidactylus. & Synalpheus

lockingtoni

 Fig. 535. Synsynella deformans (Hay) (3.1 mm)

Range: Onslow Bay, North Carolina, (20 miles off Beaufort Inlet) (18 m)

The species is found in the gill chambers of Synalpheus longicarpus, a shrimp. Hemiarthrus subcaudalis (p. 313) is also a parasite on the shrimp.

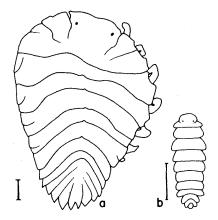


Figure 535. a. Synsynella deformans. b. Male.

Fig. 536. Bopyrinella antillensis (Nierstrasz and Brender) (3 mm)

Range: Curacao, Dutch West Indies

The host shrimp is *Thor floridanus* which is also the host of *Bopyrina thorii* in Florida (p. 335).

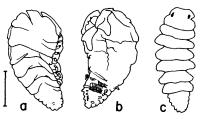


Figure 536. a. Bopyrinella antillensis. b. Ventral view. c. Male.

29a. Three or four free pleonal segments; pleotelson various 33

29b. All peraeonal segments fused, but generally indicated laterally (on one side or ventral view if not indicated on both sides in dorsal view); pleotelson never dorsally indicated (i. e., pleotelson equals pleon)

Bopyrina

GENUS BOPYRINA

The six species of the genus are found on five different shrimp genera. Two species are found on the same host from different geographic regions.

KEY TO SPECIES OF BOPYRINA

30a.	Pleon with indications of segmentation on one lateral margin only
30b.	Pleon with indications of segmentation of both lateral margins
31a.	Body asymmetrical, but medial axis almost straight B. crangona

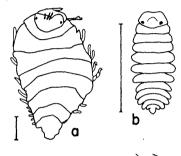


Fig. 537. Bopyrina crangona (Pearse) (5.5 mm)

Range: Coast of Carolinas (Shallow water)

The host shrimp is Crangon formosa.



Figure 537. a. Bopyrina crangona. b. Male. c. Pleon, ventral view.

31b. Body asymmetrical with curved medial axis B. abbreviata

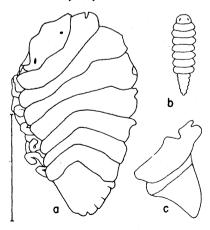


Figure 538. a. Bopyrina abbreviata. b. Male. c. Oostegite 1.

Fig. 538. Bopyrina abbreviata (Richardson) (2 mm)

Range: Puntarasa, Florida
The host shrimp is Hippolyte zostericola.

32a.	First incubatory (Fig. 542b)					
32b.	First incubatory (Fig. 538c)					
	Peraeonal segmen	•	-	_	_	

Fig. 539. Bopyrina thorii (Richardson) (3.2 mm)

Range: Key West, Florida

The host is Thor floridanus the same as for the species Bopyrinella antillensis (p. 333)

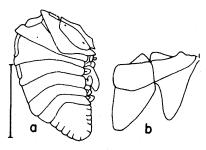


Figure 539. a. Bopyrina thorii. b. Oostegites 1.

33b. Peraeonal segment I not deeply set into peraeonal segment II B. latreuticola

Fig. 540. Bopyrina latreuticola (Gissler) (1.7 mm)

Range: Carolinas, Bermuda and Bahamas

Host animal for the species is Latreutes ensiferus. The species formerly was considered to be a member of Probopyrus, but because of the presence of eyes, the size, the fused pleon and the general body of configuration, it is included here in Bopyrina.

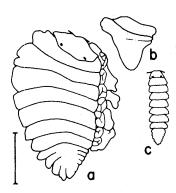


Figure 540. a. Bopyrina latreuticola. b. Oostegite 1. c. Male.

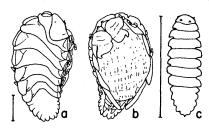


Figure 541. a. Bopyrina pontoniae. b. Ventral view. c. Male.

Fig. 541. Bopyrina pontoniae (Wells and Wells) (6 mm)

Range: Off Core Bank, North Carolina (22 m)

The host is the shrimp Pontonia margarita and it was associated with callico scallop. B. urocardis was found on the same host in Florida.

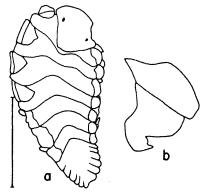


Figure 542. a. Bopyrina urocardis. b. Oostegite 1.

Fig. 542. Bopyrina urocardis (Richardson) (2.1 mm)

Range: Puntarasa, Florida

The host shrimp is *Pontonia* margarita the same as for *B.* pontoniae from North Carolina.

35a. Four free pleonal segments; fifth indicated in lateral margins of pleotelson by indentations Bopyriscus

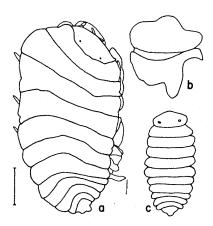


Fig. 543. Bopyriscus calmani (Richardson) (5 mm)

Range: Southern California Host unknown.

Figure 543. a. Bopyrina calmani. b. Oostegite 1. c. Male.

35b. Three free pleonal segments; fourth laterally indicated by slits in pleotelson margin

Bopyro

Fig. 544. Bopyro choprae (Pearse) (6.2 mm)

Range: North Carolina to Bahamas

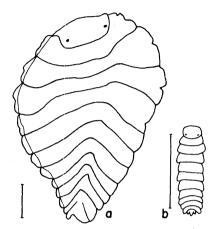


Figure 544. a. Bopyro choprae. b. Male.

CRYPTONISCIDAE

The species of the family are among the least known members of the isopod fauna of North American waters, because the female is composed of little more than just an egg sac and the males have rarely been found. It is known, however, that the species are isopods (for some species there is even a legitimate doubt of that!) because the juvenile stage looks like that of the juvenile of other species of bopyrids. Apparently the juveniles are at first planktonic and fend for themselves soon after being released from the brood sac. They soon must attach to a host, and most of the early attachment stages have never been seen by scientists because they are small and easily overlooked. The life history of no species has ever been completely explored and explained.

Most appendages including the mouth parts are absent, and the appendages which remain are greatly modified as suctorial or clinging structures. The females are composed mainly of ovarian tissue, and of eggs formed from ovarian tissue. Sometimes some sign of segmentation is found, but most frequently the form of the adult female is simply rotund or lobular, and there is only a general pattern to the body form of the various species. The details of the form depend upon the place and conditions of attachment

to the host. The males are like the males of other families of the suborder, but unfortunately they have never been extensively illustrated even when found. The isopods are probably frequently overlooked, even as adults, because they are small and look like many of the other encrusting plants and animals found on the host.

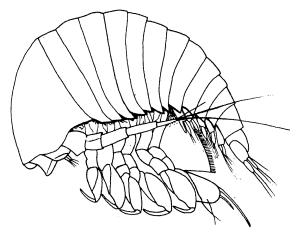


Figure 545. Clyptoniscus meinerti, lateral view larva.

There is some evidence that species of the family are host specific, but there are not enough observations to form any solid conclusions. The approximately 25 genera of the family have been divided into seven separate families, but since little other than the juvenile stage is known in most species, they are included here as one family. Members of the family are parasites on many other groups of crustaceans including other isopods. Occasionally they are parasites of crustaceans which are in turn parasites of other crustaceans or other animals. Probably they have some influence on the life history of their hosts, but not as much as that of the entoniscids. Six species in six genera have been recorded in North American waters. The key to genera which follows is only an aid, not the final word since the form of the animals is so changeable.

KEY TO GENERA AND SPECIES OF CRYPTONISCIDAE

1a. Specimen rolled into ball; much longer than wide Danalia

Fig. 546. Danalia fraissei (Nierstrasz and Brender) (4.4 mm)

Range: Curacio, Dutch West Indies.
The host of the parasite is the

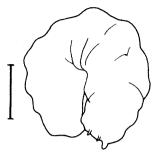


Figure 546. Danalia fraissei, lateral view.

1b. Not as above

Fig. 547. Clypeoniscus meinerti (Giard and Bonnier) (2 mm) Range: Greenland (11 to 18 m)

The species illustrated here is a parasite in the marsupium of another isopod, *Synidotea nodulosa* (p. 70), which is a relatively small isopod about 7.5 mm long. The host is found in the intertidal zone on the coast of Greenland. The species looks like a large egg in the marsupium of the female. Upon close examination it is found to be a sac of tiny eggs itself. A male is sometimes found attached to the egg mass and it looks much like other bopyrid isopod males.

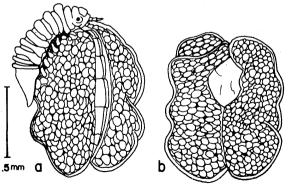


Figure 547. a. Clyptoniscus meinerti, female with male. b. Opposite view of a.

- 2b. Specimen not of only two lobes of egg masses; peraeonal segmentation not apparent _______3
- 3a. Seven lobes apparent; five lobes somewhat radially arranged

 Heptalobus

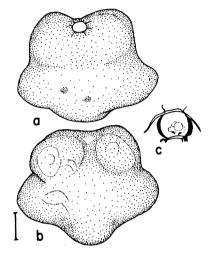


Figure 548. a. Heptalobus paradoxus, oral view. b. Aboral view. c. Detail, oral region.

Fig. 548. Heptalobus paradoxus (Nierstrasz and Brender) (4 mm)

Range: Bering Sea (730 m)

The host shring was Spironto-caris buinguis. The ring structure apparently represents the oral region. No males were found.

3b. Bean-shaped, ocarina-shaped or composed of two subspherical parts

4a. Bean-shaped

Faba

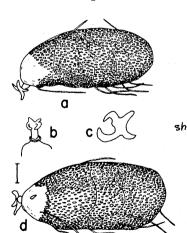


Figure 549. a. Faba setása, lateral view. b. Attachment stalk. c. Detail, attachment stalk. d. Opposite view of a.

Fig. 549. Faba setosa (Nierstrasz and Brender) (8 mm)

Range: Off central California (305 m)

The species was found on the crab Spirontocaris bispinosa. A second species of the genus is found in Hawaii. The male has never been seen. Apparently the species attaches to the host by means of the four pointed anterior (?) processes.

Faba glabra - Hawaii
? Cryptothir balani (Bate 1860)

Fig. 550(a). *Duplorbis ocarina* (Nierstrasz and Brender) (4 mm)

Range: Aleutian Islands (132 m)

The parasite shaped like the musical instrument, the ocarina, is from the branchial cavity of a crab. A cross section [A to B, Fig. 505(a)] of the animal is shown with the small intestine indicated. The large cavity of the ovary is surrounded by ovarian tissue.



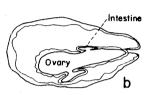


Figure 550(a). a. Duplorbis ocarina. b. Section cut at A-B.

5b. Body composed of two subspherical partsLiriopsis

Fig. 550(b). Liriopsis pygmaea (Rathkei) (5 mm)

Range Puget Sound, Washington (Shallow water)

The species is parasitic on the rhizocephalans *Peltogaster* paguri and *P. gasterella gracilis* which in turn are parasitic on hermit crabs.

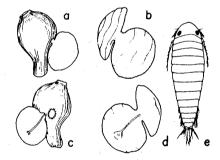


Figure 550(b). a. Liriopsis pygmaea on host, dorsal view. b. Isolated from host, dorsal view. c. Isopod and host, ventral view d. Isolated from host, ventral view. e. Male (After Sars).

DAJIDAE

The body of the female is composed mostly of a mass of eggs that is sometimes symmetrically arranged around a segmented body. The mouth, even in the young female stages, is composed of an attachment sucker. The sucker is not found in mature males, but whether or not it is present in larval males has not been determined. The antennae are severely reduced in size or absent, and only two to five pair of anterior peraeopods are present. The pleo-

pods, if present, are rudimentary, and the uropods are absent or very small if present. The males are much like those of other families, but in some species the cephalon and peraeonal segment I are fused. The pleonal segments are also fused. The species are parasites on species of Mysidacea and Euphausiacea. Four species in four genera are known from North American waters.

KEY TO GENERA AND SPECIES OF DAJIDAE

- 1a. Peraeonal segmentation visible in dorsal view; body flanked laterally by large egg masses 2

Fig. 551. Dajus mysidis (Kroyer) (4 mm)

Range: Laborador and Greenland (5 to 37 mm)

The species is a parasite which attaches to the abdomen of mysid shrimps.

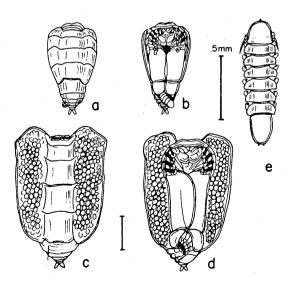


Figure 551. a. Dajus mysidis, dorsal view young female. b. Ventral view. c. Dorsal view, mature female. d. Ventral view. e. Male.

2b. Uropods not distinct; body with somewhat of a symmetry Colophryxus

Fig. 552. Colophryxus novangliae (Richardson)

Range: South of Long Island, New York (1293 m)

This species is the type and only species in the genus. It was found unattached in a fishing trawl net, therefore, the host is not known.

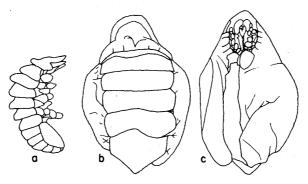


Figure 552. a. Colophryxus novangliae, male. b. Dorsal view, female. c. Ventral view.

Figs. 553 and 554. Prophryxus alascensis Richardson 1909 Range: Off Alaskan panhandle (3658 m)

This species is perhaps parasitic on a schizopod since one was caught with it. The name of the schizopod was not given.

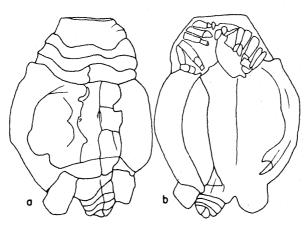
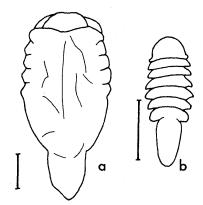


Figure 553. a. Prophryxus alascensis. b. Ventral view.



Holophryxus giandi Rich. 1908
Bering Islands, Alaska

Holophryxus californianus Rich. 1908
Central California

Figure 554. a. Prophryxus alascensis from another host). b. Male.

Fig. 535. Holophryxus alascensis (Richardson) Range: Alaska, near Juneau (269 to 640 m)

The host for this species is an unnamed stomatopod.

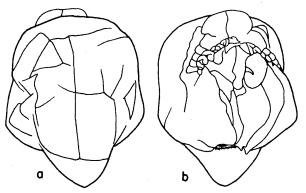


Figure 555. a, Holophryxus alascensis. b. Ventral view.

ENTONISCIDAE

Species of entoniscids are true endoparasites and are found within the body cavity of their decapod hosts. Although at first the specimens appear to be a mass of undifferentiated host tissue, it is interesting to note that all the body tagmata are present and almost all of the segments of each tagmata are represented. It is only after careful study, however, that they can be properly identified. The cephalon is generally composed of two large cephalic lobes and the pleon is long and slender. Large lamellae are present on the peraeonal segments of many species. All appendages are

rudimentary or absent, and only the mandibles are recognizable of the mouth parts. The peraeopods are present as buds or completely absent, and there are five segments plus a pleotelson present and distinct and the segments are sometimes equipped with pleopods that are long and pointed. Gravid females appear to be simply an undifferentiated mass of eggs, and only after careful examination is the body of an isopod recognized. The males are found on the body of the non-gravid female or in the egg mass of the gravid female. Their body is generally wide at the anterior end and tapers to a thin pleon, but it is in almost every other respect like that of other bopyrid males.

There is a small hole in the body wall of the host leading from the branchial cavity to the body cavity (haemocoel) where the parasite is found. In at least one species, the hole regularly pene-

trates from the eye cavity to the body cavity. The parasite is bathed in the body fluid of its host, and this is one of the criteria for calling the isopod a true endoparasite. There are about 34 species known and four are from the Atlantic coast and one from the Pacific coast of North America.

It is not easy to make a useful key to the genera or species of entoniscids so one is not attempted here. When decapods that are parasitized by the isopods are collected, they should be preserved and identified as well as the parasite. The structure of the male also is a useful identifying character. The species of entoniscids are economically important. Five species in four genera are found in North American waters.



Figure 556. Larva of

GENERA OF THE FAMILY ENTONISCID/E

Fig. 557. Cancrion carolinus (Pearse and Walker) (24 mm)

Range: Coast of Carolinas

At Beaufort, North Carolina, the species was found in the crab *Panopeus herbstii*.

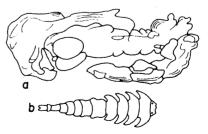
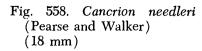


Figure 557, a. Cancrion carolinus, b. Male.



Figure 558. a. Cancrion needleri, b.



Range: Prince Edward Island, Canada

The host crab is Neopanope texana.

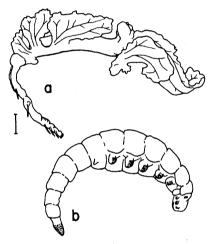


Figure 559. a. Paguritherium altum. b. Male.

Fig. 559. Paguritherium altum (Reinhard) (15 mm)

Range: Woods Hole, Massachusetts

The species is parasitic on the hermit crab Pagurus longicarpus, and it enters the body cavity through the region of the eyestalk, not the branchial cavity as in other entoniscids. In the male the peraeopods are stumps, not jointed peraeopods. The crab host is also host of another bopyrid isopod, Stegophryxus hyptius, a branchial cavity parasite (p. 321)

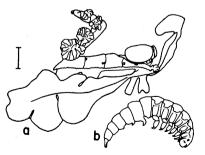


Figure 560. a. Achelion occidentalis. b. Male.

Fig. 560. Achelion occidentalis (Hartnoll) (4.8 mm)

Range: Jamaica

The parasite is found in the spider crabs Microphryus bicornutus and Stenorhynchus seticornis. Fig. 561. Portunion conformis (Muscatine) (14 mm)

Range: Puget Sound, Washington to San Francisco Bay, California

The host is Hemigrapus oregonensis, a decapod. More than one parasite is frequently found on each host.

also H. nudus (see Piltz, 1969)

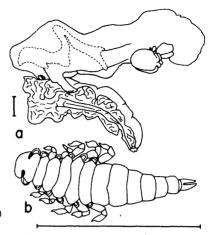


Figure 561. a. Portunion conformis. b.

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An excellent study of the ecology of a single species. More work should be done in this manner.

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Menzies, R. J. 1957. The marine borer family Limnoriidae (Crustacea, Isopoda). Bull, Mar. Sci. Gulf and Caribbean, 7(2):101-200.

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Menzies, R. J. 1962. The isopods of abyssal depths in the Atlantic Ocean. In: Abyssal Crustacea, Vema Res. Ser., 1:79-206.

A study of the deep sea isopods off the coast of North America.

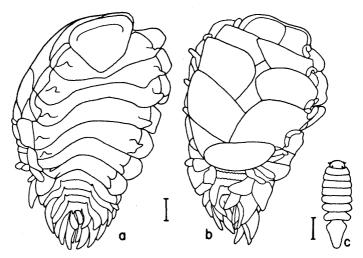


Figure 510. a. Munidion parvum. b. Ventral view. c. Male.

- 10b. Dorsal processes absent near lateral borders of peraeonal segments; uropods uniramus 11

Fig. 511. Cryptione elongata (Hansen) (19 mm)

Range: Galapagos Islands and perhaps west coast of Mexico

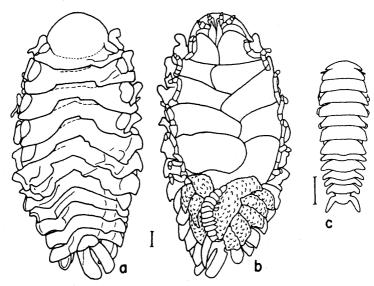


Figure 511. a. Cryptione elongata. b. Ventral view (with male on pleopods). c. Male.

Neither the male nor the female have eyes. The species is found in the branchial cavity of the crabs of the genus Nematocarcinus.

- 11b. Cephalon broadly rounded or with frontal nargin nearly straight and contained almost or completely within peraeonal segment I
- 12a. Some or all pleopods triramus; no papillae dorsolaterally placed on pleonal segment I 13

Fig. 512. Phyllodurus abdominalis (Stimpson) (14 mm)

Range: Puget Sound, Washington to San Francisco Bay, California

The species is parasitic on the erab Upogebia pugettensis.

mud shrimp

Phyllodurus robustus = Pseudione upogebiae

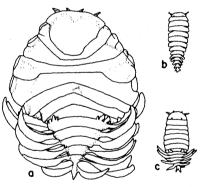


Figure 512. a. Phyllodurus abdominalis. b. Male. c. Juvenile female,

13a. All pleopods triramus Stegophryxus Fig. 513. Stegophryxus hyptius (Thompson) (9.1 mm)

Range: Massachusetts and Rhode Island & Georgia.

The species is a parasite on the abdomen of the hermit crab, *Pegurus longicarpus* which is found on the shore and in shallow water. The crab is also host of another isopod of the family Entoniscidae, *Paguruitherium altum* (p. 346), an endoparasite. Also infests f. annulipes in N. carolina & Georgia; and f. benairensis a f. miamensis in Florida.

Stegophryxus hyphalus: so. Calif. to Baja

see Markham, 1974 for review of genus

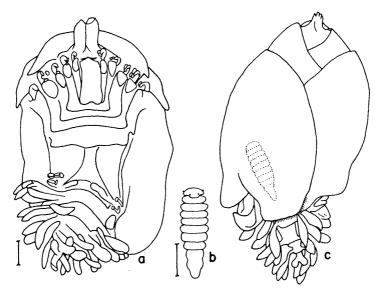


Figure 513. a. Stegophryxus hyptius. b. Male. c. Ventral view.

13b. Only anterior three pleopods triramus Stegias

Figs. 514 and 515. Stegias clibanarii (Richardson) (16 mm)

Range: Bermuda

The host species is Clibanarius tricolor.

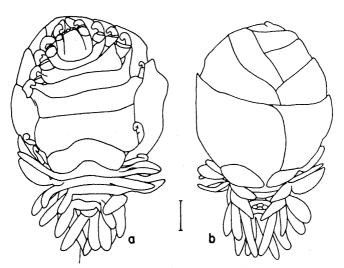
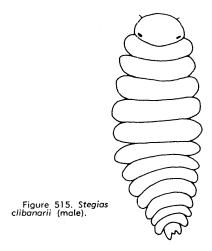


Figure 514. Stegias clibanarii. a. Dorsal view. b. Ventral View.



- 14b. Pleopods short, rudimentary or absent; pleonal lateral edges short; telson or pleotelson never with distinct rami 23
- 15a. Peraeonal segments I to III with secondarily produced posterolateral edges at least on long body side, and secondarily produced posterolateral edges on both edges of segments IV to VII; pleonal segments with lateral margins rounded

Figs. 516 and 517. Argeia pugettensis (Dana) (14 mm) Range: Bering Sea to southern California

At times half a population of shrimp have been recorded to have the parasite in their branchial chamber. Shrimp of the family Cragonidae are the hosts. Argeia pauperata Stimpson (14 mm) from San Francisco Bay and parasitic on Crago franciscorum has been considered to be a separate species although it is very similar to A. pugettensis.

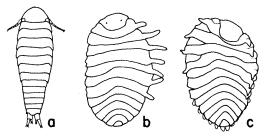


Figure 516. a. Argeia pugettensis, larva. b. Juvenile female d. Young female.

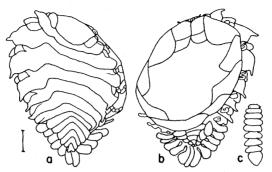


Figure 517. a. Argeia pugettensis (gravid female). b. Ventral view. b. Male.

15b. Not as above _______16

Fig. 518. Parageia ornata (Hansen) (7 mm)

Range: Acapulco, Mexico

The species is parasitic on the shrimp Sclerocrangon procax.

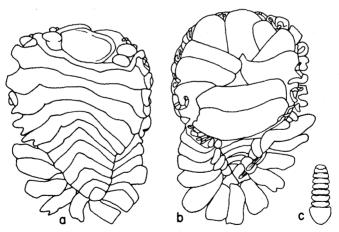


Figure 518. a. Parargeia ornata. b. Ventral view. c. Male.

16b. Pleonal segments with lateral edges moderately long and lateral margins various (for example, square, lobular, secondarily produced, etc.)

Pseudione

GENUS PSEUDIONE

The genus contains species that are parasitic on crabs and shrimp. The species constitute one of the most widespread groups of parasitic isopods, and when they are more closely examined they will probably be separated into several genera. They are most commonly encountered in the branchial cavity of hermit crabs, but they are occasionally found in shrimps.

KEY TO SPECIES OF PSEUDIONE

17a. With eyes; cephalon ovate P. trilobata

Fig. 519. Pseudione trilobata (Nierstrasz and Brender) (3 mm)

Range: Curacao, Dutch West Indies

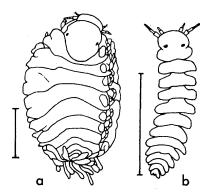


Figure 519 a. Pseudione trilobata. b.

17b. Blind; cephalon not ovate 18a. Pleopods digitate P. upogebiae

Fig. 520. Pseudione upogebiae (Hay) (11 mm)

Range: Beaufort, North Caro-

The species is a parasite of the gill chambers of the shrimp Upogebia affinis. The distinct digitate pleopods distinguish the species from other species in the genus. When more observations are made, the peculiar pleopods might serve as criteria for establishing a new genus.

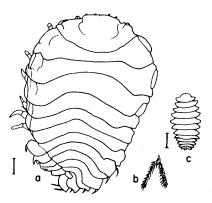


Figure 520. a. Pseudione urogebiae. b. Detail, pleopod. c. Male.

Markham (1974) placed Phyllodorus robustus Peanse (1953) into ir synonomy with Pseudicne UDOasbias

18b. Pleopods not digitate 19
19a. Posterior part of body abruptly narrower than anterior part; pleonal secondary lamellae not distinct P. giardi
Fig. 521. Pseudione giardi (Calman) (12 mm)

Range: Puget Sound, Washington

The species is parasitic on the hermit crab Pagurus ochotensis.

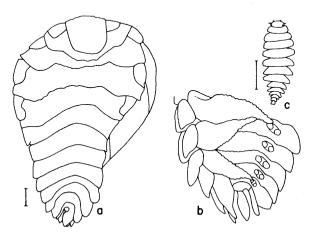


Figure 521. a. Pseudione giardi. b. Ventral view, posterior part. c. Male.

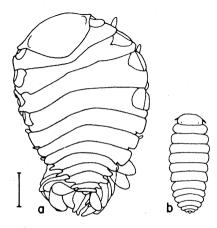


Fig. 522. Pseudione curtata (Richardson) (5.5 mm) Range: Key West, Florida

Only one specimen of the species was taken from the branchial cavity of the crab *Petrolisthes sexspinous*.

Figure 522. a. Pseudione curtata, b. Male

20b.	Pleonal terga not produced or produced into large secondary lamellae 21				
21a.	Pleonal terga not produced into large lamellae 22				
21b.	Pleonal terga produced into large secondary lamellae				
	P. galacanthae				

Fig. 523. Pseudione galacanthae (Hansen) (11 mm)

Range: Gulf of California to Washington State

The species was found in the branchial cavity of the galatheid crab Munnida quadrispina.

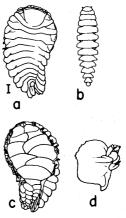


Figure 523. a. Pseudione galacanthae. b. Male. c. Ventral view. d. Oostegite

Fig. 524. Pseudione furcata (Richardson) (11 mm)

Range: Virginia Host unknown.

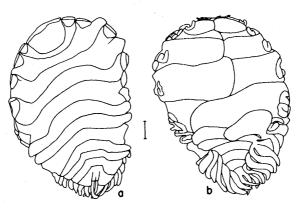


Figure 524. a. Pseudione furcata, b. Ventral view.

Menzies, R. J. and D. Frankenberg 1966. Handbook on the Common Marine Isopod Crustacea of Georgia. Univ. Georgia Press, pp. viii-93.

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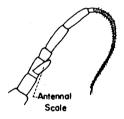


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Aporobopyrus

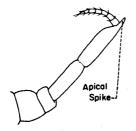


Figure 563. Apical spike (Antennulonis-cus).

muguensis, 315 mouth part, antenna, Appendage: peraeopod or pleopod Appendix masculina: see male pleopod 2 Fig. 567 Apsidoniscus perplexus, 270 Arcturidae, 50 Arcturus baffini, 58 beringanus, 57 caribbaeus, 58 glaber, 57 longispinus, 57 purpureus, 58 Archetype isopod, 7 Argeia pauperata, 323 pugettensis, 323 Arthropoda, 1 Article: segment of flagella of antennae and mandibular palp Asellidae: family of isopods which are found in freshwater habitats (only Asellus sp. is included here as an example, 241) Aselloidea (superfamily), 230 Pleopods, 241 Asellota (suborder), 229 General nomenclature, 230 Antennal one types, 231 Maxillipeds, 233, 250 Peraeopods, 232 Asellus sp., 242 Astacilla caeca, 51 grandulata, 51 lauffi, 51 Austrosignum tillerae, 292

В

Baiting for isopods, 22 Basipod: see basis, 40

Basis (pl. bases): segment closet to body except where coxal segment is present Bathura luna, 98 Bathygnathia curvirostris, 221 Bathygyge grandis, 312 Bathynomus giganteus, 169 Benthic: bottom dwelling animal Bopyrella maginitiei, 332 harmopleon, 332 Bopyridae, 312 Bopyrina abbreviata, 334 cragona, 334 latreuticola, 335 pontoniae, 336 thorii, 335 urocaridis, 336 Bopyrinella antillensis, 333 Bopyriscus calmani, 336 Bopyro choprae, 337 Bopyroides hippolytes, 329 Boss: an enlargment; a large tubercle Braga cichlae, 154 occidentalis, 155 Branchial chamber: gill cavity in decapods (where parasitic isopods are sometimes found); pleopodal cavity in Asellota (230) and Valvifera (48) Caecianiropsis

needleri, 346
Carcinology: study of crustaceans
Carpias
bermudensis, 260
Carpus: peraeopodal segment (5), 32
Cassidinidea
lunifrons, 115
ovalis, 115
Cephalization: process of formation
through evolution of anterior
appendages to sensory and gna-

psammophila, 257

horvathi, 257

branchiata, 102

carolinus, 345

Caecijaera

Calathura

Cancrion

thal from locomotoral ones. Fig. Cephalon: head-anterior most tagma where gnathal and sensory appendages are located Cerathothoa impressa, 155 Chelate: clawed Chelifera: see Tanaidacea. 4 Chiridotea almyra, 61 arenicola, 62 coeca, 62 nigrescens, 62 stenops, 61 tufsi, 61 Cirolana albida, 180 borealis, 182 californiensis, 178 concharum, 181 diminuta, 184 gracilis, 185 harfordi, 183 impressa, 184 joanneae, 179 minuta, 180 obtruncata, 181 parva, 185 polita, 183 sphaeromiformis, 178 Cirolanidae, 168 Cirolanides texensis, 186 Cleantis heathii, 82 occidentalis, 83 planicaudata, 82 Clypeoniscus meinerti, 338, 339 Clypeus: front of cephalon above labrum, 37, 75 Colanthura squamossima, 90 tenuis, 89 Colidotea rostrata, 74 Collecting methods, 21, places, 20 Colophryxus novangliae, 343 Colopisthus parvus, 168 Conilera cylindracea, 174 stygia, 174 Corallanidae, 212 COUPLING HOOKS: structures found on medial edge of maxilliped (p. 38) and pleopods (Fig. 564) which apparently fastens them together

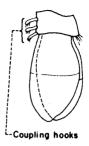


Figure 564. Pleopod with coupling hooks on basis.

Coxa: peraeopodal segment, (1), 32 Coxal plate: see epimere Creaseriella anops, 187 Cryptione elongata, 320 Cryptoniscan larvae or stage, 309 Cryptoniscidae, 337 Culture (or rearing), 23 Cumacea, 4 Cvathura burbanki, 107 carinata, 107 cruis, 105 Curassavica, 104 munda, 105 polita, 106 specus, 104 Cymodoce faxoni, 127 japonica, 127 Cymothoa caraibica, 160 excisa, 346, 160 exigua, 161 oestrum, 161 Cymothoidea, 145 Mouth parts, 146

Dactylus: peraeopodal segment, (7)
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Dendrotion
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birsteini, 279 lobiceps, 280 plebjum, 280 symmetrica, 279 tenuimanum, 281 Desmosomatidae, 278 Development, 24 Digitate: finger-like; branched Diplophryxus synalphei, 314 Discerceis granulosa, 126 linguicauda, 126 Distribution, 14 (Map, Fig. 13) Dorsal pit, 86 Dorsolateral keel, groove, 86 Dorsum: upper surface of body or of tagmata of body Duplorbis ocarina, 341 Dynamene dilatata, 122 Dynamenella angulata, 125 benedicti, 125 glabra, 122 moorei, 124 perforata, 124 sheareri, 123 Dynameniscus carinatus, 118 Dynamenopsis dianae, 123

Echinothambema ophiuroides, 248 Echinothambematidae, 247 Ecology, general, 9 Economic importance of isopods, 9 Limnoria, 138 Sphaeroma destructor, 128 Sphaeroma pentadon, 128 Edanthura linearis: this species from California was not described in sufficient detail to be recognized again, hence, it is a nomen nundem Edotea acuta, 81 montosa, 80 sublittoralis, 81 triloba, 81 Eisothistos atlanticus, 97 Endopod: inner branch of appendage Entoniscidae, 344 Epicaridea (suborder), 309 General nomenclature, 309 Epicaridian larva, 309

EPIMERE: modified coxal segment of pleopod sometimes forming extension of peraeon, Fig. 565

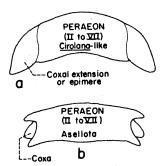


Figure 565. Coxa and coxal extensions (epimeres) on peraeonal segments, a. General Flabellifera. b. Some Asellota.

Erichsonella attenuata, 84 crenulata, 85 filiformis, 84 floridana, 84 pseudoculata, 85 Erygne rissoi, 315 Eucarida, 5 Eurycope californiensis, 305 complanata, 305 cornuta, 304 mutica, 304 Eurycopidae, 302 Eurydice branchuropus, 173 caudata, 173 convexa, 172 littoralis, 171 piperata, 172 spinigera, 171 Eusymmerus antennatus, 83 Excirolana chiltoni, 175 kincaidi, 175 linguifrons, 174 mayana, 174 vancouverensis, 175 Excorollana antilliensis, 211 berbicensis, 206 fissicauda, 208 kathae, 209 mexicana, 209

oculata, 207 quadricornis, 210 rapax, 211 sexticornis, 210 subtilis, 206 tricornis, 210 truncata, 208 warmingii, 207 Excorallanidae, 205
Exopod: outer branch of appendage Exosphaeroma amplicauda, 131 antillense, 134 crenulatum, 132 diminutum, 136 dugesi, 134 inornata, 133 media, 133 octoneum, 135 papillae, 132 rhomburum, 135 thermophilum, 135

F

Faba
setosa, 340
Flabellifera (suborder), 109
General nomenclature, 110
Maxillipeds, 111, 177
Flagellum (pl. flagella): distal small
segments of antennae, 35
Fossorial: modified for digging or
burrowing
Frontal lamina: protuberance ventral on cephalon usually between
antennae, Fig. 566

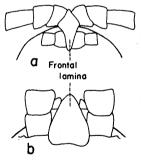


Figure 566. Frontal lamina between antennae (ventral view). a. Eurydice-like. b. Antennuloniscus.

Frontal plate: see frontal lamina

G

General Nomenclature, 7 Geniculate: bent like a knee

Gnathal: having to do with jaw or mouth Gnathia cerina, 226 clementensis, 223 coronadoensis, 222 crenulatifrons, 224 cristata, 225 elongata, 228 hirsuta, 225 multispinis, 224 productatridens, 228 serrata, 222 steveni, 227 triospathiona, 223 tridens, 229 trilobata, 227 Gnathiidae, 217 Gnathiidea (suborder), 217 General nomenclature, 218 Female, 25, 226 Juvenile, 220 Gnathopod: peraeopod modified for eating or food gathering (usually peraeopod I) Gnorimosphaeroma insularis, 130 lutea, 130 nobeli, 131 oregonensis, 129 Grapsicepon choprae, 319 edwardii, 318 Gravid: with eggs or offspring in marsupium

ш

Habitats, 16, 20 (Figs 14, 15, 16) Haliophasma geminata, 103 Haplomesus quadrispinosus, 247 tropicalis, 247 Haplomunna caeca, 242 Haploniscidae, 270 Haploniscus excisus, 272 retrospinis, 273 tropicalis, 273 unicornis, 272 Haptolana trichostoma, 186 Head: see cephalon Hemirathus abdominalis, 313 schmitti, 314 subcaudalis, 313

Heptalobus

paradoxus, 340

Heteromesus bifurcatus, 246 granulatus, 246 spinescens, 245 Holophryxus	Ione brevicauda, 316 cornuta, 316 thompsoni, 316 Incisor process: cutting structure of
alascensis, 344	mandible, 37
Horolanthura	Incubatory lamellae: see oostegites
irpex, 93	Incubatory pouch: see marsupium
Hydroniscus abyssi, 274	Indurate: hard, stiff or thick
quadrifrons, 274	Indusa carinata, 159
Hypopharynx: sensory structure just	Inner antenna: antenna one
behind mandibles, 38	Irona: see Mothocya, 162
Hyssura	Ischium: peraeonal segment (3), 32
producta, 93	Ischnomesidae, 243
profunda, 93	Ischnomesus
<u> </u>	armatus, 383
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caribbica, 256	j
lobata, 255	Jaera
vemae, 255	marina, 269
Ianiropsis	wakishiana, 269
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epilittoralis, 262	Jaeropsis Jaeropsis
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magnocola, 263	coralicola, 285
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Idotea (Idothea, old spelling)	tricornis, 259
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metallica, 78	alascensis, 268 erostrata, 268 holmsei, 268
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phosphorea, 76	rajata, 265
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hirticeps, 302	Isopod suborders, 47
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libbeyi, 252	Flagellifera, 111 Gnathiidea, 221
speciosa, 251	Microcerberidea, 214
spinosa, 252	Valvifera, 49
•	•

L

Labeling of specimens, 30 posterior lip-see Labium: hypopharynx Labrum: anterior lip below clypeus, 37, 75 Lacinia mobilis: articulate structure near incisor process of mandible, Lappet, lateral: produced extensions of peraeonal segment (or cephalon, or pleon)-e.g., Asellota Largest isopod, 169 Leidva bimini, 318 distorta, 317 Leptanthura tenuis, 96 thori, 96 Limnoria algarum, 140 lingnorum, 145 pfefferi, 144 platycaudata, 143 quadripunctata, 143 saseboensis, 144 simulata, 142 tripunctata, 142 Limnoridae, 138 Liriopsis pygmaea, 341 Lironeca (Livoneca, old spelling) californica, 166 convexa, 167 ovalis, 164 panamaensis, 167 redmannii, 163 reniformis, 167 symmetrica, 165 tenuistylis, 164 texana, 166 vulgaris, 165

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Macrostylidae, 275
Macrostylis
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caribbicus, 278
galatheae, 275
minutus, 276
setifer, 276
truncatex, 275
vemae, 277
Malacostraca, 3
Male pleopod 2 Fig. 567
MALE STYLET: modified endopod
of pleopod 2 male Fig. 567

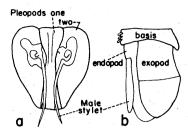


Figure 567. Male sexual pleopods 2 a. Asellota (Paraselloidea). b. General,

Mandibular palp: small appendage of one, two or three articles coming from mandibule—absent oniscoids and most valviferans

MANUS: hand, usually refering to propodus and dactylus of peraeopod I, Fig. 568



Figure 568. Peraeopod I, Stenetrium-like.

Marsupium: brood pouch of female where offspring develop, 24 Maxilla one: see mouth parts Maxilla two: see mouth parts Maxilliped: General nomenclature, 38 Anthuridea, 115 Asellota, 250, 333 Epicaridea, 309 Flabellifera, 111 Valvifera, 49 Maxillula: maxilla one-see parts Meinertia deplanata, 157 gaudichaudii, 157 gilberti, 158 transversa, 156 Membraneous pleopods: fleshy thin pleopods found in some

sphaeromatids, (4 and 5) Fig. Merus: pleopodal segment (4), 32 Mesanthura occidentalis, 109 pulchra, 109 Mesidotea: see Saduria, 59 Mesosignidae, 298 Mesosignum kohleri, 298 usheri, 299 Microarcturus tannerensis, 56 Microcerberidae, 214 Microcerberidea (suborder), 214 Microcerberus abbotti, 216 littoralis, 217 mexicanus, 215 mirabilis, 216 renaudi, 217 Microniscus larva, 309 Microproctus caecus, 254 Molar process: process variously modified arising from mandible, 37 Morphology, general, 31 Mothocya nana, 147 MOUTH PARTS General, 35 and Fig. 569 Asellota, 231 Epicaridea, 309 Flabellifera, 32, 146 Gnathiidea, 348 Valvifera, 49



Figure 569. Mouth parts, 1-6 anterior to posterior. 1. Labrum. 2. Mandible. 3. Hypopharynx. 4. Maxilla one (Maxillula). 5. Maxilla two. 6. Maxilliped,

Munidion parvum, 319 Munna chromatocephala, 297 fabricii, 296

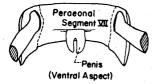
fernaldi. 294 halei, 294 kroyeri, 295 magnifica, 296 reynoldsi, 295 spinifrons, 293 stephenseni, 297 truncata, 293 ubiquita, 298 Munnidae, 289 Munnogonium erratum, 289 waldronensis, 290 Munnopsidae, 288 Munnopsis longiremis, 288 typica, 288

Nannoniscidae, 281 Nannonisconus latipleonus, 282 Nannoniscus arcticus, 283 camayae, 283 oblongus, 282 primitivus, 283 Natatory: swimming structures especially peraeopods, Fig. 31 Neastacilla californica, 53 Neoanthura coeca, 90 Neomunna: see Munna Nerocila acuminata, 152 californica, 151 munda, 151 lanceolata, 150 Non-obligate parasite: parasite which does not depend upon its host for completion of its life cycle

Ocellus (pl. ocelli): a unit of com-

pound eye; one to many found if present Olencira praegustator, 152 Omnivores, 9 Oniscoidea (suborder): isopods which live on land (only mentioned here-p. 2 and Fig. 9) Oostegite: incubatory lamella-structure arising from peraeopod which covers marsupium, 24 Operculate: door-like-covering structure of pleopods or valves Outer antenna: antenna two Ovigerous: see gravid

Paguritherium altum, 346 Palaegyge meeki, 328 Pananthura formosa, 97 Pancarida, 5 Paracerceis caudata, 119 cordata, 120 gilliana, 121 sculpta, 119 tomentosa, 119 Paradynamene benjamensis, 121 Paralimnoria andrewsi, 139 Paranthura antillensis, 95 elegans, 94 infundibulata, 95 Parargeia ornata, 324 Paraselloidea (superfamily), 230 Pleopods, Fig. 366 Parasitic species, Fig. 12 of fish, 160, 190 of other crustaceans, 309 Peduncle: segment or segments which are closest to body; first 5 (or 6) segments of antenna two; first 3 of antenna one Pelagic: see planktonic PENIS: sexual structure located on peraeonal segment VII; sometimes of taxonomic value, Fig.



male stylet—Fig. 567)

570 (not to be confused with

Figure 570, Penis on peraeonal segment VII (ventral view). type 1-5.

Pentidotea aculeata, 73 montereyensis, 72 resecta, 71 schmittii, 72 stenops, 74 wosnesenskii, 73 Peracarida, 4

Peraeon: thorax-body middle or tagma where locomotoral appendages are located, 7 Peraeopods: legs or locomotoral structures, 32 Phreatoicidea (suborder): isopods found mostly in Southern Hemisphere (only mentioned herep. 43 and Fig. 9) Phryxus: see Hemiarthus Phycolimnoria algarum, 140 Phyllodurus abdominalis, 321 Planktonic: living in open water Pleon: abdomen-posterior most tagma where sexual, respiratory and sometimes natatory appendages are located, 7 PLEOPODS: appendages of pleonal

segments except uropods, Fig.

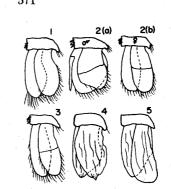


Figure 571. Pleopods - Sphaeromatid

Pleotelson: fused telson and one or more posterior pleonal segments, Fig. 572

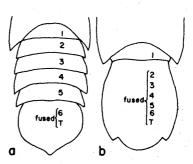


Figure 572. Fusion of pleonal segments. "T" equals telson. a. Gene al. b. Aselluta.

Pleurogonium	dumerilii, 204
californiense, 291	insularis, 202
inerme, 291	laticauda, 199
rubicundum, 290	maculata, 202
spinosissmum, 291	oculata, 200
Pleuroprion	propodialis, 204
intermedium, 52	signata, 201 tridens, 198
murdochi, 52	tridens, 198
Portunion	tuberculosa, 203
conformis, 347	Rock-boring isopod, 129
Prehensile: modified for grasping—	Ronalea: see Erichsonella, 85
e.g., peraeopods	Rostrum: medial cephalic anterior
Preservation of isopods, 21, 29	pointing projection or process
Probopyrus	S
alphei, 329	Saduria
bithynis, 331	
floridensis, 331	entomon, 59 sabini, 59
ovitormis, 330	sibirica, 58
panamensis, 331 pandalicola, 330	Scale of drawings, 45
Process: projection from another	Schizopoda: taxon which included
structure	Mysidacea and Euphausiacea
Propodus: peraeopodal segment (6),	Serolidae, 136
32	Serolis
Produced: drawn out or extended	cardinata, 137
Prophryxus	mgravi. 137
alascensis, 343	vemae, 136
Protogynous: animal which is first	Seta (pl. setae): hair or spine on
female then becomes male-e.g.,	isopod (no true setae are ever
anthurids	found on arthropods, but the
Psammophil: animal which lives be-	term is in general use)
tween grains of sand on beach	Skuphonura
or in cave—e.g., Microcerberus,	laticeps, 91
214, Caecianiropsis, 257	Specirolana
Pseudione	bolivari, 176
curtata, 326	pelaezi, 176
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trilobata, 325	quadridentatum, 128
urogebiae, 325	Sphaeromatidae, 114
Pterisopodus: see Nerocila, 152	Spiniform: type of molar process in
Ptilanthura	Jaeropsis, Fig. 37e, and p. 286
tenuis, 108	Squama: antennal scale, 35
tricarina, 108	Statocyst, 91
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R Paris A	Stegophryxus
	hyptius, 321
Ramus: branch-usually endopod or	Stenetriidae, 238
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magnispinis, 308 snanoi, 307 truncata, 306 vemae, 308 Subchelate: modified claw, 5, 32 Subrostral process: see frontal lamina Sympod: basis of uropod, 40 Syncarida, 5 Syneurycope hanseni, 304 parallela, 303 Synidotea angulata, 68 bicuspida, 63 calcarea, 68 consolidata, 66 erosa, 66 harfordi, 67 laevis, 70 laticauda, 67 magnifica, 69 marmorata, 64 muricata, 69 nebulosa, 67 nodulosa, 70 pallida, 65 pettiboneae, 65 picta, 70 ritteri, 64 Synsynella deformans, 333 Syscenus infelix, 97

Tagma (pl. tagmata): functional group of body segments—cephalon, peraeon and pleon, 7
Tanaidacea, 4, 5
Taxon (pl. taxa): any category of classification—e.g., species, genus, suborder, class, etc.
Tecticeps alascensis, 116

convexus, 116 pugettensis, 117 Telotha henselii, 162 Telson: posterior or last body segment-see also pleotelson Tergum (pl. terga): upper part of peraeon Thorax: see peraeon Tridentella virginiana, 213 Trilobite, 2 Troglocirolana cubensis, 188 Tubercles: small bumps Tubiferous: isopod from tubular burrow-e.g., anthurid Type-species: species, usually first described, on which genus is defined

U

Unguis: claw
Uromunna: see Munna
Uropod, 40
Exopod—outer branch
Endopod—inner branch

٧

Valve: uropod covering branchial cavity of valviferans, 48 Valvifera (suborder), 48 General nomenclature, 48 Maxillipeds, 49

W

Wood-boring isopod, 128, 138

X

Xananthura brevitelson, 92 masculinum masculina
mobilis mobiles

frontal
lawina (epistome)

clypeus
labrum

Tsopods with

first coxal plate free appendix mesculinum

mothered to somite) attached distally on endopod

Bathyrromus

Plakarthrium (mediodistally on endopod)

Anuropus Plakarthrium

rlakarthrium some asellotes 4

ornquar

penis

Some epicarids
Some phreatoicids

ii eatoicios

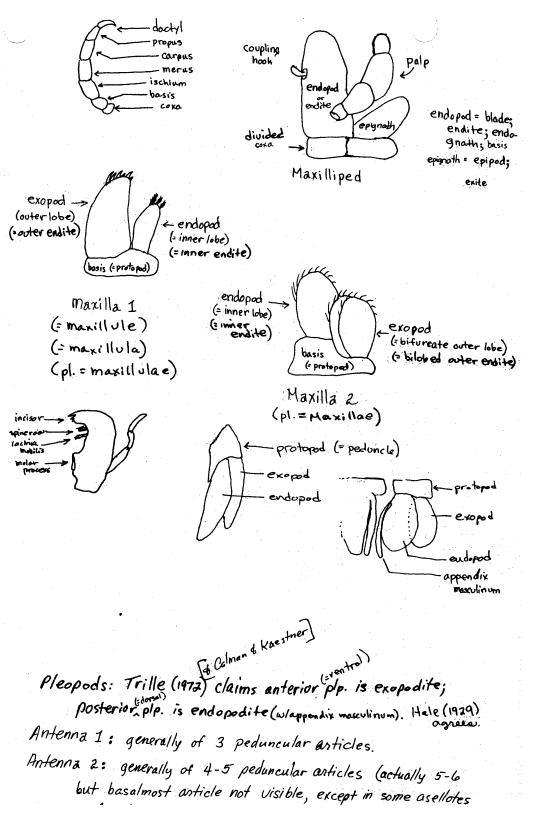
serolidae Cerceis, Bracerceis, Geocerceis

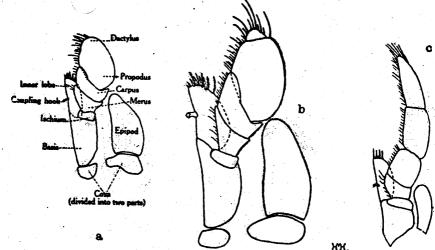
of percon, perconited percopod corresponds with tanail

workers (Torbin Wolff, L.

Gardiner, J. Sieg, R. winn)

<u>Cleantioides</u> (medially on endopod)





Fro. 316.—Maxilliped of (a) Euidotea peronii, (b) Synischia levidensis and (c) Crabyzos longicaudatus (x 16-40).

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